

Fig. 34. Glutathione-S-transferase activity in *Lamellidens corrianus* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.

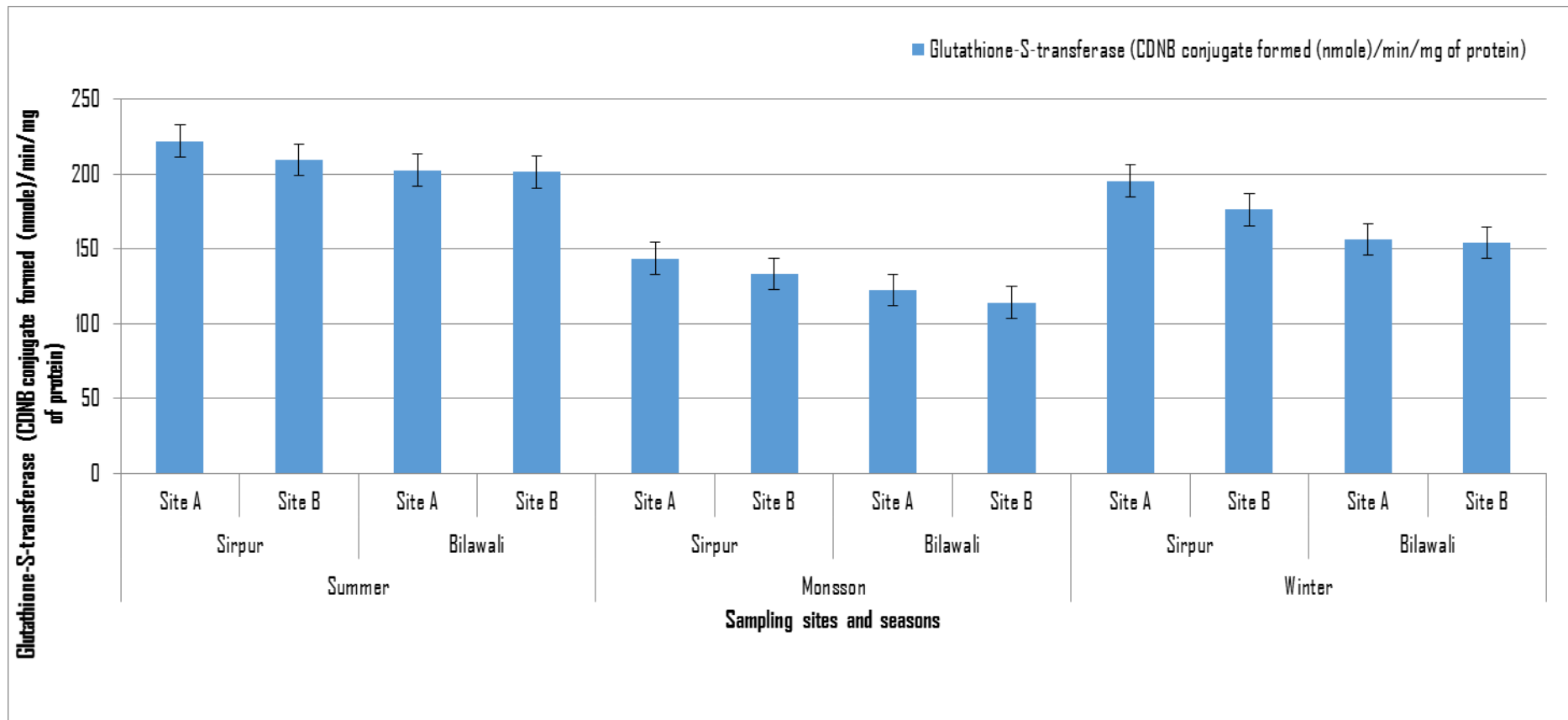
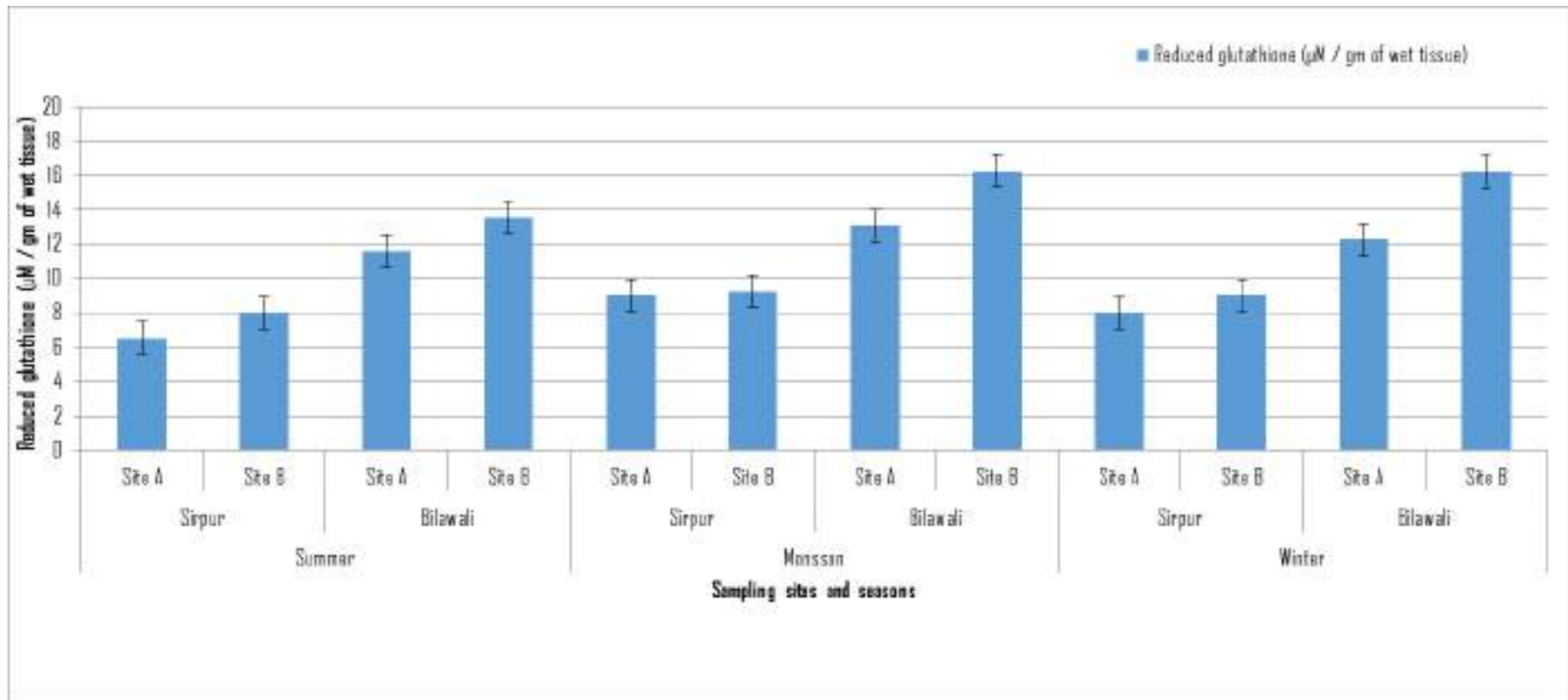


Fig. 35. Reduced glutathione activity in *Lamellidens corrianus* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.



13.58±0.21 in summer; 13.09±0.13 and 16.28±0.11 in monsoon; 12.29±0.12 and 16.24±0.17 in winter seasons respectively.

#### **3.4.1.4 Superoxide dismutase:**

Superoxide dismutase with standard error per mg protein in collected *Lamellidens corrianus* bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 7. Superoxide dismutase with standard deviation per mg protein in *Lamellidens corrianus* bivalve samples collected from Sirpur Lake site A and site B were 119.84±1.29 and 125.64±1.39 in summer; 149.84±1.92 and 169.24±2.09 in monsoon; 128.36±1.28 and 151.31±1.34 in winter seasons whereas, Bilawali Tank site A and site B were reported 135.47±2.58 and 143.57±1.97 in summer; 190.18±2.16 and 201.26±2.84 in monsoon; 149.75±1.19 and 160.75±1.17 in winter seasons respectively.

#### **3.4.1.5 Catalase**

Catalase with standard error per mg protein in collected *Lamellidens corrianus* bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 7. Catalase with standard deviation per mg protein in *Lamellidens corrianus* bivalve samples collected from Sirpur Lake site A and site B were 92.35±1.14 and 93.27±1.18 in summer; 131.06±1.84 and 132.56±1.79 in monsoon;

Fig. 36. Superoxide dismutase activity in *Lamellidens corrianus* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.

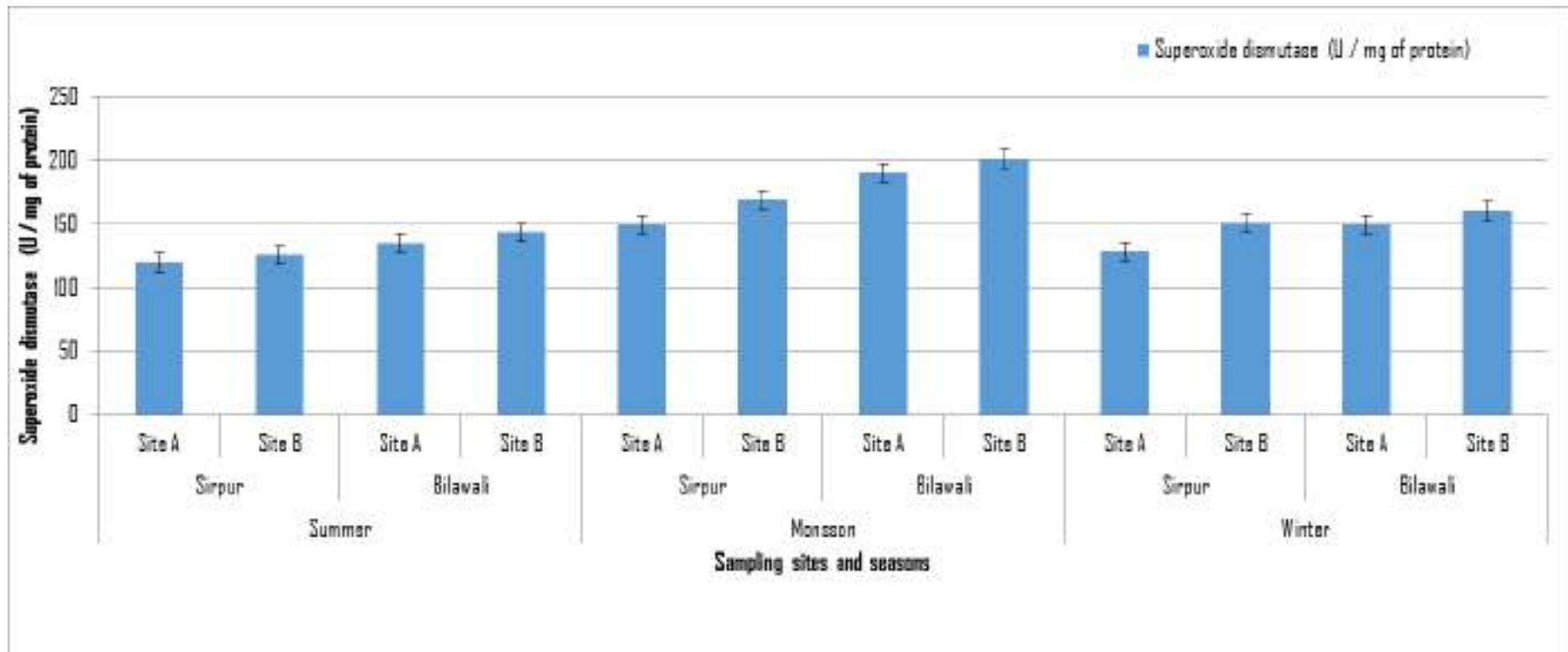
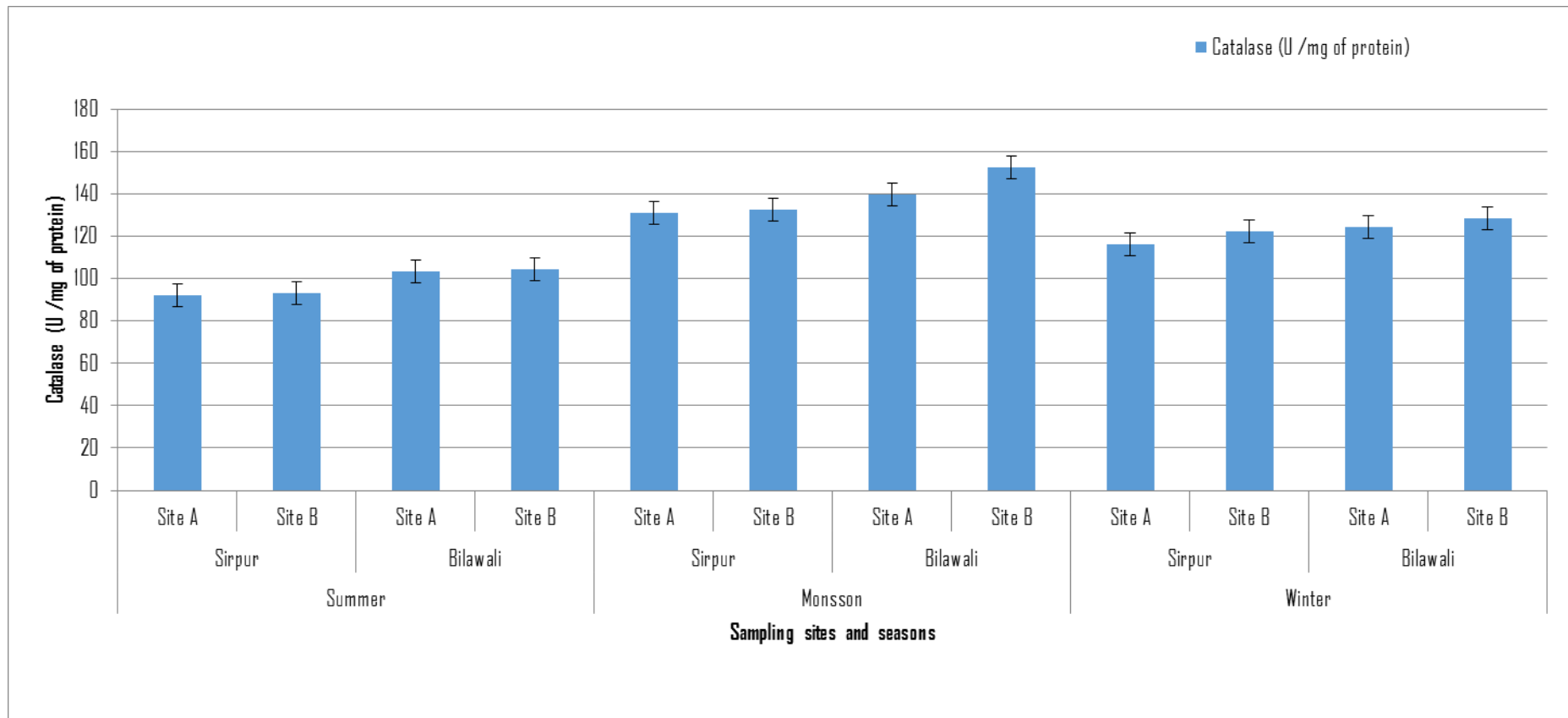


Fig. 37. Catalase activity in *Lamellidens corrianus* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.

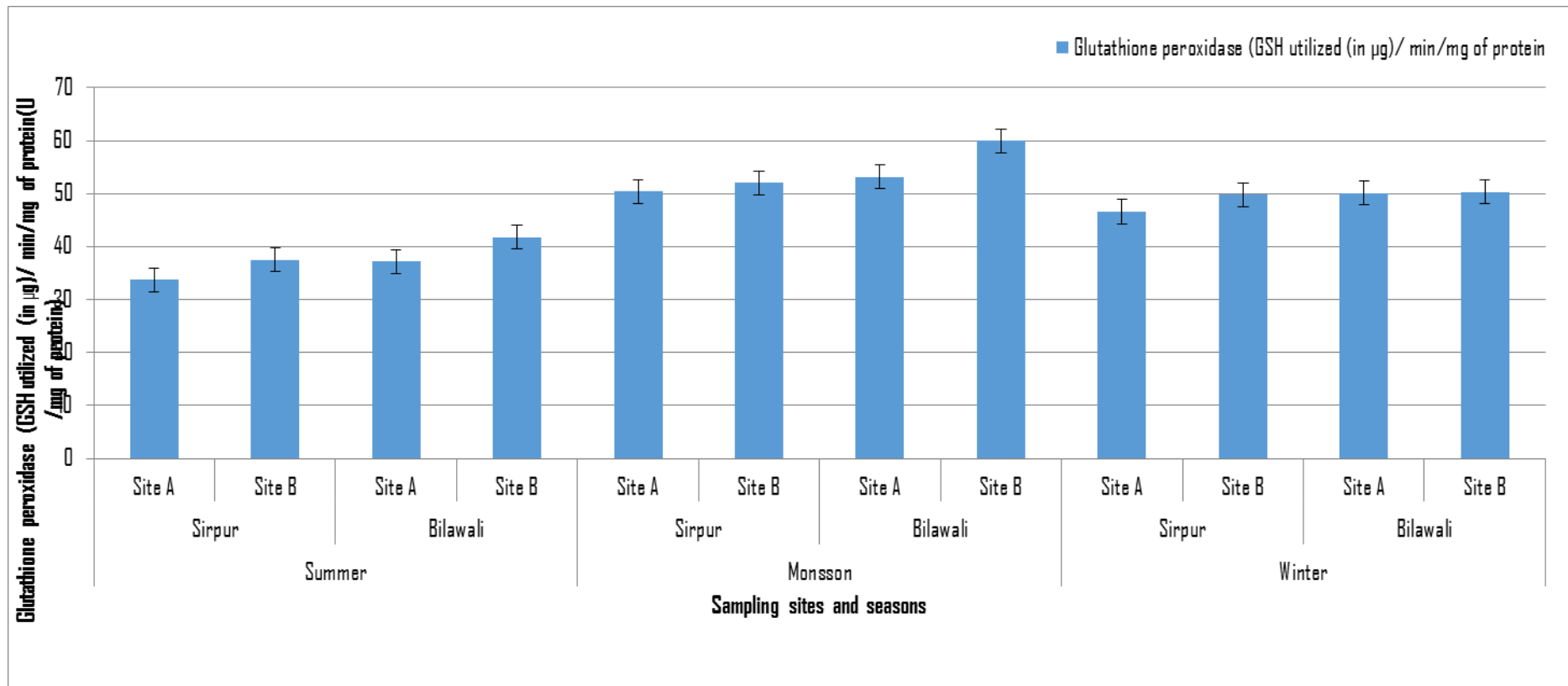


116.24±1.16 and 122.58±1.29 in winter seasons whereas, Bilawali Tank site A and site B were reported 103.52±2.13 and 104.26±1.96 in summer; 139.86±1.29 and 152.45±1.09 in monsoon; 124.51±1.05 and 128.36±1.76 in winter seasons respectively.

#### **3.4.1.6 Glutathione peroxidase**

Superoxide dismutase with standard error per mg protein in collected *Lamellidens corrianus* bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 7. Superoxide dismutase with standard deviation per mg protein in *Lamellidens corrianus* bivalve samples collected from Sirpur Lake site A and site B were 33.75±0.26 and 37.45±0.23 in summer; 50.42±0.17 and 52.01±0.14 in monsoon; 46.59±0.11 and 49.78±0.11 in winter seasons whereas, Bilawali Tank site A and site B were reported 37.19±0.22 and 41.79±0.16 in summer; 53.18±0.11 and 60.01±0.14 in monsoon; 50.14±0.09 and 50.28±0.08 in winter seasons respectively.

Fig. 38. Glutathione peroxidase activity in *Lamellidens corrianus* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.



### 3.4.2 *Lamellidens marginali*

Table 8 presents the mean with standard deviation of collected observations of oxidative biomarkers in *Lamellidens marginali* samples.



Table 8. Seasonal variations of oxidative stress in *Lamellidens marginali* collected from Sirpur and Bilawali Tanks of Indore.

Parameters	Summer				Monson				Winter			
	Sirpur		Bilawali		Sirpur		Bilawali		Sirpur		Bilawali	
	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B
<b>Lipid Peroxidation</b>	216.59± 4.51	196.28± 4.44	191.26± 4.19	179.94± 4.63	143.24± 4.72	126.59± 3.87	121.05± 3.88	118.42± 3.09	189.64± 4.92	175.96± 3.51	167.26± 4.01	161.27± 3.12
<b>Glutathione-S-transferase</b>	249.56± 4.63	201.57± 4.06	213.25± 4.02	201.58± 4.17	163.28± 4.98	124.23± 3.67	134.26± 4.01	109.25± 3.11	199.24± 5.02	171.68± 3.45	172.36± 4.03	156.34± 3.09
<b>Reduced glutathione</b>	8.69±0.0 7	9.02±0.0 6	15.01±0. 09	15.12±0. 10	10.24±0. 11	9.94±0.1 1	16.02±0. 09	17.23±0. 10	9.03±0.0 8	9.05±0.0 7	15.46±0. 12	16.25±0. 09
<b>Superoxide dismutase</b>	121.45± 1.86	123.51± 1.68	138.25± 1.28	149.56± 1.15	165.84± 1.72	178.42± 1.96	194.58± 2.06	194.53± 1.92	128.37± 1.11	149.68± 1.94	146.27± 2.31	155.24± 1.65
<b>Catalase</b>	91.2±1.2 3	99.27±1. 22	98.57±1. 14	108.69± 1.10	135.26± 1.39	141.26± 1.78	153.67± 0.09	158.69± 1.49	119.26± 1.09	128.67± 1.78	120.32± 1.98	131.27± 1.53
<b>Glutathione peroxidase</b>	33.01±0. 13	38.46±0. 12	40.41±0. 11	45.62±0. 08	46.27±0. 12	48.36±0. 13	49.15±0. 14	54.27±0. 14	37.36±0. 05	42.26±0. 09	44.12±0. 10	48.56±0. 10

### 3.4.2.1 Lipid peroxidation

Lipid peroxidation with standard error per mg protein in collected *Lamellidens marginali* bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 8. Lipid peroxidation with standard deviation per mg protein in *Lamellidens marginali* bivalve samples collected from Sirpur Lake site A and site B were  $216.59 \pm 4.51$  and  $196.28 \pm 4.44$  in summer;  $143.24 \pm 4.72$  and  $126.59 \pm 3.87$  in monsoon;  $189.64 \pm 4.92$  and  $175.96 \pm 3.51$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $191.26 \pm 4.19$  and  $179.94 \pm 4.63$  in summer;  $121.05 \pm 3.88$  and  $118.42 \pm 3.09$  in monsoon;  $167.26 \pm 4.01$  and  $161.27 \pm 3.12$  in winter seasons.

### 3.4.2.2 Glutathione-S-transferase

Glutathione-S-transferase with standard error per mg protein in collected *Lamellidens marginali* bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 8. Glutathione-S-transferase with standard deviation per mg protein in *Lamellidens marginali* bivalve samples collected from Sirpur Lake site A and site B were  $249.56 \pm 4.63$  and  $201.57 \pm 4.06$  in summer;

Fig. 39. Lipid Peroxidation activity in *Lamellidens marginali* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.

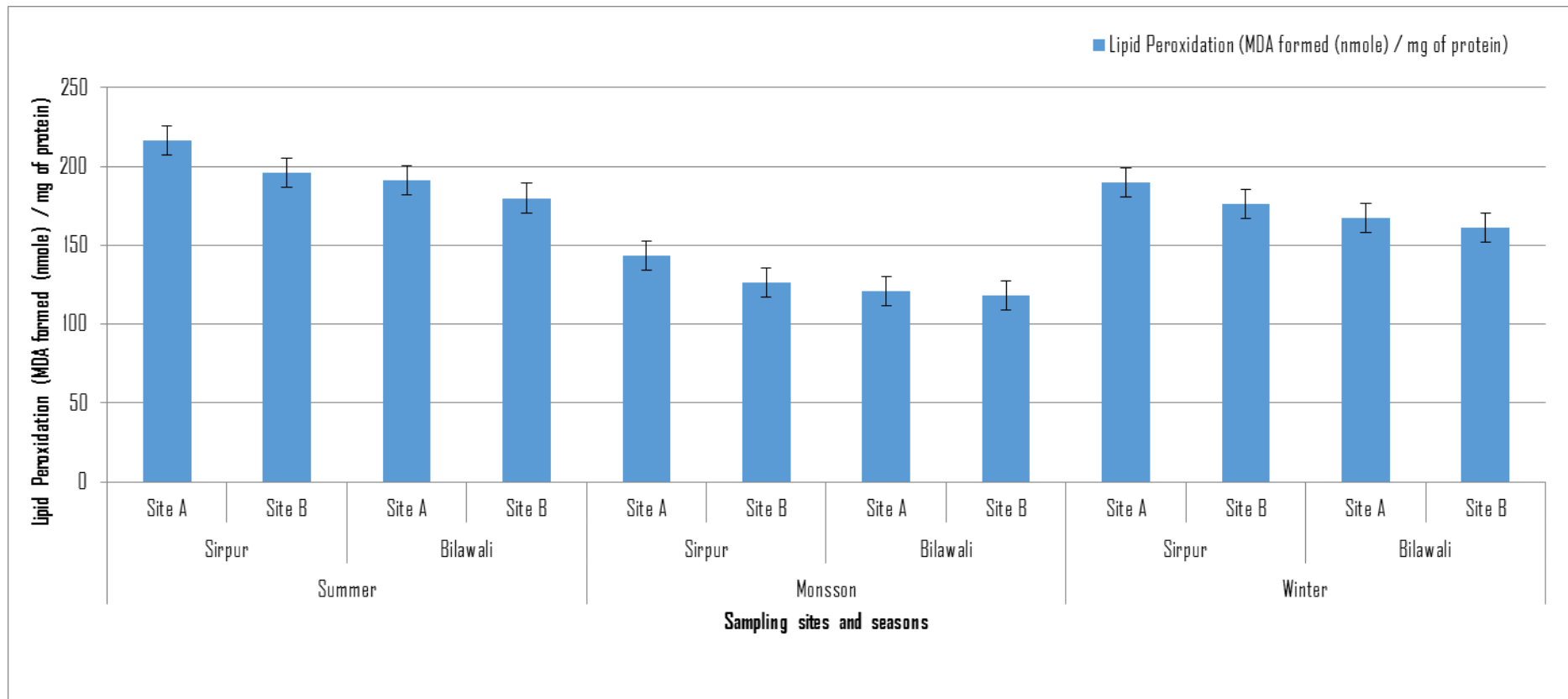
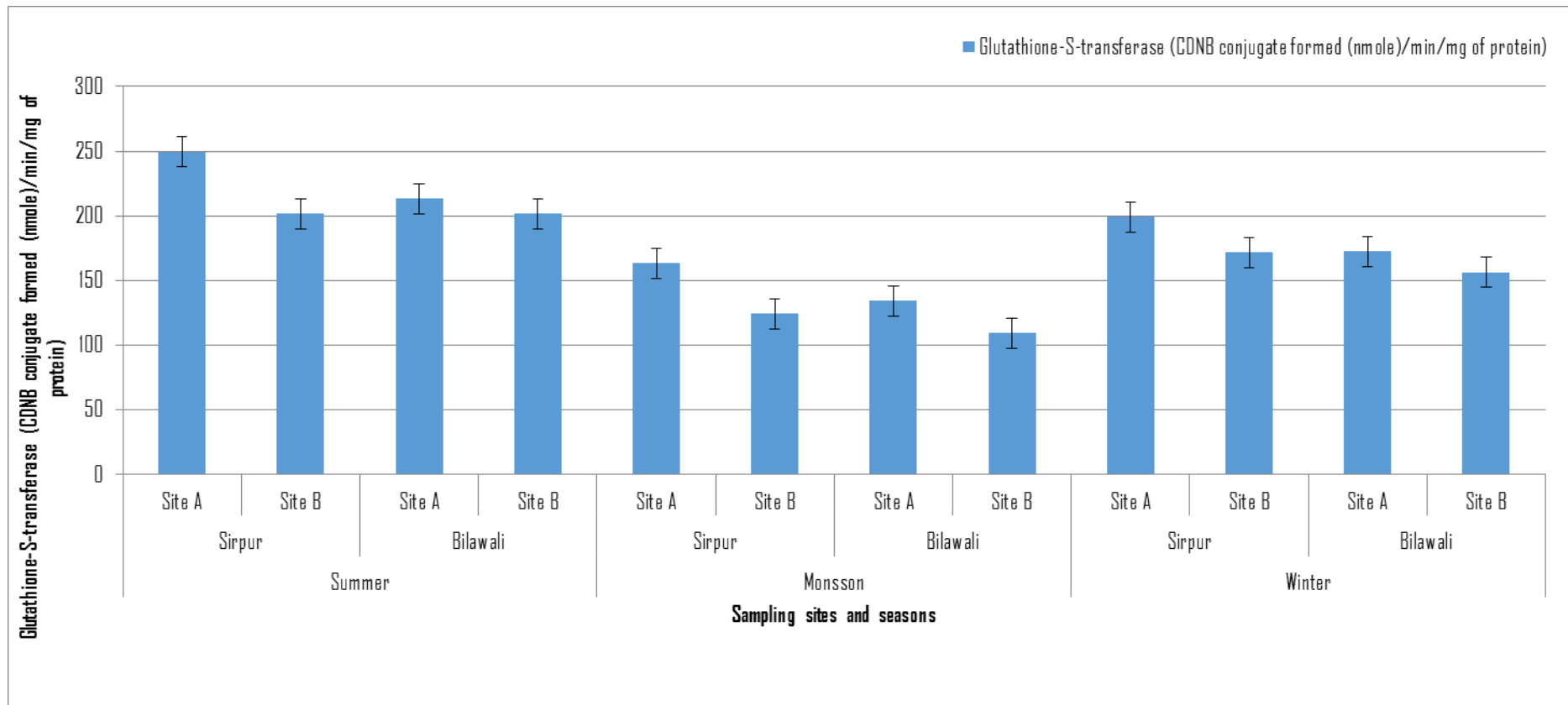


Fig. 40. Glutathione-S-transferase activity in *Lamellidens marginali* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.



163.28±4.98 and 124.23±3.67 in monsoon; 199.24±5.02 and 171.68±3.45 in winter seasons whereas, Bilawali Tank site A and site B were reported 213.25±4.02 and 201.58±4.17 in summer; 134.26±4.01 and 109.25±3.11 in monsoon; 172.36±4.03 and 156.34±3.09 in winter seasons respectively.

### **3.4.2.3 Reduced glutathione**

Reduced glutathione with standard error per mg protein in collected *Lamellidens marginali* bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 8. Reduced glutathione with standard deviation per mg protein in *Lamellidens marginali* bivalve samples collected from Sirpur Lake site A and site B were 8.69±0.07 and 9.02±0.06 in summer; 10.24±0.11 and 9.94±0.11 in monsoon; 9.03±0.08 and 9.05±0.07 in winter seasons whereas, Bilawali Tank site A and site B were reported 15.01±0.09 and 15.12±0.10 in summer; 16.02±0.09 and 17.23±0.10 in monsoon; 15.46±0.12 and 16.25±0.09 in winter seasons respectively.

### **3.4.2.4 Superoxide dismutase:**

Superoxide dismutase with standard error per mg protein in collected *Lamellidens marginali* bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 8. Superoxide dismutase with standard deviation per mg protein in *Lamellidens*

Fig. 41. Reduced glutathione activity in *Lamellidens marginali* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.

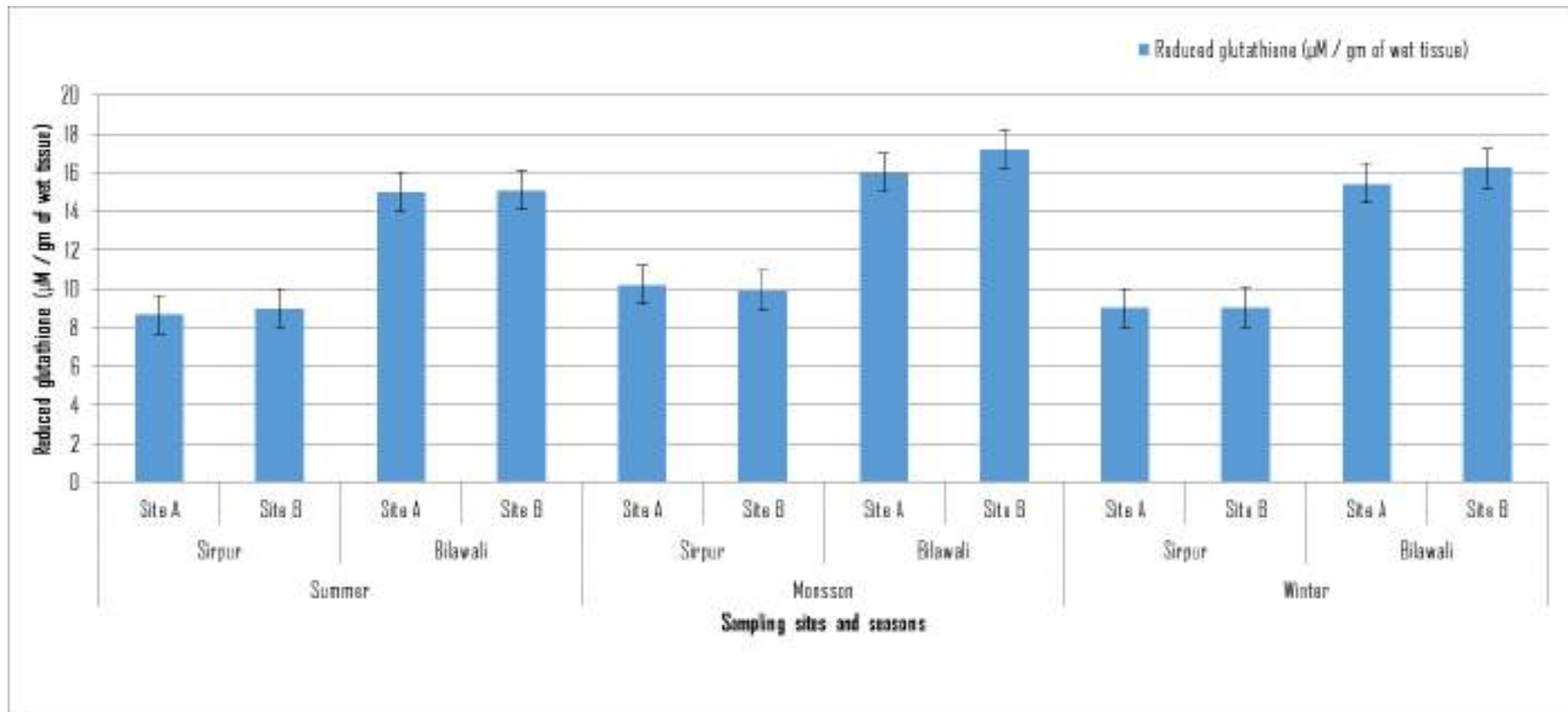
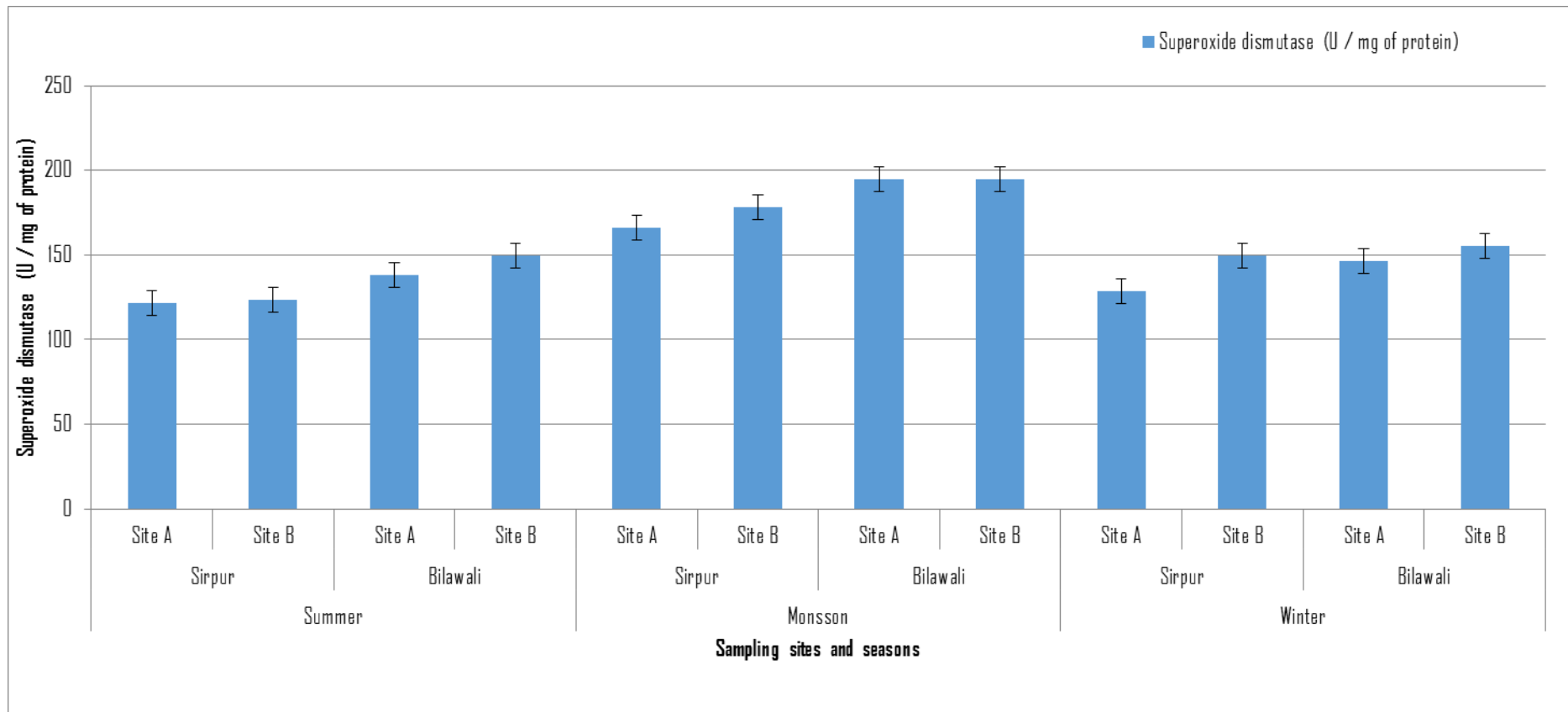


Fig. 42. Superoxide activity in *Lamellidens marginalis* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.



*marginali* bivalve samples collected from Sirpur Lake site A and site B were  $121.45 \pm 1.86$  and  $123.51 \pm 1.68$  in summer;  $165.84 \pm 1.72$  and  $178.42 \pm 1.96$  in monsoon;  $128.37 \pm 1.11$  and  $149.68 \pm 1.94$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $138.25 \pm 1.28$  and  $149.56 \pm 1.15$  in summer;  $194.58 \pm 2.06$  and  $194.53 \pm 1.92$  in monsoon;  $146.27 \pm 2.31$  and  $155.24 \pm 1.65$  in winter seasons respectively.

#### **3.4.2.5 Catalase**

Catalase with standard error per mg protein in collected *Lamellidens marginali* bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 8. Catalase with standard deviation per mg protein in *Lamellidens marginali* bivalve samples collected from Sirpur Lake site A and site B were  $91.2 \pm 1.23$  and  $99.27 \pm 1.22$  in summer;  $135.26 \pm 1.39$  and  $141.26 \pm 1.78$  in monsoon  $119.26 \pm 1.09$  and  $128.67 \pm 1.78$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $98.57 \pm 1.14$  and  $108.69 \pm 1.10$  in summer;  $153.67 \pm 0.09$  and  $158.69 \pm 1.49$  in monsoon;  $120.32 \pm 1.98$  and  $131.27 \pm 1.53$  in winter seasons respectively.

#### **3.4.2.6 Glutathione peroxidase**

Superoxide dismutase with standard error per mg protein in collected *Lamellidens marginali* bivalve samples of Sirpur and Bilawali Tank in



Fig. 43. Catalase activity in *Lamellidens marginali* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.

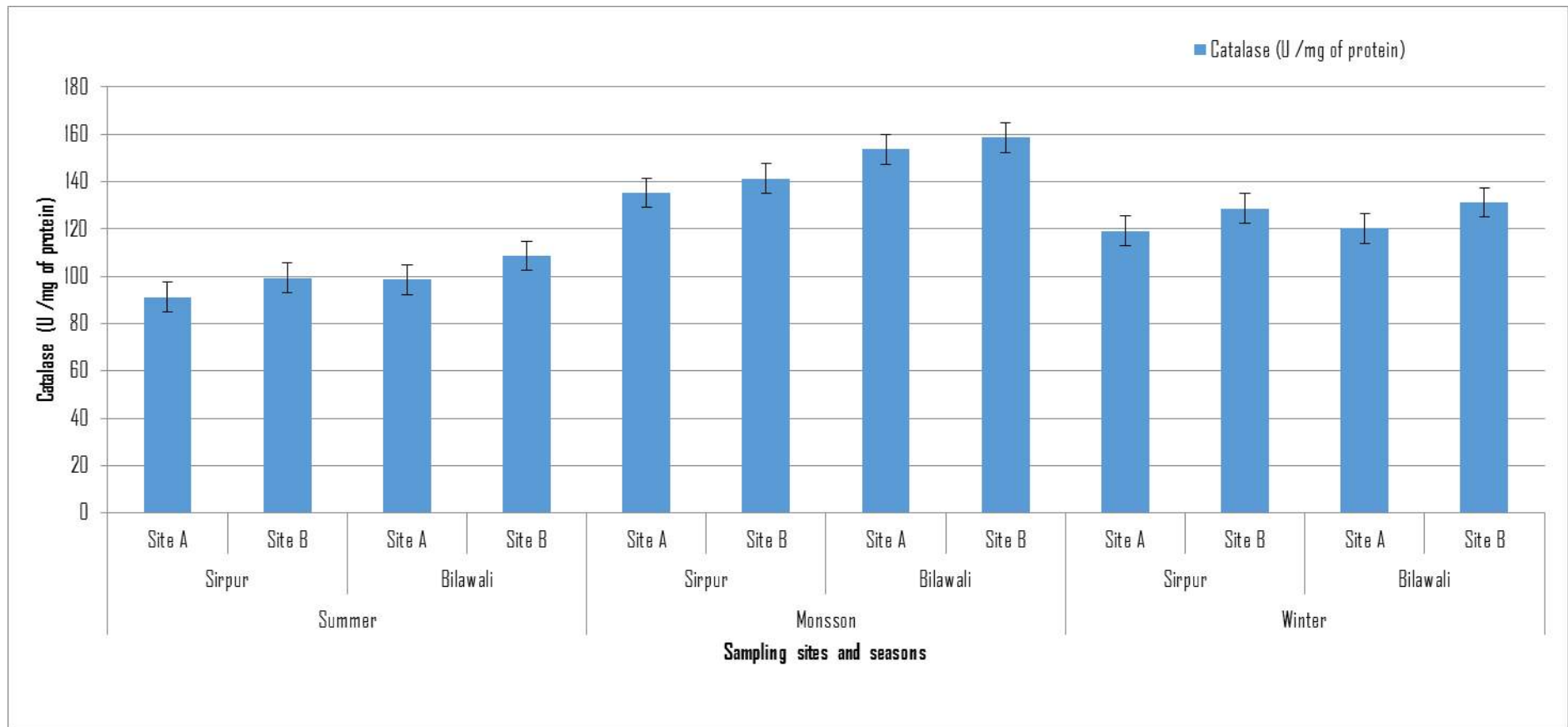
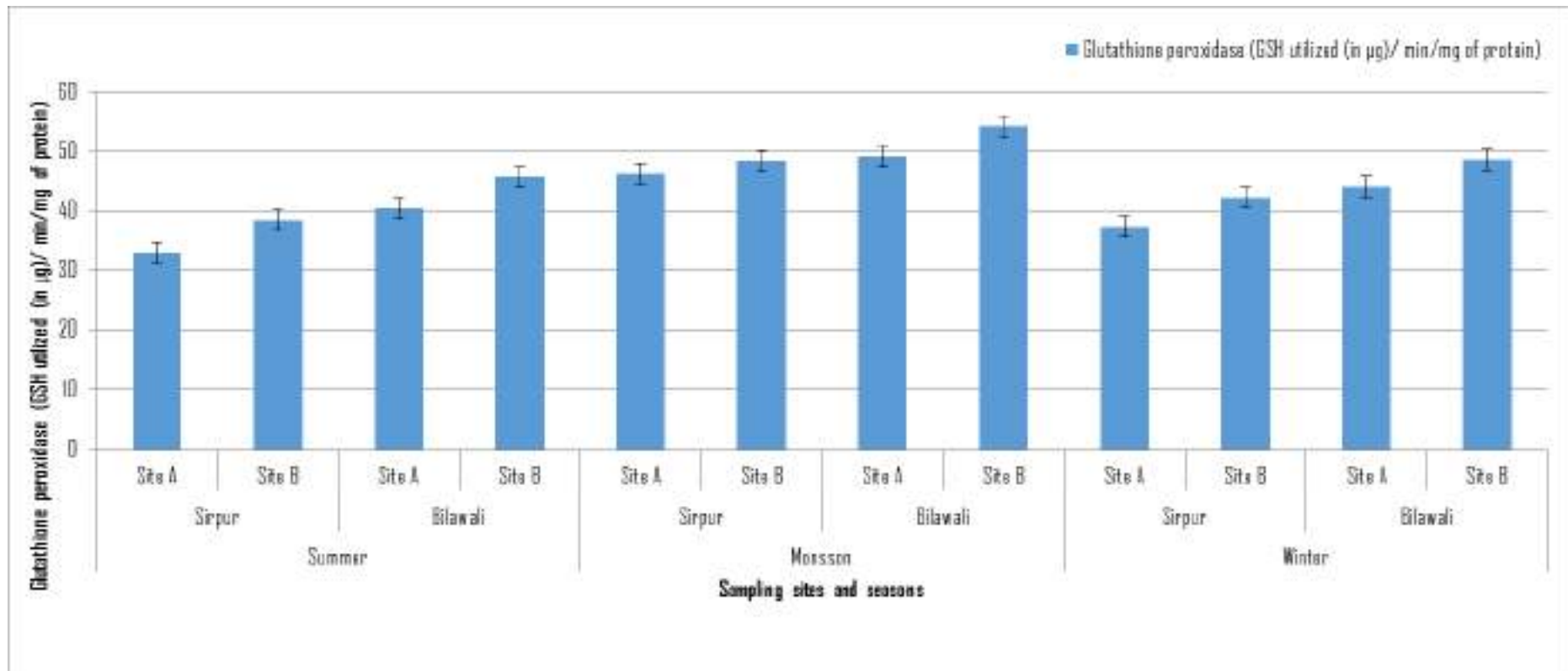


Fig. 44. Glutathione peroxidase activity in *Lamellidens marginali* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.



summer, monsoon and winter seasons is shown in Table 8. Superoxide dismutase with standard deviation per mg protein in *Lamellidens marginali* bivalve samples collected from Sirpur Lake site A and site B were  $33.01\pm 0.13$  and  $38.46\pm 0.12$  in summer;  $46.27\pm 0.12$  and  $48.36\pm 0.13$  in monsoon;  $37.36\pm 0.05$  and  $42.26\pm 0.09$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $40.41\pm 0.11$  and  $45.62\pm 0.08$  in summer;  $49.15\pm 0.14$  and  $54.27\pm 0.14$  in monsoon;  $44.12\pm 0.10$  and  $48.56\pm 0.10$  in winter seasons respectively.

### **3.4.3 *Unio* sp.**

Table 9 presents the mean with standard deviation of collected observations of oxidative biomarkers in *Unio* sp. samples.

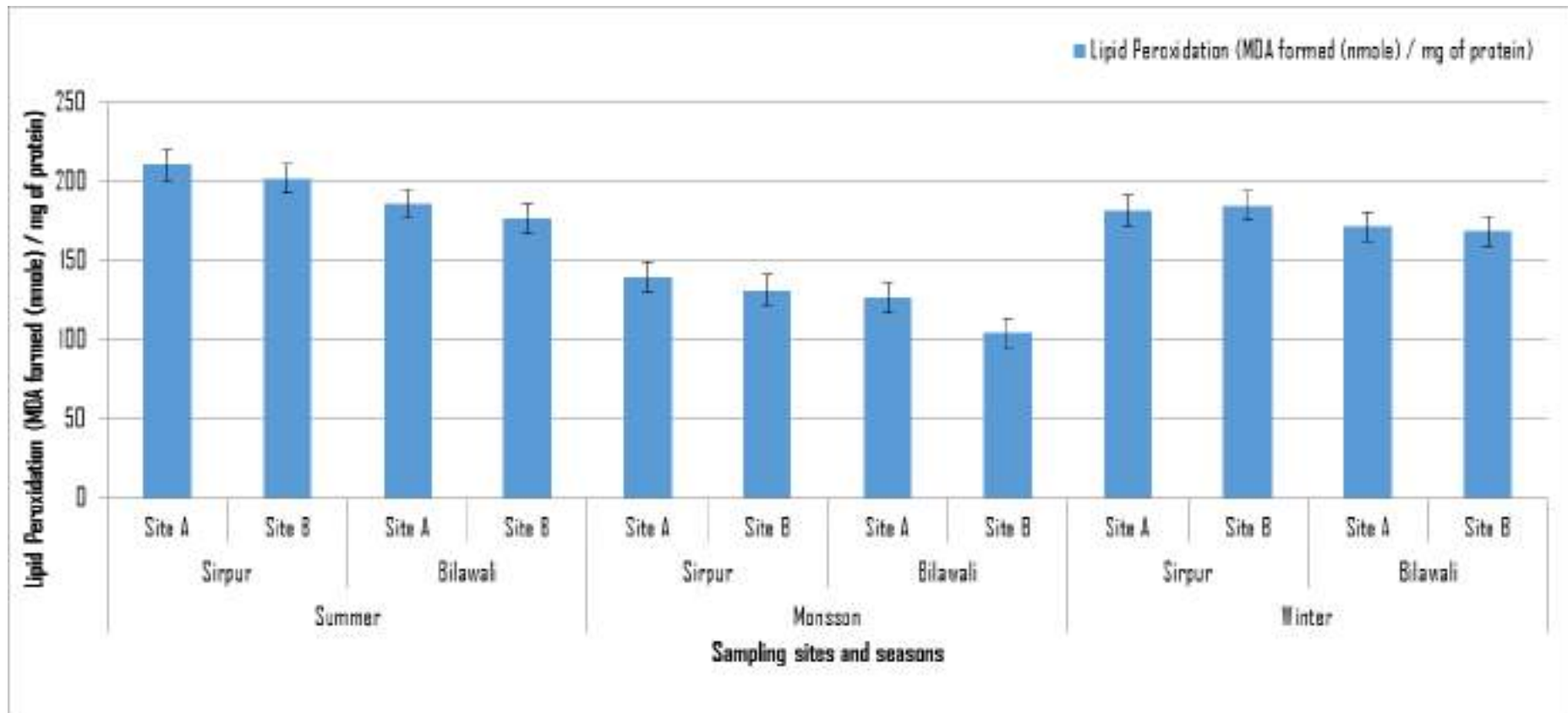
Table 9. Seasonal variations of oxidative stress in *Unio* sp. collected from Sirpur and Bilawali Tanks of Indore.

Parameters	Summer				Monson				Winter			
	Sirpur		Bilawali		Sirpur		Bilawali		Sirpur		Bilawali	
	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B
<b>Lipid Peroxidation</b>	210.23± 5.21	201.45± 5.02	185.74± 5.09	176.59± 5.42	139.57± 4.23	131.42± 4.23	126.25± 4.67	104.23± 3.24	181.45± 5.21	184.56± 4.96	171.25± 4.85	168.46± 4.12
<b>Glutathione-S-transferase</b>	230.18± 5.01	210.52± 4.98	201.56± 4.65	201.45± 5.86	142.56± 4.42	124.62± 3.28	119.51± 3.68	116.24± 3.65	192.45± 4.98	171.24± 3.99	154.86± 3.86	156.28± 3.21
<b>Reduced glutathione</b>	5.86±0.0 9	6.76±0.1 1	9.23±0.0 9	11.23±0. 08	7.01±0.1 0	7.59±0.1 2	11.2±0.1 1	14.26±0. 12	6.95±0.1 1	6.95±0.1 2	11.03±0. 09	13.06±0. 1
<b>Superoxide dismutase</b>	120.45± 2.12	125.64± 2.03	130.45± 1.98	140.25± 1.48	160.58± 1.56	180.46± 1.14	186.54± 1.69	191.42± 1.94	125.34± 2.01	145.36± 3.02	141.29± 2.38	151.24± 2.16
<b>Catalase</b>	88.64±1. 01	96.58±1. 12	105.28± 1.86	105.36± 1.23	129.54± 1.86	143.51± 1.12	154.75± 1.23	152.01± 1.64	124.23± 1.96	121.48± 2.12	118.46± 2.10	128.34± 1.13
<b>Glutathione peroxidase</b>	32.56±0. 16	34.27±0. 11	36.15±0. 14	40.81±0. 11	40.16±0. 07	46.02±0. 10	47.2±0.1 3	51.2±0.1 1	39.56±0. 12	40.58±0. 13	41.03±0. 11	44.52±0. 12

### 3.4.3.1 Lipid peroxidation

Lipid peroxidation with standard error per mg protein in collected *Unio* sp. bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 9. Lipid peroxidation with standard deviation per mg protein in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $210.23 \pm 5.21$  and  $201.45 \pm 5.02$  in summer;  $139.57 \pm 4.23$  and  $131.42 \pm 4.23$  in monsoon;  $181.45 \pm 5.21$  and  $184.56 \pm 4.96$  in winter seasons whereas, Bilawali Tank site A and site B

Fig. 45. Lipid Peroxidation activity in *Unio* sp. bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.



were reported  $185.74 \pm 5.09$  and  $176.59 \pm 5.42$  in summer;  $126.25 \pm 4.67$  and  $104.23 \pm 3.24$  in monsoon;  $171.25 \pm 4.85$  and  $168.46 \pm 4.12$  in winter seasons.

### **3.4.3.2 Glutathione-S-transferase**

Glutathione-S-transferase with standard error per mg protein in collected *Unio* sp. bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 9. Glutathione-S-transferase with standard deviation per mg protein in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $230.18 \pm 5.01$  and  $210.52 \pm 4.98$  in summer  $142.56 \pm 4.42$  and  $124.62 \pm 3.28$  in monsoon;  $192.45 \pm 4.98$  and  $171.24 \pm 3.99$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $201.56 \pm 4.65$  and  $201.45 \pm 5.86$  in summer;  $119.51 \pm 3.68$  and  $116.24 \pm 3.65$  in monsoon;  $154.86 \pm 3.86$  and  $156.28 \pm 3.21$  in winter seasons respectively.

### **3.4.3.3 Reduced glutathione**

Reduced glutathione with standard error per mg protein in collected *Unio* sp. bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 9. Reduced glutathione with standard deviation per mg protein in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $5.86 \pm 0.09$  and  $6.76 \pm 0.11$  in summer;  $7.01 \pm 0.10$  and  $7.59 \pm 0.12$  in

Fig. 46. Glutathione-S-transferase activity in *Unio* sp. bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.

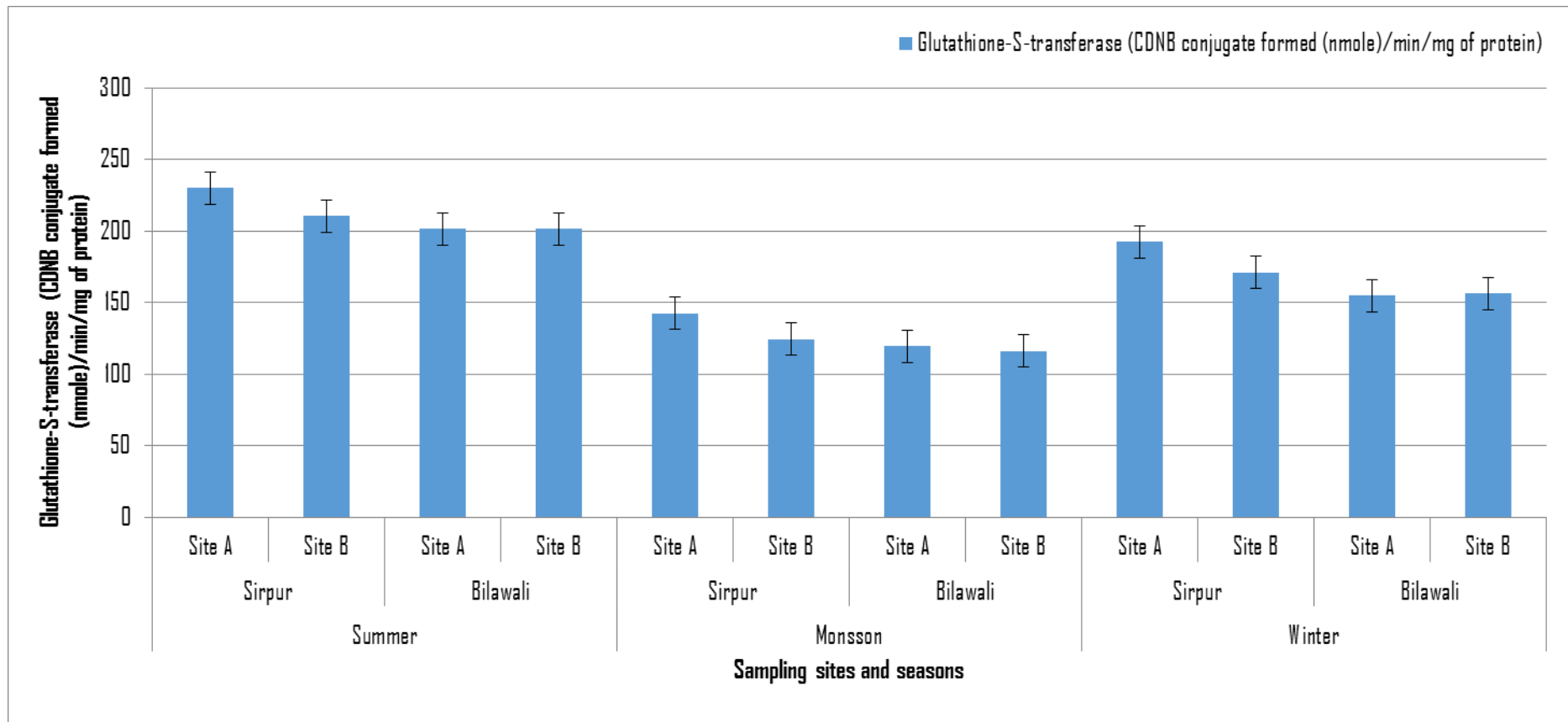
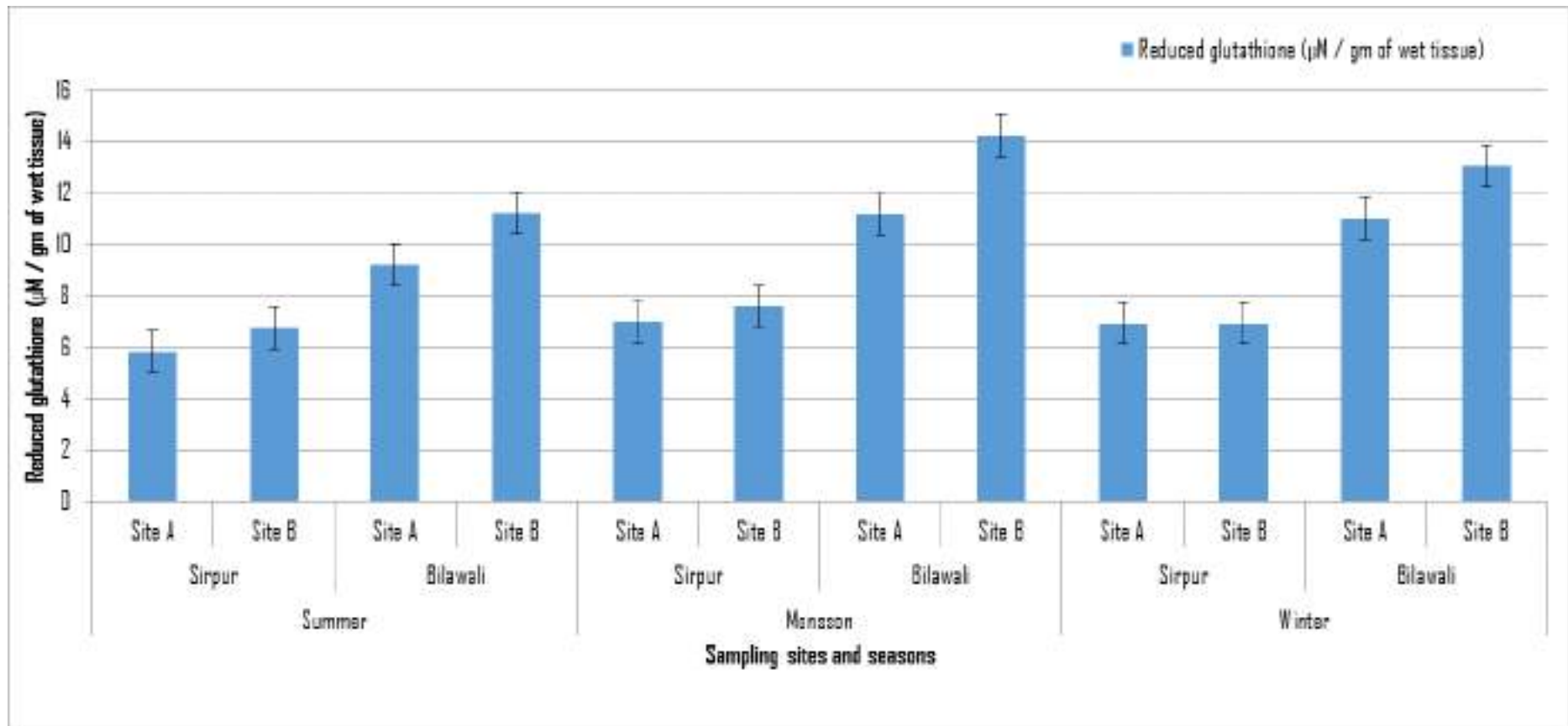




Fig. 47. Reduced glutathione activity in *Unio* sp. bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.



monsoon;  $6.95 \pm 0.11$  and  $6.95 \pm 0.12$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $9.23 \pm 0.09$  and  $11.23 \pm 0.08$  in summer;  $11.2 \pm 0.11$  and  $14.26 \pm 0.12$  in monsoon;  $11.03 \pm 0.09$  and  $13.06 \pm 0.1$  in winter seasons respectively.

#### **3.4.3.4 Superoxide dismutase:**

Superoxide dismutase with standard error per mg protein in collected *Unio* sp. bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 9. Superoxide dismutase with standard deviation per mg protein in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $120.45 \pm 2.12$  and  $125.64 \pm 2.03$  in summer;  $160.58 \pm 1.56$  and  $180.46 \pm 1.14$  in monsoon;  $125.34 \pm 2.01$  and  $145.36 \pm 3.02$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $130.45 \pm 1.98$  and  $140.25 \pm 1.48$  in summer;  $186.54 \pm 1.69$  and  $191.42 \pm 1.94$  in monsoon;  $141.29 \pm 2.38$  and  $151.24 \pm 2.16$  in winter seasons respectively.

#### **3.4.3.5 Catalase**

Catalase with standard error per mg protein in collected *Unio* sp. bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 9. Catalase with standard deviation per mg protein in *Unio* sp. bivalve samples collected from Sirpur Lake site A

Fig. 48. Superoxide activity in *Unio* sp. bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.

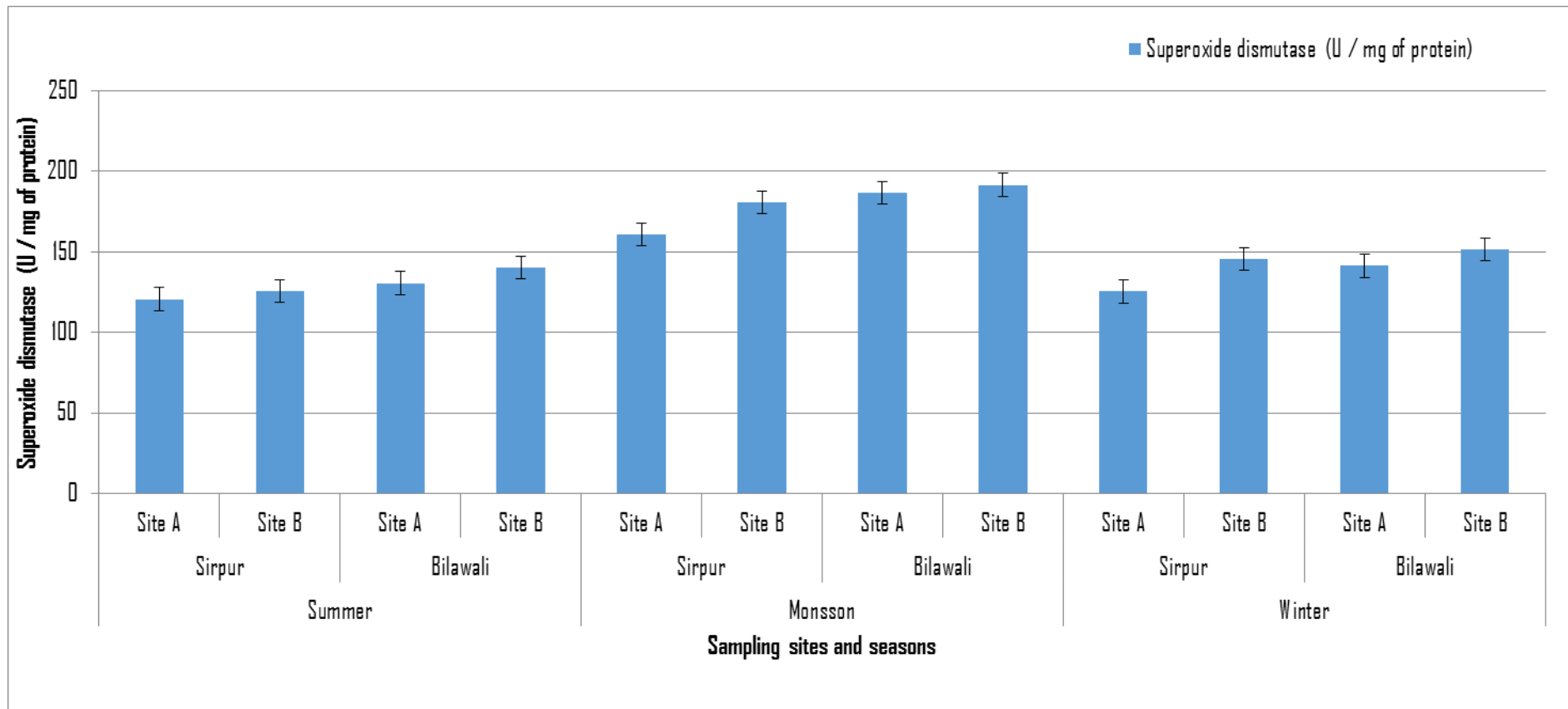
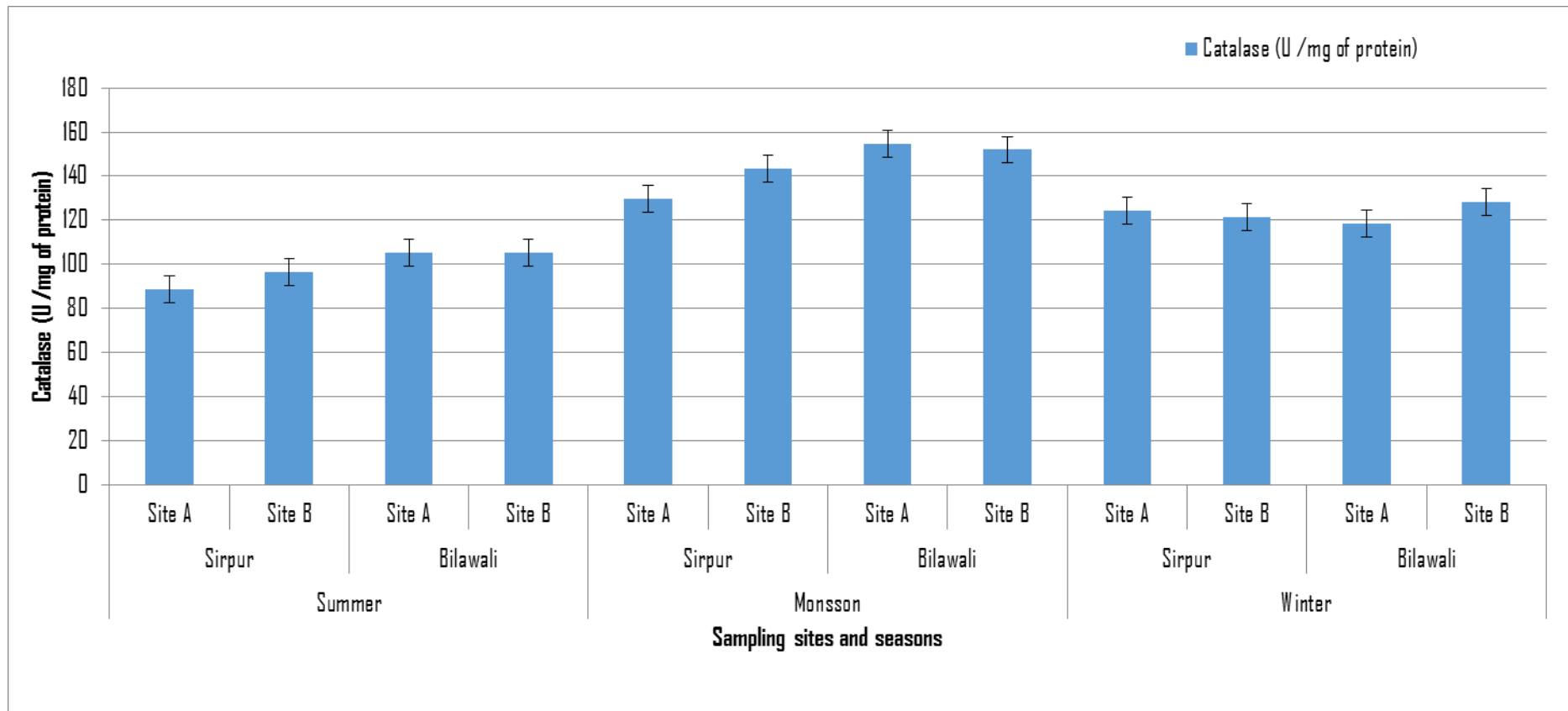


Fig. 49. Catalase activity in *Unio* sp. bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.

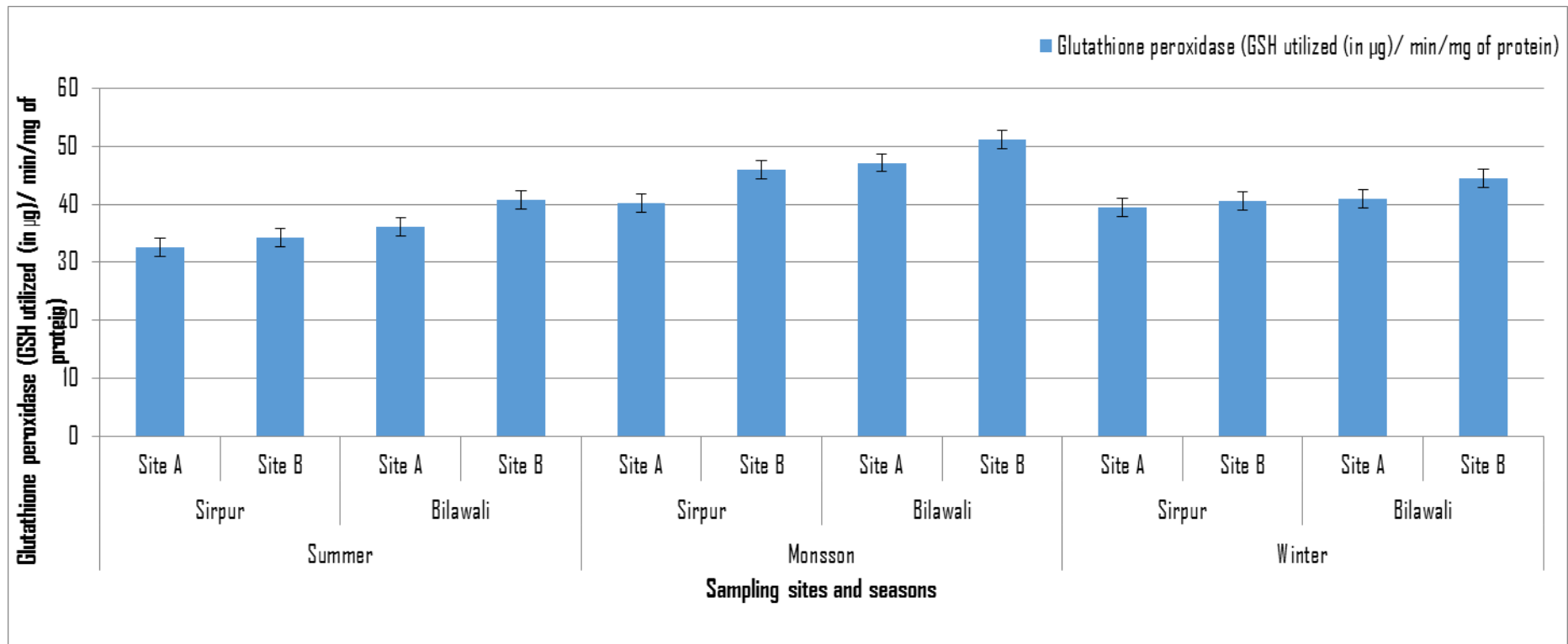


and site B were  $88.64 \pm 1.01$  and  $96.58 \pm 1.12$  in summer;  $129.54 \pm 1.86$  and  $143.51 \pm 1.12$  in monsoon  $124.23 \pm 1.96$  and  $121.48 \pm 2.12$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $105.28 \pm 1.86$  and  $105.36 \pm 1.23$  in summer;  $154.75 \pm 1.23$  and  $152.01 \pm 1.64$  in monsoon;  $118.46 \pm 2.10$  and  $128.34 \pm 1.13$  in winter seasons respectively.

#### **3.4.3.6 Glutathione peroxidase**

Superoxide dismutase with standard error per mg protein in collected *Unio* sp. bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 9. Superoxide dismutase with standard deviation per mg protein in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $32.56 \pm 0.16$  and  $34.27 \pm 0.11$  in summer;  $40.16 \pm 0.07$  and  $46.02 \pm 0.10$  in monsoon;  $39.56 \pm 0.12$  and  $40.58 \pm 0.13$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $36.15 \pm 0.14$  and  $40.81 \pm 0.11$  in summer;  $47.2 \pm 0.13$  and  $51.2 \pm 0.11$  in monsoon;  $41.03 \pm 0.11$  and  $44.52 \pm 0.12$  in winter seasons respectively.

Fig. 50. Glutathione peroxidase activity in *Unio* sp. bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.



### **3.5 Biochemical studies in bivalves:**

Biochemical studies viz. Protein, Ascorbic acid ,RNA and DNA were assessed in *Lamellidens corrianus*, *Lamellidens marginali* and *Unio* sp bivalves samples collected from two different sites of Sirpur and Bilawali Tanks, Indore in summer, monsoon and winter seasons and presented in table 10-12.

#### **3.5.1 *Lamellidens corrianus***

Table 10 presents the mean with standard deviation of collected observations of biochemical studies in *Lamellidens corrianus* samples.

Table 10. Seasonal variations of biochemical parameters in *Lamellidens corrianus* collected from Sirpur and Bilawali Tanks of Indore.

Body part	Summer				Monson				Winter			
	Sirpur		Bilawali		Sirpur		Bilawali		Sirpur		Bilawali	
	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B
<b>PROTEIN</b>												
<b>Gills</b>	49.59±1.25	52.01±1.34	52.42±1.23	53.21±1.47	60.25±1.58	61.56±1.37	63.24±1.34	63.15±1.28	59.68±1.19	61.29±1.39	61.02±1.49	61.05±1.39
<b>Digestive gland</b>	47.56±1.12	51.2±1.42	50.27±1.42	51.12±1.49	61.10±1.29	61.03±1.29	62.13±1.31	64.29±1.22	57.62±1.18	59.68±1.38	59.61±1.38	60.31±1.39
<b>Whole soft body</b>	48.51±1.14	49.35±1.29	49.23±1.29	50.01±1.43	58.67±1.27	60.01±1.37	60.03±1.27	61.20±1.31	53.24±1.11	56.24±1.34	57.23±1.36	59.64±1.32
<b>ASCORBIC ACID</b>												
<b>Gills</b>	0.712±0.001	0.786±0.003	0.805±0.001	0.814±0.001	1.005±0.005	1.132±0.009	1.142±0.011	1.112±0.009	0.986±0.009	1.002±0.009	1.009±0.008	1.008±0.009
<b>Digestive gland</b>	0.856±0.002	0.886±0.003	0.926±0.002	0.953±0.002	1.103±0.005	1.296±0.010	1.298±0.012	1.236±0.010	1.023±0.009	1.124±0.010	1.143±0.010	1.112±0.008
<b>Whole soft body</b>	0.702±0.001	0.736±0.002	0.792±0.001	0.756±0.001	1.009±0.004	1.124±0.010	1.136±0.009	1.152±0.012	0.958±0.008	0.995±0.009	0.995±0.009	1.020±0.008
<b>DNA</b>												
<b>Gills</b>	1.12±0.23	1.18±0.22	1.23±0.21	1.29±0.21	1.46±0.23	1.36±0.21	1.76±0.22	1.86±0.23	1.62±0.12	1.54±0.21	1.56±0.22	1.59±0.23
<b>Digestive gland</b>	1.19±0.20	1.25±0.19	1.31±0.22	1.35±0.23	1.79±0.23	1.85±0.22	1.89±0.23	1.97±0.22	1.63±0.17	1.59±0.22	1.62±0.23	1.68±0.22
<b>Whole soft body</b>	1.23±0.21	1.32±0.14	1.41±0.23	1.42±0.21	1.94±0.25	1.99±0.23	1.95±0.23	1.96±0.21	1.79±0.21	1.86±0.23	1.95±0.22	1.94±0.23



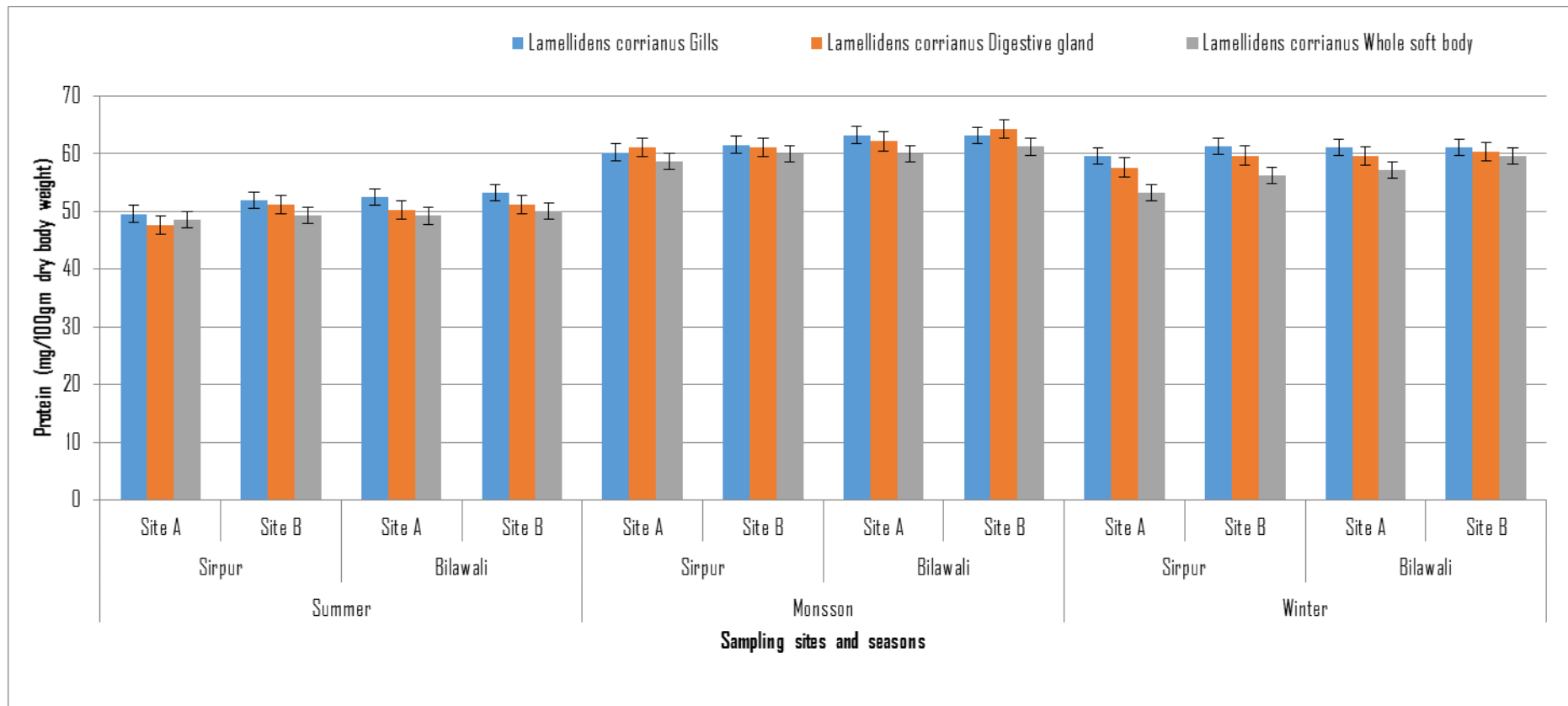
RNA												
Body part	Summer				Monsoon				Winter			
	Sirpur		Bilawali		Sirpur		Bilawali		Sirpur		Bilawali	
	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B
<b>Gills</b>	5.01±0.34	5.98±0.41	6.07±0.33	6.12±0.30	7.01±0.29	8.36±0.33	8.49±0.41	8.50±0.39	7.50±0.38	8.96±0.42	9.08± 0.37	9.10±0.43
<b>Digestive gland</b>	6.39± 0.41	6.89± 0.33	7.02± 0.37	7.19±0.32	8.94±0.37	9.63±0.27	9.82±0.43	9.98±0.42	9.57±0.42	9.61±0.39	9.96± 0.39	9.69±0.39
<b>Whole soft body</b>	4.42± 0.29	4.93± 0.27	5.96± 0.36	6.02±0.44	6.18±0.28	6.89±0.27	8.34±0.41	8.36±0.39	6.62±0.34	7.38±0.43	8.91± 0.41	8.95±0.47

### 3.5.1.1 Protein

Protein with standard error per mg dry weight of gills in collected *Lamellidens corrianus* bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 10. Protein with standard deviation per mg dry weight of gills in *Lamellidens corrianus* bivalve samples collected from Sirpur Lake site A and site B were  $49.59 \pm 1.25$  and  $52.01 \pm 1.34$  in summer;  $60.25 \pm 1.58$  and  $61.56 \pm 1.37$  in monsoon;  $59.68 \pm 1.19$  and  $61.29 \pm 1.39$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $52.42 \pm 1.23$  and  $53.21 \pm 1.47$  in summer;  $63.24 \pm 1.34$  and  $63.15 \pm 1.28$  in monsoon;  $61.02 \pm 1.49$  and  $61.05 \pm 1.39$  in winter seasons respectively. Protein with standard deviation per mg dry weight of digestive gland in *Lamellidens corrianus* bivalve samples collected from Sirpur Lake site A and site B were  $47.56 \pm 1.12$  and  $51.2 \pm 1.42$  in summer;  $61.10 \pm 1.29$  and  $61.03 \pm 1.29$  in monsoon;  $57.62 \pm 1.18$  and  $59.68 \pm 1.38$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $50.27 \pm 1.42$  and  $51.12 \pm 1.49$  in summer;  $62.13 \pm 1.31$  and  $64.29 \pm 1.22$  in monsoon;  $59.61 \pm 1.38$  and  $60.31 \pm 1.39$  in winter seasons respectively. Protein with standard deviation per mg dry weight of whole body soft tissue in *Lamellidens corrianus* bivalve samples collected from Sirpur Lake site A and site B were  $48.51 \pm 1.14$  and  $49.35 \pm 1.29$  in summer;  $58.67 \pm 1.27$  and  $60.01 \pm 1.37$  in monsoon;  $53.24 \pm 1.11$  and  $56.24 \pm 1.34$  in winter

seasons whereas, Bilawali Tank site A and site B were reported 49.23±1.29 and 50.01±1.43 in summer; 60.03±1.27 and 61.20±1.31 in monsoon; 57.23±1.36 and 59.64±1.32 in winter seasons respectively.

Fig. 51. Total protein in gills, digestive gland and whole soft body of *Lamellidens corrianus* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.

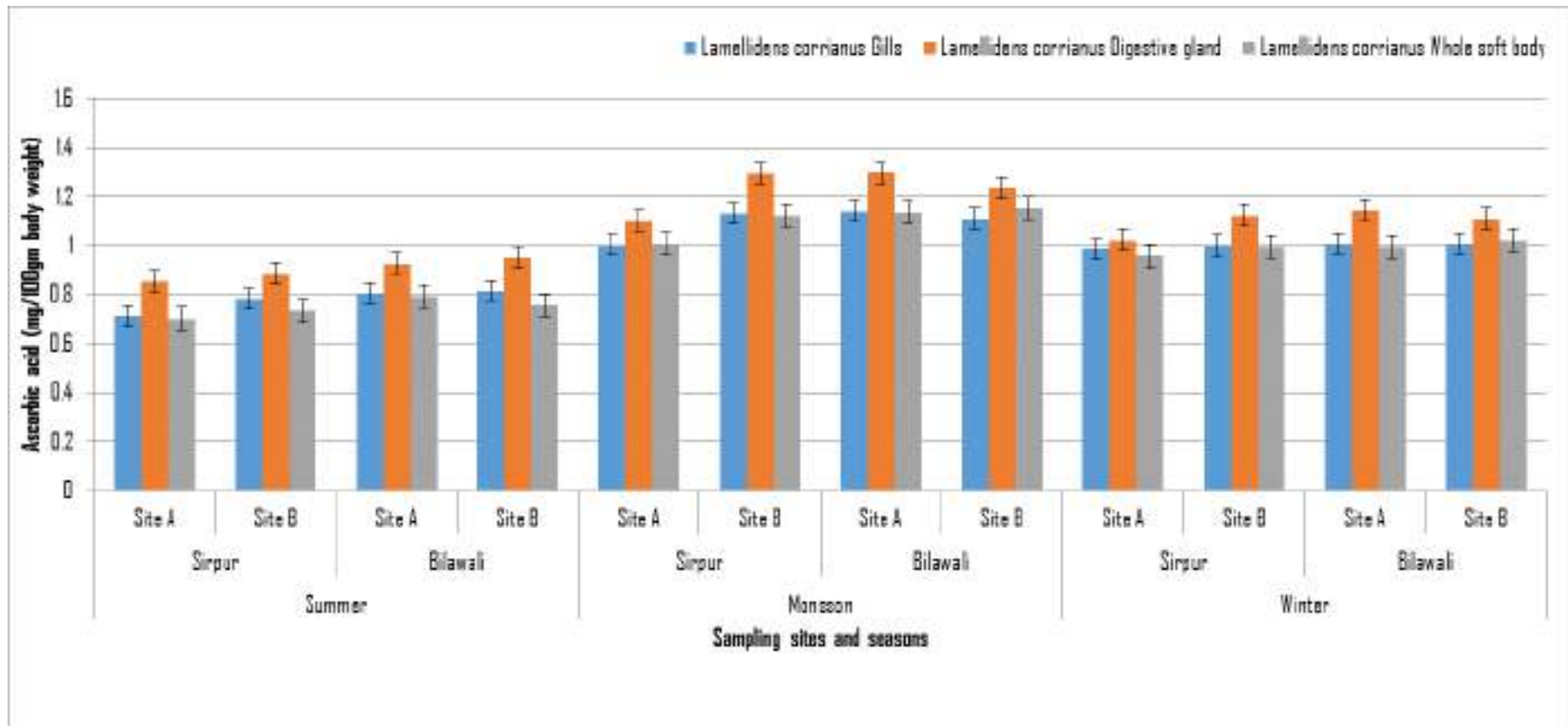


### 3.5.1.2 Ascorbic acid

Ascorbic acid with standard error per mg dry weight of gills in collected *Lamellidens corrianus* bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 9. Ascorbic acid with standard deviation per mg dry weight of gills in *Lamellidens corrianus* bivalve samples collected from Sirpur Lake site A and site B were  $0.712\pm 0.001$  and  $0.786\pm 0.003$  in summer;  $1.005\pm 0.005$  and  $1.132\pm 0.009$  in monsoon;  $0.986\pm 0.009$  and  $1.002\pm 0.009$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $0.805\pm 0.001$  and  $0.814\pm 0.001$  in summer;  $1.142\pm 0.011$  and  $1.112\pm 0.009$  in monsoon;  $1.009\pm 0.008$  and  $1.008\pm 0.009$  in winter seasons respectively. Ascorbic acid with standard deviation per mg dry weight of digestive gland in *Lamellidens corrianus* bivalve samples collected from Sirpur Lake site A and site B were  $0.856\pm 0.002$  and  $0.886\pm 0.003$  in summer;  $1.103\pm 0.005$  and  $1.296\pm 0.010$  in monsoon;  $1.023\pm 0.009$  and  $1.124\pm 0.010$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $0.926\pm 0.002$  and  $0.953\pm 0.002$  in summer;  $1.298\pm 0.012$  and  $1.236\pm 0.010$  in monsoon;  $1.143\pm 0.010$  and  $1.112\pm 0.008$  in winter seasons respectively. Ascorbic acid with standard deviation per mg dry weight of whole body soft tissue in *Lamellidens corrianus* bivalve samples collected from Sirpur Lake site A and site B were  $0.702\pm 0.001$  and  $0.736\pm 0.002$  in summer;  $1.009\pm 0.004$  and  $1.124\pm 0.010$  in monsoon;  $0.958\pm 0.008$  and  $0.995\pm 0.009$  in winter seasons whereas, Bilawali Tank

site A and site B were reported  $0.792\pm 0.001$  and  $0.756\pm 0.001$  in summer;  $1.136\pm 0.009$  and  $1.152\pm 0.012$  in monsoon;  $0.995\pm 0.009$  and  $1.020\pm 0.008$  in winter seasons respectively.

Fig. 52. Ascorbic acid in gills, digestive gland and whole soft body of *Lamellidens corrianus* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.

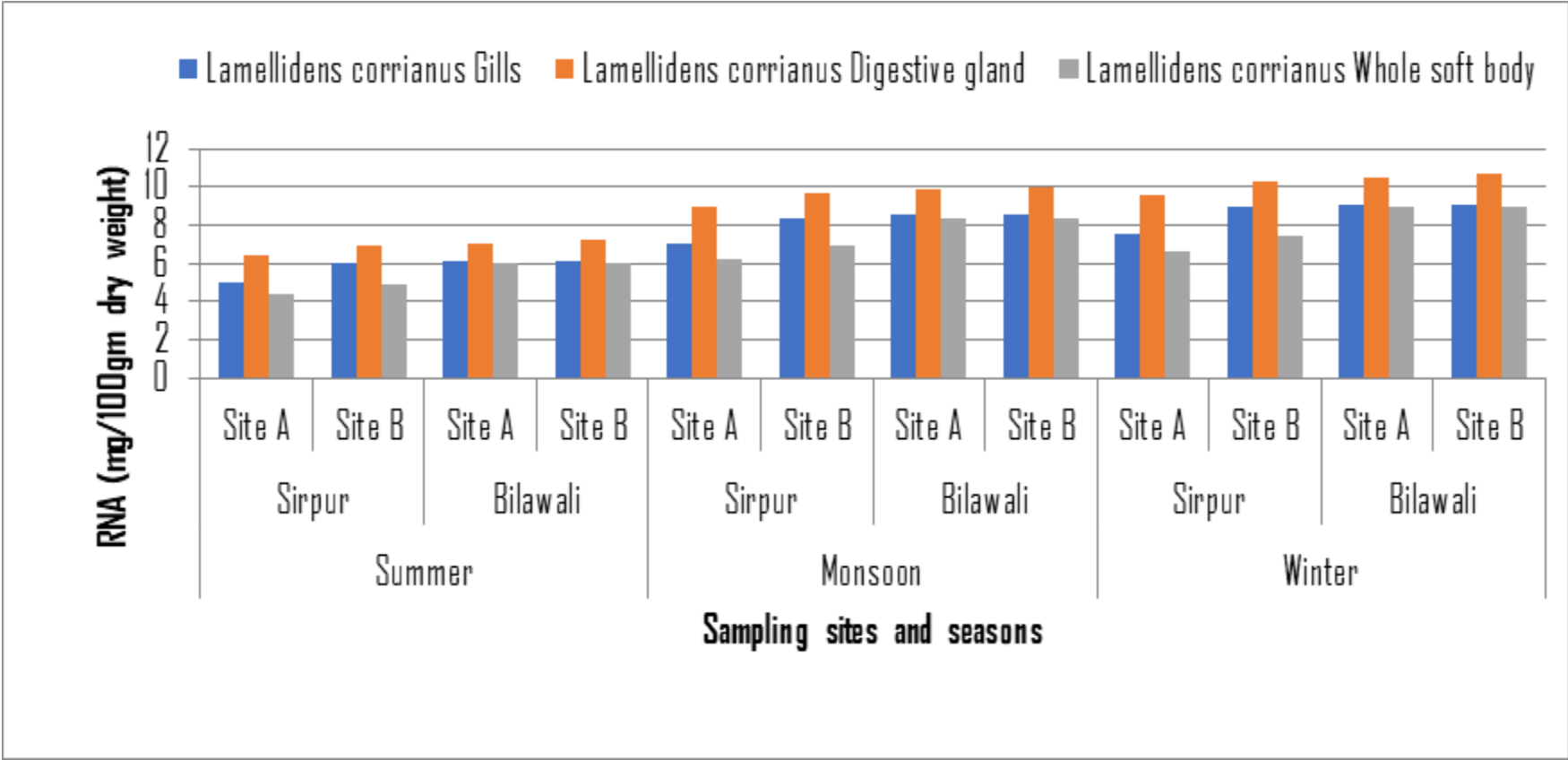


### 3.5.1.3 RNA

RNA with standard error per mg dry weight of gills in collected *Lamellidens corrianus* bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 9. RNA with standard deviation per mg dry weight of gills in *Lamellidens corrianus* bivalve samples collected from Sirpur Lake site A and site B were  $5.01 \pm 0.34$  and  $5.98 \pm 0.41$  in summer;  $7.01 \pm 0.29$  and  $8.36 \pm 0.33$  in monsoon;  $7.50 \pm 0.38$  and  $8.96 \pm 0.42$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $6.07 \pm 0.33$  and  $6.12 \pm 0.30$  in summer;  $8.49 \pm 0.41$  and  $8.50 \pm 0.39$  in monsoon;  $9.08 \pm 0.37$  and  $9.10 \pm 0.43$  in winter seasons respectively. RNA with standard deviation per mg dry weight of digestive gland in *Lamellidens corrianus* bivalve samples collected from Sirpur Lake site A and site B were  $6.39 \pm 0.41$  and  $6.89 \pm 0.33$  in summer;  $8.94 \pm 0.37$  and  $9.63 \pm 0.27$  in monsoon;  $9.57 \pm 0.42$  and  $9.61 \pm 0.39$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $7.02 \pm 0.37$  and  $7.19 \pm 0.32$  in summer;  $9.82 \pm 0.43$  and  $9.98 \pm 0.42$  in monsoon;  $9.96 \pm 0.39$  and  $9.69 \pm 0.39$  in winter seasons respectively. RNA with standard deviation per mg dry weight of whole body soft tissue in *Lamellidens corrianus* bivalve samples collected from Sirpur Lake site A and site B were  $4.42 \pm 0.29$  and  $4.93 \pm 0.27$  in summer;  $6.18 \pm 0.28$  and  $6.89 \pm 0.27$  in monsoon;  $6.62 \pm 0.34$  and  $7.38 \pm 0.43$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $5.96 \pm 0.36$  and  $6.02 \pm 0.44$  in summer;  $8.34 \pm 0.41$  and  $8.36 \pm 0.39$  in monsoon;  $8.91 \pm 0.41$  and  $8.95 \pm 0.47$  in winter seasons respectively



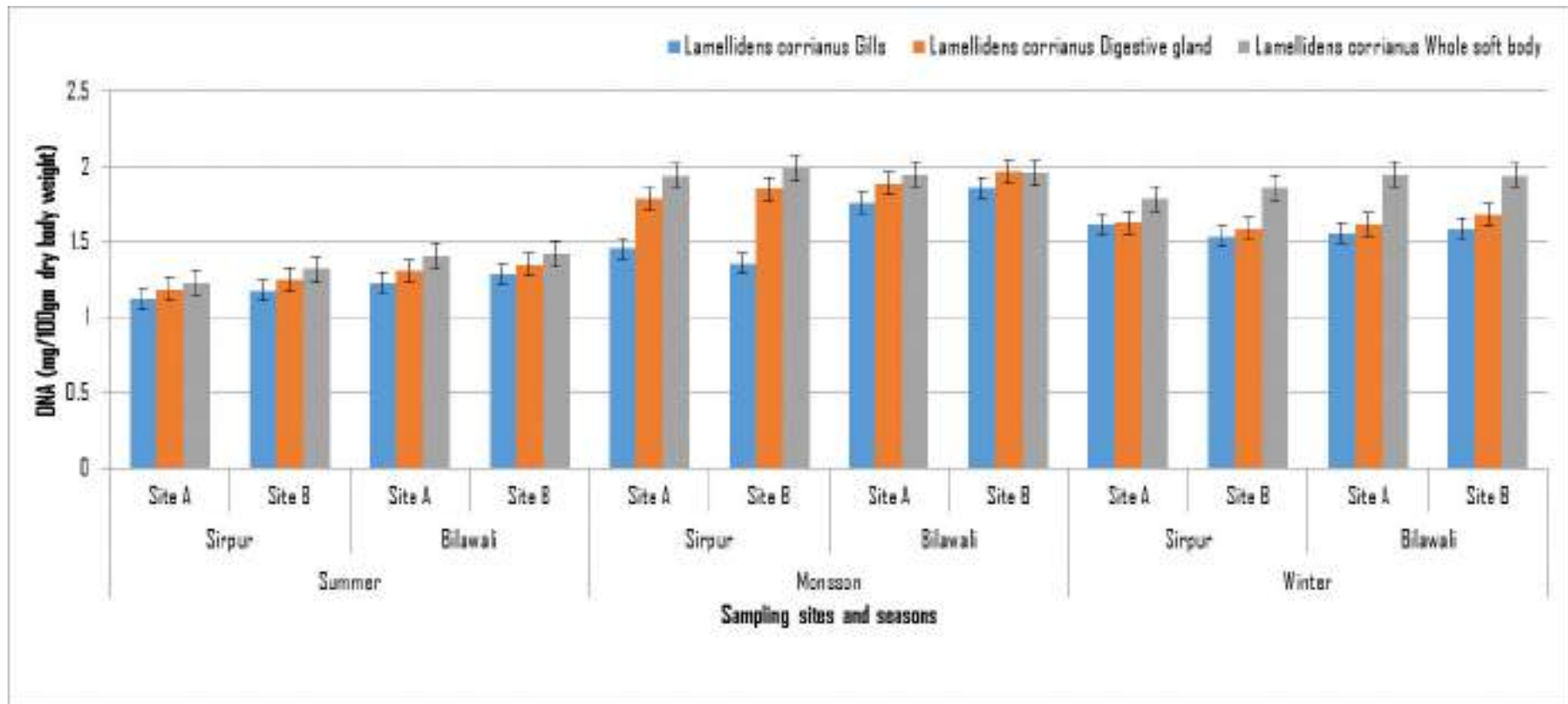
Fig. 53. RNA in gills, digestive gland and whole soft body of *Lamellidens corrianus* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons



#### 3.5.1.4 DNA

DNA with standard error per mg dry weight of gills in collected *Lamellidens corrianus* bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 9. DNA with standard deviation per mg dry weight of gills in *Lamellidens corrianus* bivalve samples collected from Sirpur Lake site A and site B were  $1.12 \pm 0.23$  and  $1.18 \pm 0.22$  in summer;  $1.46 \pm 0.23$  and  $1.36 \pm 0.21$  in monsoon;  $1.62 \pm 0.12$  and  $1.54 \pm 0.21$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $1.23 \pm 0.21$  and  $1.29 \pm 0.21$  in summer;  $1.76 \pm 0.22$  and  $1.86 \pm 0.23$  in monsoon;  $1.56 \pm 0.22$  and  $1.59 \pm 0.23$  in winter seasons respectively. DNA with standard deviation per mg dry weight of digestive gland in *Lamellidens corrianus* bivalve samples collected from Sirpur Lake site A and site B were  $1.19 \pm 0.20$  and  $1.25 \pm 0.19$  in summer;  $1.79 \pm 0.23$  and  $1.85 \pm 0.22$  in monsoon;  $1.63 \pm 0.17$  and  $1.59 \pm 0.22$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $1.31 \pm 0.22$  and  $1.35 \pm 0.23$  in summer;  $1.89 \pm 0.23$  and  $1.97 \pm 0.22$  in monsoon;  $1.62 \pm 0.23$  and  $1.68 \pm 0.22$  in winter seasons respectively. DNA with standard deviation per mg dry weight of whole body soft tissue in *Lamellidens corrianus* bivalve samples collected from Sirpur Lake site A and site B were  $1.23 \pm 0.21$  and  $1.32 \pm 0.14$  in summer;  $1.94 \pm 0.25$  and  $1.99 \pm 0.23$  in monsoon;  $1.79 \pm 0.21$  and  $1.86 \pm 0.23$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $1.41 \pm 0.23$  and  $1.42 \pm 0.21$  in summer;  $1.95 \pm 0.23$  and  $1.96 \pm 0.21$  in monsoon;  $1.95 \pm 0.22$  and  $1.94 \pm 0.23$  in winter seasons respectively.

Fig. 54. DNA in gills, digestive gland and whole soft body of *Lamellidens corrianus* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.



### 3.5.2 *Lamellidens marginali*

Table 11 presents the mean with standard deviation of collected observations of biochemical studies in *Lamellidens marginali* samples.

Table 11. Seasonal variations of biochemical parameters in *Lamellidens marginali* collected from Sirpur and Bilawali Tanks of Indore.

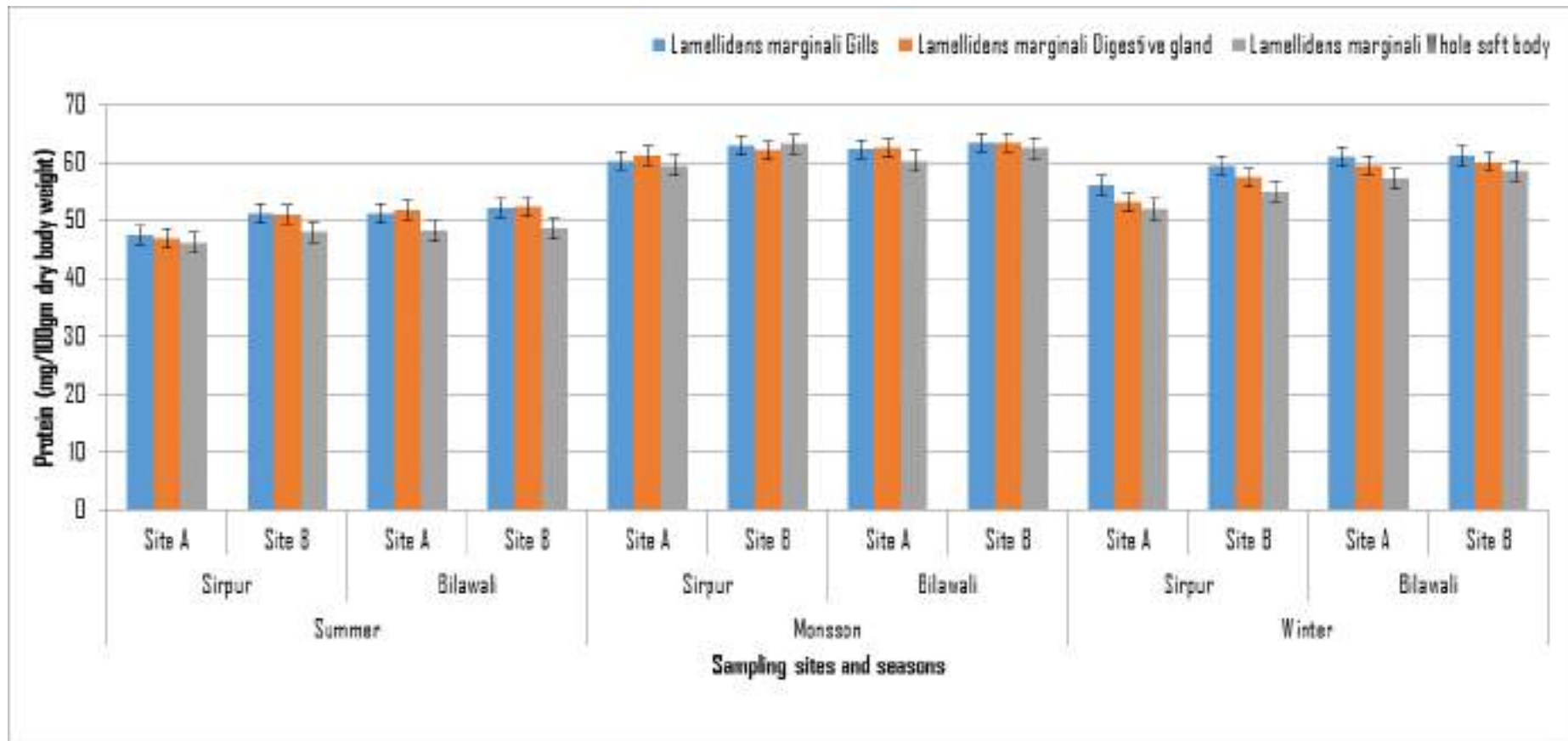
Body part	Summer				Monson				Winter			
	Sirpur		Bilawali		Sirpur		Bilawali		Sirpur		Bilawali	
	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B
<b>PROTEIN</b>												
<b>Gills</b>	47.52±1.23	51.24±1.41	51.24±1.29	52.16±1.47	60.29±1.34	62.98±1.41	62.46±1.41	63.59±1.36	56.16±1.15	59.63±1.29	61.02±1.36	61.25±1.41
<b>Digestive gland</b>	46.89±1.15	51.11±1.39	51.82±1.30	52.36±1.49	61.24±1.45	62.14±1.28	62.53±1.39	63.49±1.32	53.24±1.14	57.42±1.27	59.64±1.41	60.12±1.36
<b>Whole soft body</b>	46.25±1.10	48.10±1.15	48.24±1.41	48.67±1.23	59.58±1.39	63.25±1.32	60.28±1.27	62.59±1.37	52.03±1.14	55.12±1.21	57.26±1.29	58.59±1.30
<b>ASCORBIC ACID</b>												
<b>Gills</b>	0.776±0.001	0.812±0.003	0.812±0.002	0.824±0.002	1.100±0.006	1.129±0.009	1.149±0.012	1.186±0.009	0.986±0.008	0.997±0.009	0.997±0.008	1.061±0.008
<b>Digestive gland</b>	0.882±0.002	0.945±0.004	0.964±0.003	0.942±0.003	1.235±0.006	1.325±0.011	1.324±0.019	1.321±0.010	1.023±0.009	1.129±0.010	1.124±0.010	1.123±0.009
<b>Whole soft body</b>	0.721±0.001	0.795±0.002	0.752±0.001	0.806±0.002	1.112±0.004	1.142±0.009	1.145±0.012	1.154±0.010	0.984±0.008	0.998±0.009	1.023±0.010	1.006±0.010
<b>DNA</b>												
<b>Gills</b>	1.12±0.21	1.24±0.26	1.22±0.19	1.31±0.23	1.63±0.21	1.79±0.20	1.88±0.21	1.84±0.22	1.48±0.20	1.62±0.21	1.62±0.19	1.68±0.20
<b>Digestive gland</b>	1.22±0.28	1.25±0.23	1.31±0.18	1.33±0.19	1.68±0.21	1.91±0.19	1.76±0.19	1.96±0.21	1.39±0.23	1.54±0.21	1.64±0.19	1.71±0.1
<b>Whole soft body</b>	1.29±0.22	1.35±0.22	1.42±0.23	1.42±0.19	1.94±0.19	1.98±0.20	1.98±0.19	1.98±0.19	1.89±0.21	1.92±0.22	1.95±0.20	1.93±0.29

RNA												
Body part	Summer				Monsoon				Winter			
	Sirpur		Bilawali		Sirpur		Bilawali		Sirpur		Bilawali	
	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B
Gills	5.07± 0.28	6.01±0.38	6.13± 0.34	6.24± 0.29	7.09± 0.37	8.40± 0.39	8.58± 0.44	8.66± 0.33	7.59± 0.33	9.00± 0.41	9.17± 0.42	9.28± 0.43
Digestive gland	6.22± 0.33	7.03±0.24	7.20± 0.41	7.31± 0.27	8.70± 0.39	9.49± 0.31	9.81± 0.45	9.59± 0.37	9.31± 0.37	9.43± 0.46	9.92± 0.62	9.63± 0.53
Whole soft body	4.59± 0.21	5.71±0.39	6.04± 0.38	6.22± 0.35	6.42± 0.27	7.98± 0.33	8.45± 0.39	8.63± 0.41	6.87± 0.38	8.55± 0.46	9.03± 0.39	9.25± 0.35

### 3.5.2.1 Protein

Protein with standard error per mg dry weight of gills in collected *Lamellidens marginali* bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 11. Protein with standard deviation per mg dry weight of gills in *Lamellidens marginali* bivalve samples collected from Sirpur Lake site A and site B were  $49.59 \pm 1.25$  and  $52.01 \pm 1.34$  in summer;  $60.25 \pm 1.58$  and  $61.56 \pm 1.37$  in monsoon;  $59.68 \pm 1.19$  and  $61.29 \pm 1.39$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $52.42 \pm 1.23$  and  $53.21 \pm 1.47$  in summer;  $63.24 \pm 1.34$  and  $63.15 \pm 1.28$  in monsoon;  $61.02 \pm 1.49$  and  $61.05 \pm 1.39$  in winter seasons respectively. Protein with standard deviation per mg dry weight of digestive gland in *Lamellidens marginali* bivalve samples collected from Sirpur Lake site A and site B were  $47.56 \pm 1.12$  and  $51.2 \pm 1.42$  in summer;  $61.10 \pm 1.29$  and  $61.03 \pm 1.29$  in monsoon;  $57.62 \pm 1.18$  and  $59.68 \pm 1.38$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $50.27 \pm 1.42$  and  $51.12 \pm 1.49$  in summer;  $62.13 \pm 1.31$  and  $64.29 \pm 1.22$  in monsoon;  $59.61 \pm 1.38$  and  $60.31 \pm 1.39$  in winter seasons respectively. Protein with standard deviation per mg dry weight of whole body soft tissue in *Lamellidens marginali* bivalve samples collected from Sirpur Lake site A and site B were  $48.51 \pm 1.14$  and  $49.35 \pm 1.29$  in summer;  $58.67 \pm 1.27$  and  $60.01 \pm 1.37$  in monsoon;  $53.24 \pm 1.11$  and  $56.24 \pm 1.34$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $49.23 \pm 1.29$  and  $50.01 \pm 1.43$  in summer;  $60.03 \pm 1.27$  and  $61.20 \pm 1.31$  in monsoon;  $57.23 \pm 1.36$  and  $59.64 \pm 1.32$  in winter seasons respectively.

Fig. 55. Total protein in gills, digestive gland and whole soft body of *Lamellidens marginali* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.

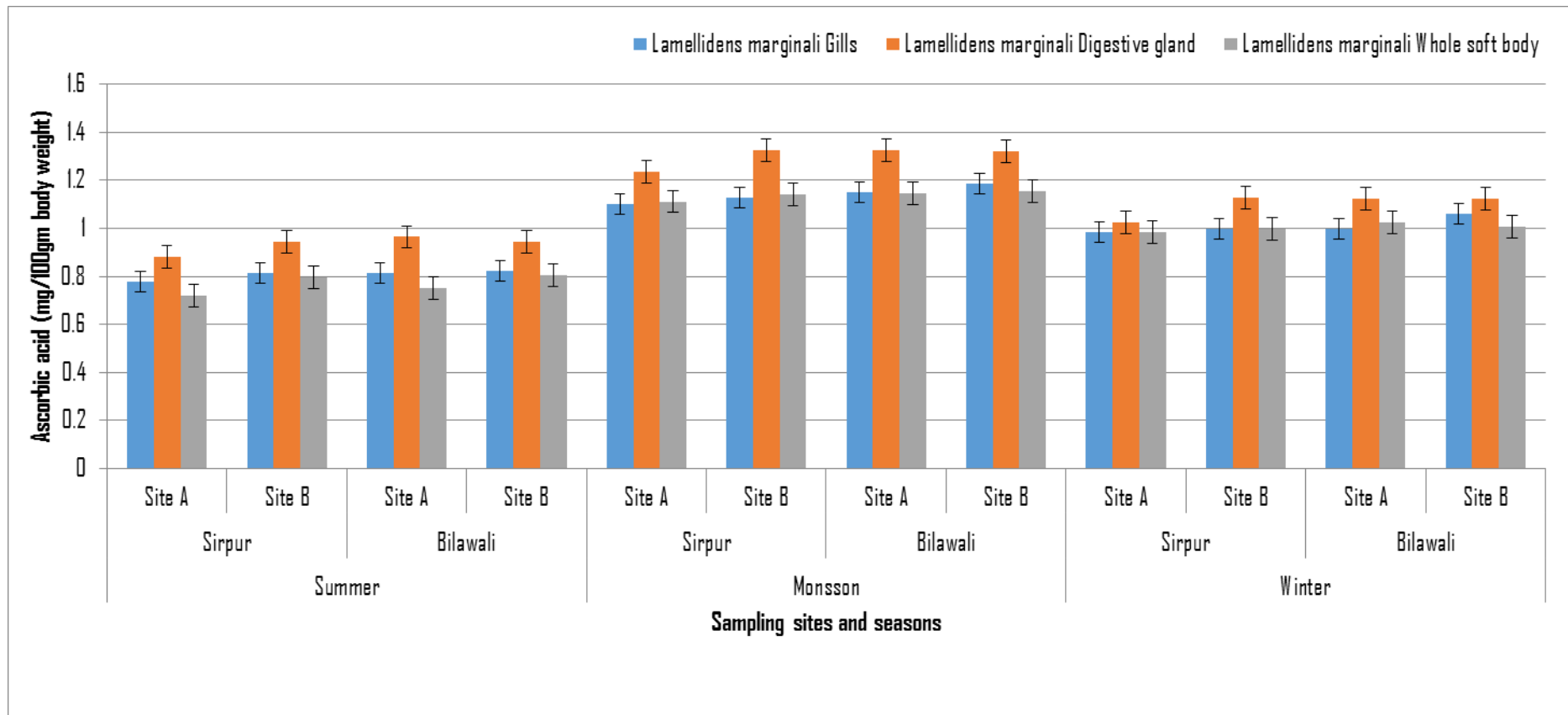




### 3.5.1.2 Ascorbic acid

Ascorbic acid with standard error per mg dry weight of gills in collected *Lamellidens marginali* bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 11. Ascorbic acid with standard deviation per mg dry weight of gills in *Lamellidens marginali* bivalve samples collected from Sirpur Lake site A and site B were  $0.712\pm 0.001$  and  $0.786\pm 0.003$  in summer;  $1.005\pm 0.005$  and  $1.132\pm 0.009$  in monsoon;  $0.986\pm 0.009$  and  $1.002\pm 0.009$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $0.805\pm 0.001$  and  $0.814\pm 0.001$  in summer;  $1.142\pm 0.011$  and  $1.112\pm 0.009$  in monsoon;  $1.009\pm 0.008$  and  $1.008\pm 0.009$  in winter seasons respectively. Ascorbic acid with standard deviation per mg dry weight of digestive gland in *Lamellidens marginali* bivalve samples collected from Sirpur Lake site A and site B were  $0.856\pm 0.002$  and  $0.886\pm 0.003$  in summer;  $1.103\pm 0.005$  and  $1.296\pm 0.010$  in monsoon;  $1.023\pm 0.009$  and  $1.124\pm 0.010$  in winter seasons whereas, Bilawali Tank site A and site B

Fig. 56. Ascorbic acid in gills, digestive gland and whole soft body of *Lamellidens marginali* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.



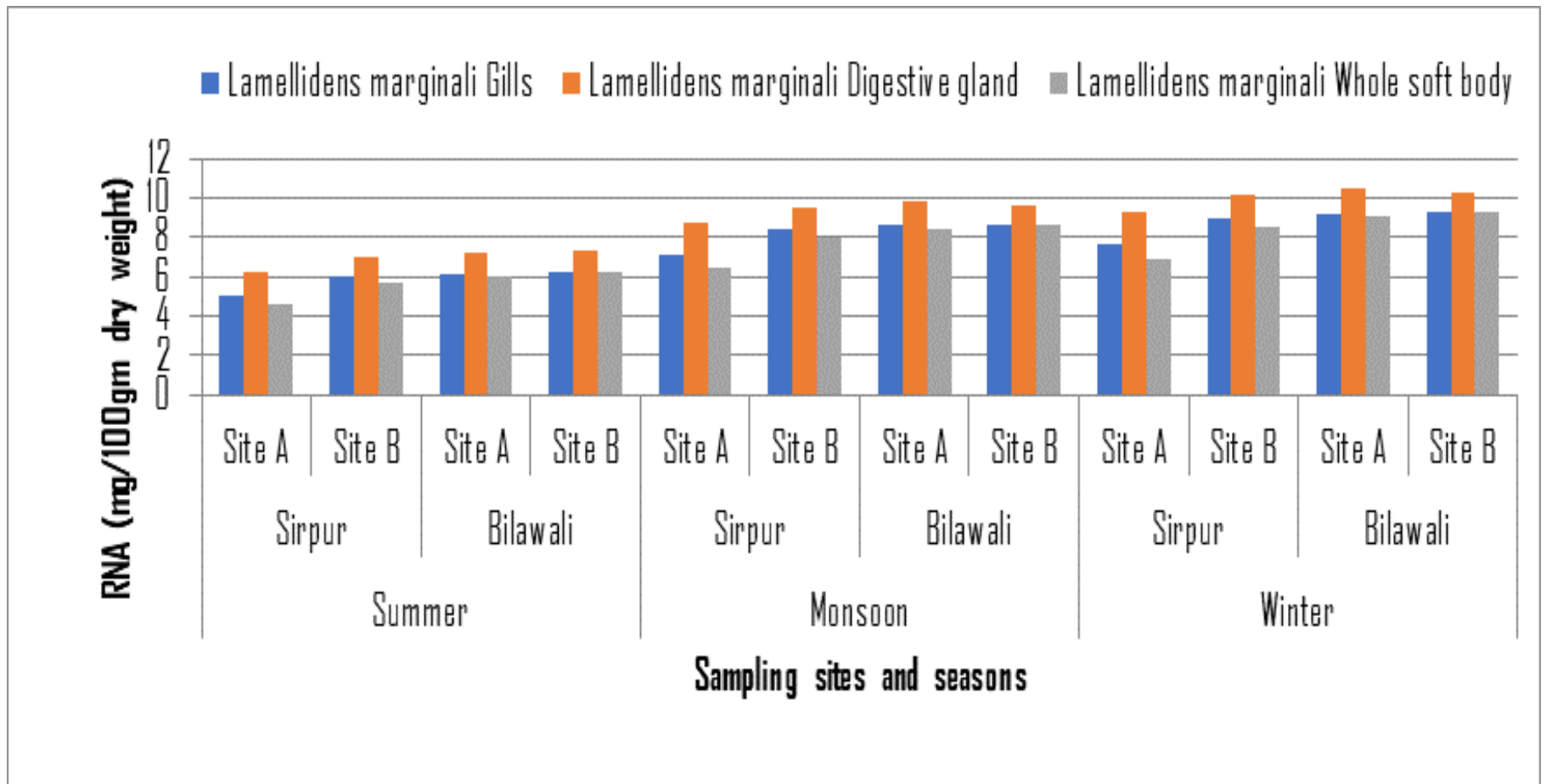
were reported  $0.926\pm 0.002$  and  $0.953\pm 0.002$  in summer;  $1.298\pm 0.012$  and  $1.236\pm 0.010$  in monsoon;  $1.143\pm 0.010$  and  $1.112\pm 0.008$  in winter seasons respectively. Ascorbic acid with standard deviation per mg dry weight of whole body soft tissue in *Lamellidens marginali* bivalve samples collected from Sirpur Lake site A and site B were  $0.702\pm 0.001$  and  $0.736\pm 0.002$  in summer;  $1.009\pm 0.004$  and  $1.124\pm 0.010$  in monsoon;  $0.958\pm 0.008$  and  $0.995\pm 0.009$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $0.792\pm 0.001$  and  $0.756\pm 0.001$  in summer;  $1.136\pm 0.009$  and  $1.152\pm 0.012$  in monsoon;  $0.995\pm 0.009$  and  $1.020\pm 0.008$  in winter seasons respectively.

### 3.5.1.3 RNA

RNA with standard error per mg dry weight of gills in collected *Lamellidens marginali* bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 11. RNA with standard deviation per mg dry weight of gills in *Lamellidens marginali* bivalve samples collected from Sirpur Lake site A and site B were  $5.07\pm 0.28$  and  $6.01\pm 0.38$  in summer;  $7.09\pm 0.37$  and  $8.40\pm 0.39$  in monsoon;  $7.59\pm 0.33$  and  $9.00\pm 0.41$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $6.13\pm 0.34$  and  $6.12\pm 0.30$  in summer;  $8.58\pm 0.44$  and  $8.66\pm 0.33$  in monsoon;  $9.17\pm 0.42$  and  $9.28\pm 0.43$  in winter seasons respectively. RNA with standard deviation per mg dry weight of digestive gland in *Lamellidens marginali* bivalve samples collected from

Sirpur Lake site A and site B were  $6.22 \pm 0.33$  and  $7.03 \pm 0.24$  in summer;  $8.70 \pm 0.39$  and  $9.49 \pm 0.31$  in monsoon;  $9.31 \pm 0.37$  and  $9.43 \pm 0.46$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $7.20 \pm 0.41$  and  $7.31 \pm 0.27$  in summer;  $9.81 \pm 0.45$  and  $9.59 \pm 0.37$  in monsoon;  $9.92 \pm 0.62$  and  $9.63 \pm 0.53$  in winter seasons respectively. RNA with standard deviation per mg dry weight of whole body soft tissue in *Lamellidens marginali* bivalve samples collected from Sirpur Lake site A and site B were  $4.59 \pm 0.21$  and  $5.71 \pm 0.39$  in summer;  $6.42 \pm 0.27$  and  $7.98 \pm 0.33$  in monsoon;  $6.87 \pm 0.38$  and  $8.55 \pm 0.46$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $6.04 \pm 0.38$  and  $6.22 \pm 0.35$  in summer;  $8.45 \pm 0.39$  and  $8.63 \pm 0.41$  in monsoon;  $9.03 \pm 0.39$  and  $9.25 \pm 0.35$  in winter seasons respectively

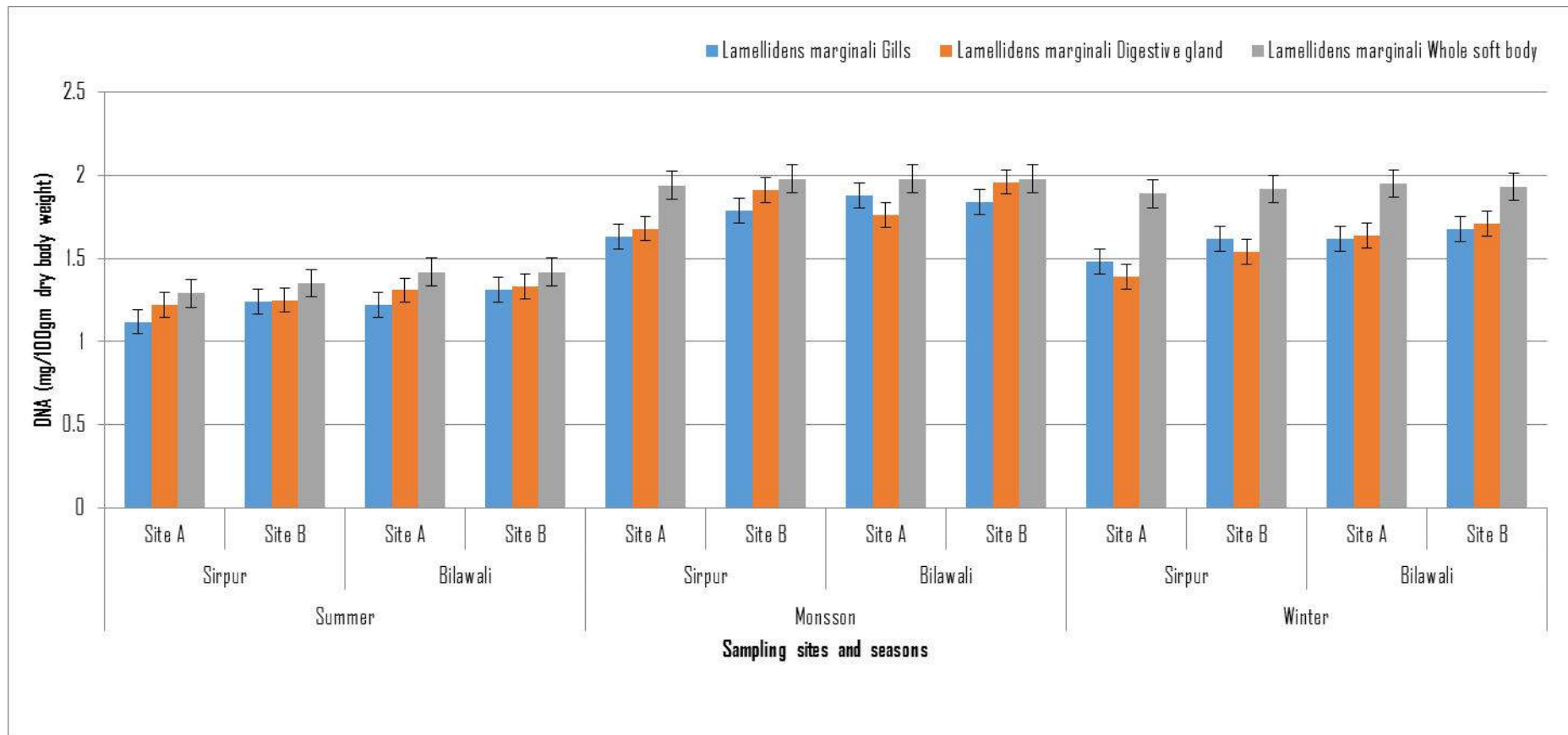
Fig. 57. RNA in gills, digestive gland and whole soft body of *Lamellidens marginali* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons



#### 3.5.1.4 DNA

DNA with standard error per mg dry weight of gills in collected *Lamellidens marginali* bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 11. DNA with standard deviation per mg dry weight of gills in *Lamellidens marginali* bivalve samples collected from Sirpur Lake site A and site B were  $1.12\pm 0.23$  and  $1.18\pm 0.22$  in summer;  $1.46\pm 0.23$  and  $1.36\pm 0.21$  in monsoon;  $1.62\pm 0.12$  and  $1.54\pm 0.21$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $1.23\pm 0.21$  and  $1.29\pm 0.21$  in summer;  $1.76\pm 0.22$  and  $1.86\pm 0.23$  in monsoon;  $1.56\pm 0.22$  and  $1.59\pm 0.23$  in winter seasons respectively. DNA with standard deviation per mg dry weight of digestive gland in *Lamellidens marginali* bivalve

Fig. 58. DNA in gills, digestive gland and whole soft body of *Lamellidens marginali* bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.



samples collected from Sirpur Lake site A and site B were  $1.19\pm 0.20$  and  $1.25\pm 0.19$  in summer;  $1.79\pm 0.23$  and  $1.85\pm 0.22$  in monsoon;  $1.63\pm 0.17$  and  $1.59\pm 0.22$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $1.31\pm 0.22$  and  $1.35\pm 0.23$  in summer;  $1.89\pm 0.23$  and  $1.97\pm 0.22$  in monsoon;  $1.62\pm 0.23$  and  $1.68\pm 0.22$  in winter seasons respectively. DNA with standard deviation per mg dry weight of whole body soft tissue in *Lamellidens marginali* bivalve samples collected from Sirpur Lake site A and site B were  $1.23\pm 0.21$  and  $1.32\pm 0.14$  in summer;  $1.94\pm 0.25$  and  $1.99\pm 0.23$  in monsoon;  $1.79\pm 0.21$  and  $1.86\pm 0.23$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $1.41\pm 0.23$  and  $1.42\pm 0.21$  in summer;  $1.95\pm 0.23$  and  $1.96\pm 0.21$  in monsoon;  $1.95\pm 0.22$  and  $1.94\pm 0.23$  in winter seasons respectively.



### **3.5.3 *Unio* sp.**

Table 12 presents the mean with standard deviation of collected observations of biochemical studies in *Unio* sp. samples.

Table 12. Seasonal variations of biochemical parameters in *Unio* sp. collected from Sirpur and Bilawali Tanks of Indore.

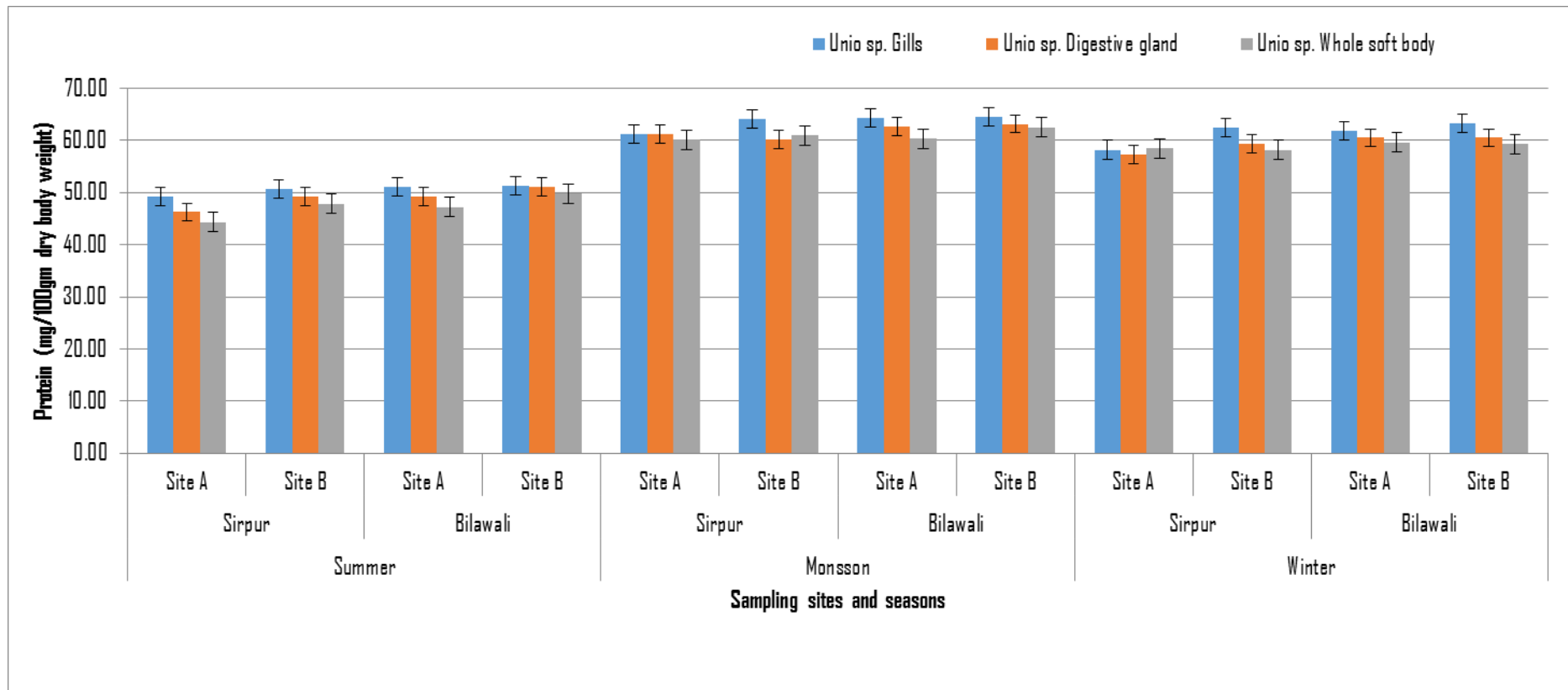
Body part	Summer				Monson				Winter			
	Sirpur		Bilawali		Sirpur		Bilawali		Sirpur		Bilawali	
	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B
<b>PROTEIN</b>												
<b>Gills</b>	49.21±1. 12	50.67±1. 62	51.02±1. 42	51.28±1. 21	61.24±1. 60	64.16±1. 29	64.23±1. 19	64.57±1. 29	58.21±1. 12	62.38±1. 39	61.85±1. 29	63.24±1. 27
<b>Digestive gland</b>	46.24±1. 28	49.16±1. 52	49.26±1. 39	51.09±1. 12	61.25±1. 18	60.25±1. 32	62.58±1. 32	63.16±1. 32	57.31±1. 14	59.42±1. 37	60.49±1. 21	60.49±1. 24
<b>Whole soft body</b>	44.32±1. 13	47.86±1. 28	47.21±1. 38	49.75±1. 13	60.16±1. 38	60.95±1. 27	60.28±1. 19	62.54±1. 33	58.42±1. 15	58.12±1. 34	59.64±1. 23	59.25±1. 11
<b>ASCORBIC ACID</b>												
<b>Gills</b>	0.774±0. 001	0.812±0. 003	0.812±0. 002	0.818±0. 001	1.110±0. 006	1.119±0. 009	1.136±0. 012	1.169±0. 011	0.916±0. 008	1.012±0. 010	1.041±0. 010	1.004±0. 008
<b>Digestive gland</b>	0.920±0. 003	0.912±0. 003	0.965±0. 002	0.924±0. 002	1.254±0. 006	1.321±0. 011	1.321±0. 014	1.325±0. 015	0.998±0. 007	1.109±0. 010	1.116±0. 009	1.128±0. 008
<b>Whole soft body</b>	0.719±0. 002	0.795±0. 002	0.789±0. 002	0.795±0. 002	1.110±0. 006	1.294±0. 011	1.142±0. 015	1.176±0. 015	0.937±0. 006	0.994±0. 009	0.994±0. 008	1.012±0. 006
<b>DNA</b>												
<b>Gills</b>	1.16±0.1 9	1.17±0.1 9	1.21±0.2 1	1.26±0.1 6	1.43±0.1 9	1.65±0.2 0	1.82±0.2 2	1.78±0.1 7	1.42±0.2 1	1.46±0.1 9	1.65±0.2 1	1.69±0.2 4
<b>Digestive gland</b>	1.14±0.2 3	1.20±0.1 7	1.24±0.2 2	1.29±0.2 1	1.62±0.2 8	1.79±0.2 1	1.86±0.2 1	1.95±0.1 9	1.41±0.1 8	1.47±0.2 4	1.58±0.2 1	1.68±0.2 4
<b>Whole soft body</b>	1.25±0.2 1	1.31±0.2 2	1.39±0.2 3	1.39±0.1 7	1.98±0.2 8	1.99±0.2 0	1.84±0.2 1	1.97±0.1 9	1.95±0.1 9	1.95±0.2 5	1.89±0.2 0	1.93±0.2 9

RNA												
Body part	Summer				Monsoon				Winter			
	Sirpur		Bilawali		Sirpur		Bilawali		Sirpur		Bilawali	
	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B	Site A	Site B
<b>Gills</b>	5.02± 0.33	5.43±0.34	5.61± 0.34	5.98± 0.36	7.02± 0.39	7.59± 0.37	7.85± 0.39	8.30± 0.41	7.51± 0.34	8.13± 0.43	8.39± 0.40	8.89± 0.39
<b>Digestive gland</b>	6.13± 0.39	6.03±0.29	6.14± 0.29	6.25± 0.34	8.58± 0.41	8.42± 0.34	8.59± 0.41	8.68± 0.43	9.18± 0.39	9.03± 0.39	9.18± 0.39	9.30± 0.37
<b>Whole soft body</b>	4.87± 0.27	5.37±0.34	5.47± 0.24	5.84± 0.32	6.81± 0.39	7.50± 0.32	7.65± 0.39	8.11± 0.37	7.29± 0.46	8.04± 0.38	8.18± 0.46	8.69± 0.39

### 3.5.3.1 Protein

Protein with standard error per mg dry weight of gills in collected *Unio* sp. bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 12. Protein with standard deviation per mg dry weight of gills in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $49.21 \pm 1.12$  and  $50.67 \pm 1.62$  in summer;  $61.24 \pm 1.60$  and  $64.16 \pm 1.29$  in monsoon;  $58.21 \pm 1.12$  and  $62.38 \pm 1.39$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $51.02 \pm 1.42$  and  $51.28 \pm 1.21$  in summer;  $64.23 \pm 1.19$  and  $64.57 \pm 1.29$  in monsoon;  $61.85 \pm 1.29$  and  $63.24 \pm 1.27$  in winter seasons respectively. Protein with standard deviation per mg dry weight of digestive gland in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $46.24 \pm 1.28$  and  $49.16 \pm 1.52$  in summer;  $61.25 \pm 1.18$  and  $60.25 \pm 1.32$  in monsoon;  $57.31 \pm 1.14$  and  $59.42 \pm 1.37$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $49.26 \pm 1.39$  and  $51.09 \pm 1.12$  in summer;  $62.58 \pm 1.32$  and  $63.16 \pm 1.32$  in monsoon;  $60.49 \pm 1.21$  and  $60.49 \pm 1.24$  in winter seasons respectively. Protein with standard deviation per mg dry weight of whole body soft tissue in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $44.32 \pm 1.13$  and  $47.86 \pm 1.28$  in summer;  $60.16 \pm 1.38$  and  $60.95 \pm 1.27$  in monsoon;  $58.42 \pm 1.15$  and  $58.12 \pm 1.34$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $47.21 \pm 1.38$  and  $49.75 \pm 1.13$  in summer;  $60.28 \pm 1.19$  and  $62.54 \pm 1.33$  in monsoon;  $59.64 \pm 1.23$  and  $59.25 \pm 1.11$  in winter seasons respectively.

Fig. 59. Total protein in gills, digestive gland and whole soft body of *Unio* sp. bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.

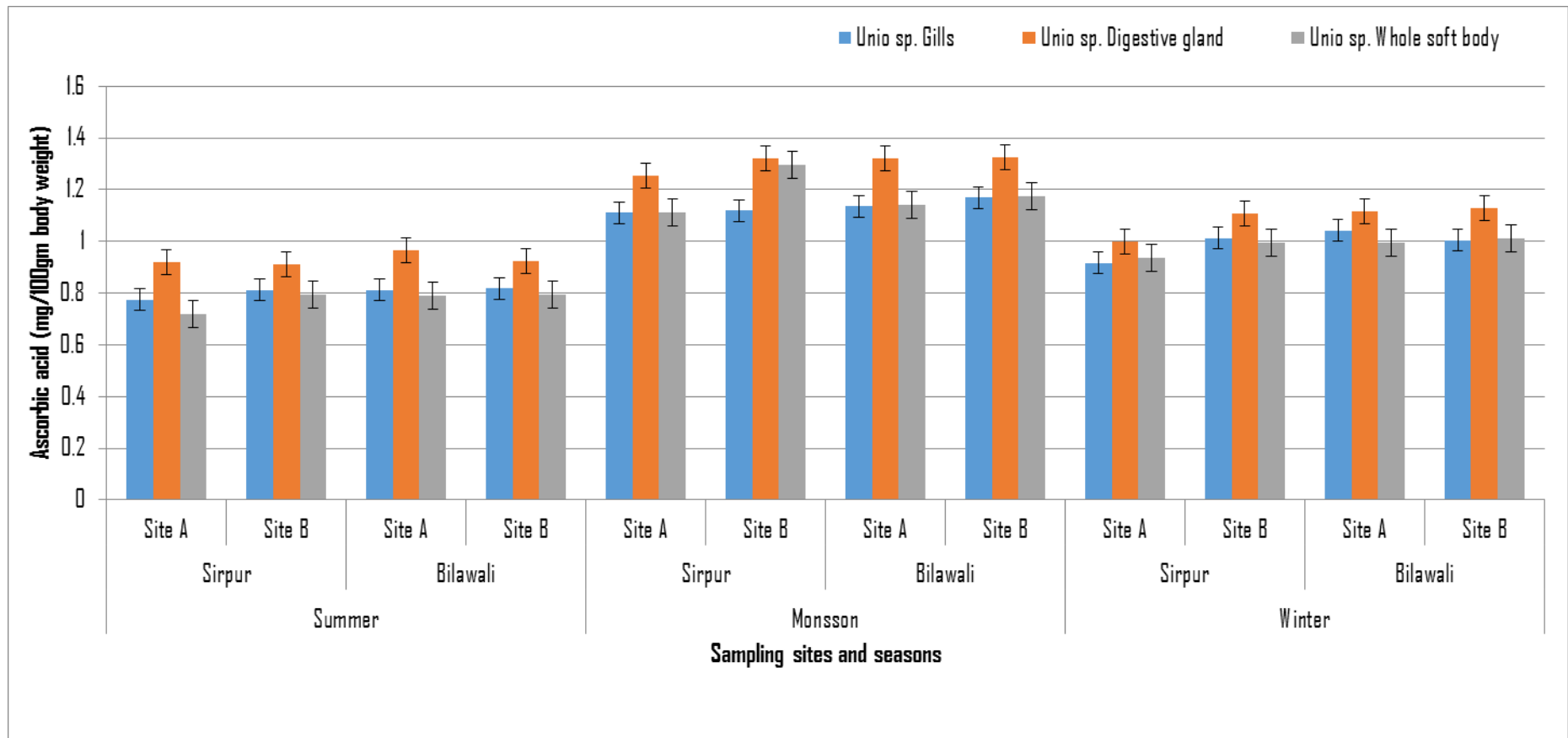


### 3.5.1.2 Ascorbic acid

Ascorbic acid with standard error per mg dry weight of gills in collected *Unio* sp. bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 12. Ascorbic acid with standard deviation per mg dry weight of gills in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $0.774 \pm 0.001$  and  $0.812 \pm 0.003$  in summer;  $1.110 \pm 0.006$  and  $1.119 \pm 0.009$  in monsoon;  $0.916 \pm 0.008$  and  $1.012 \pm 0.010$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $0.812 \pm 0.002$  and  $0.818 \pm 0.001$  in summer;  $1.136 \pm 0.012$  and  $1.169 \pm 0.011$  in monsoon;  $1.041 \pm 0.010$  and  $1.004 \pm 0.008$  in winter seasons respectively. Ascorbic acid with standard deviation per mg dry weight of digestive gland in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $0.920 \pm 0.003$  and  $0.912 \pm 0.003$  in summer;  $1.254 \pm 0.006$  and  $1.321 \pm 0.011$  in monsoon;  $0.998 \pm 0.007$  and  $1.109 \pm 0.010$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $0.965 \pm 0.002$  and  $0.924 \pm 0.002$  in summer;  $1.321 \pm 0.014$  and  $1.325 \pm 0.015$  in monsoon;  $1.116 \pm 0.009$  and  $1.128 \pm 0.008$  in winter seasons respectively. Ascorbic acid with standard deviation per mg dry weight of whole body soft tissue in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $0.719 \pm 0.002$  and  $0.795 \pm 0.002$  in summer;  $1.110 \pm 0.006$  and  $1.294 \pm 0.011$  in monsoon;  $0.937 \pm 0.006$  and  $0.994 \pm 0.009$  in winter seasons whereas,

Bilawali Tank site A and site B were reported  $0.789\pm 0.002$  and  $0.795\pm 0.002$  in summer;  $1.142\pm 0.015$  and  $1.176\pm 0.015$  in monsoon;  $0.994\pm 0.008$  and  $1.012\pm 0.006$  in winter seasons respectively.

Fig. 60. Ascorbic acid in gills, digestive gland and whole soft body of *Unio* sp. bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.

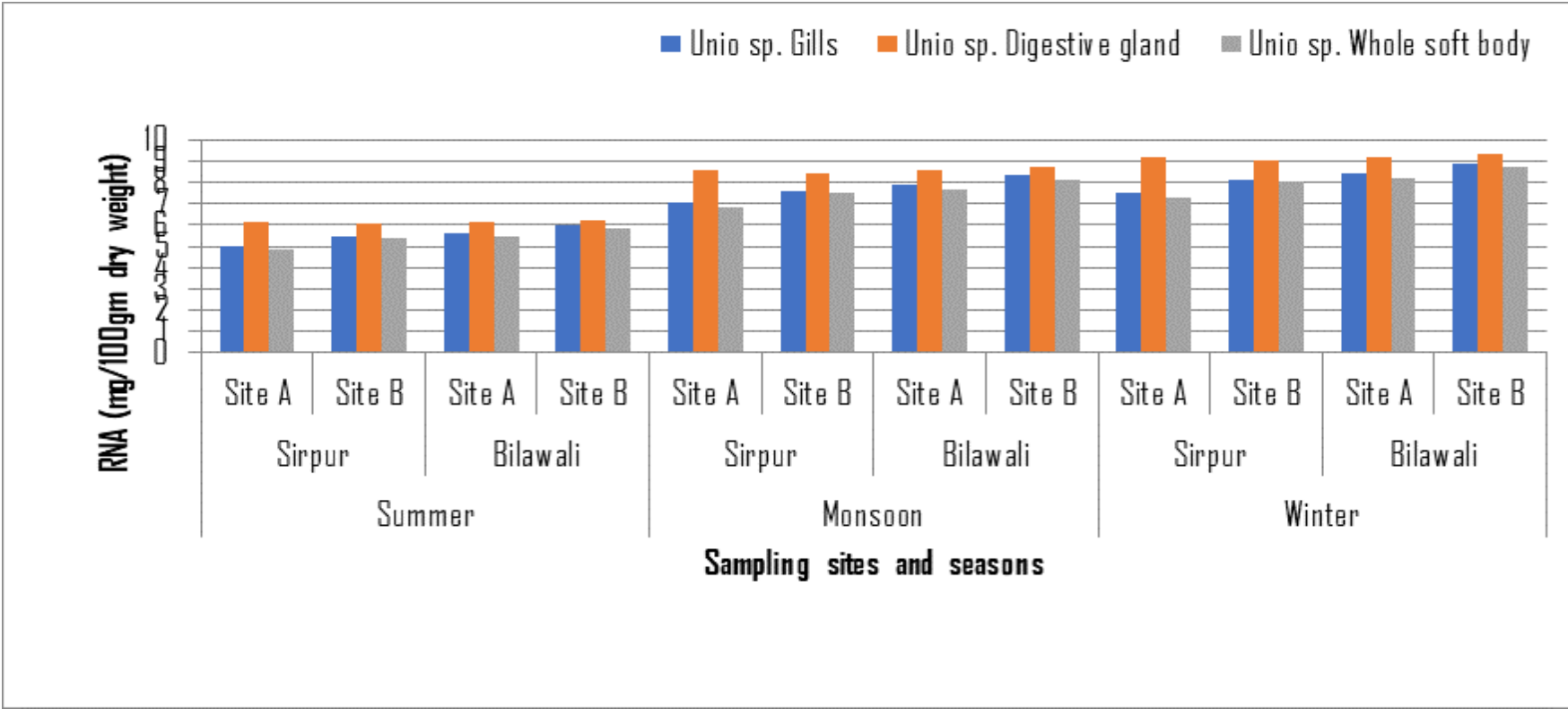




### 3.5.1.3 RNA

RNA with standard error per mg dry weight of gills in collected *Unio* sp. bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 12. RNA with standard deviation per mg dry weight of gills in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $5.02 \pm 0.33$  and  $5.43 \pm 0.34$  in summer;  $7.02 \pm 0.39$  and  $7.59 \pm 0.37$  in monsoon;  $7.51 \pm 0.34$  and  $8.13 \pm 0.43$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $5.61 \pm 0.34$  and  $5.98 \pm 0.36$  in summer;  $7.85 \pm 0.39$  and  $8.30 \pm 0.41$  in monsoon;  $8.39 \pm 0.40$  and  $8.89 \pm 0.39$  in winter seasons respectively. RNA with standard deviation per mg dry weight of digestive gland in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $6.13 \pm 0.39$  and  $6.03 \pm 0.29$  in summer;  $8.58 \pm 0.41$  and  $8.42 \pm 0.34$  in monsoon;  $9.18 \pm 0.39$  and  $19.03 \pm 0.39$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $6.14 \pm 0.29$  and  $6.25 \pm 0.34$  in summer;  $8.59 \pm 0.41$  and  $8.68 \pm 0.43$  in monsoon;  $9.18 \pm 0.39$  and  $9.30 \pm 0.37$  in winter seasons respectively. RNA with standard deviation per mg dry weight of whole body soft tissue in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $4.87 \pm 0.27$  and  $5.37 \pm 0.34$  in summer;  $6.81 \pm 0.39$  and  $7.50 \pm 0.32$  in monsoon;  $7.29 \pm 0.46$  and  $8.04 \pm 0.38$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $5.47 \pm 0.24$  and  $5.84 \pm 0.32$  in summer;  $7.65 \pm 0.39$  and  $8.11 \pm 0.37$  in monsoon;  $8.18 \pm 0.46$  and  $8.69 \pm 0.39$  in winter seasons respectively.

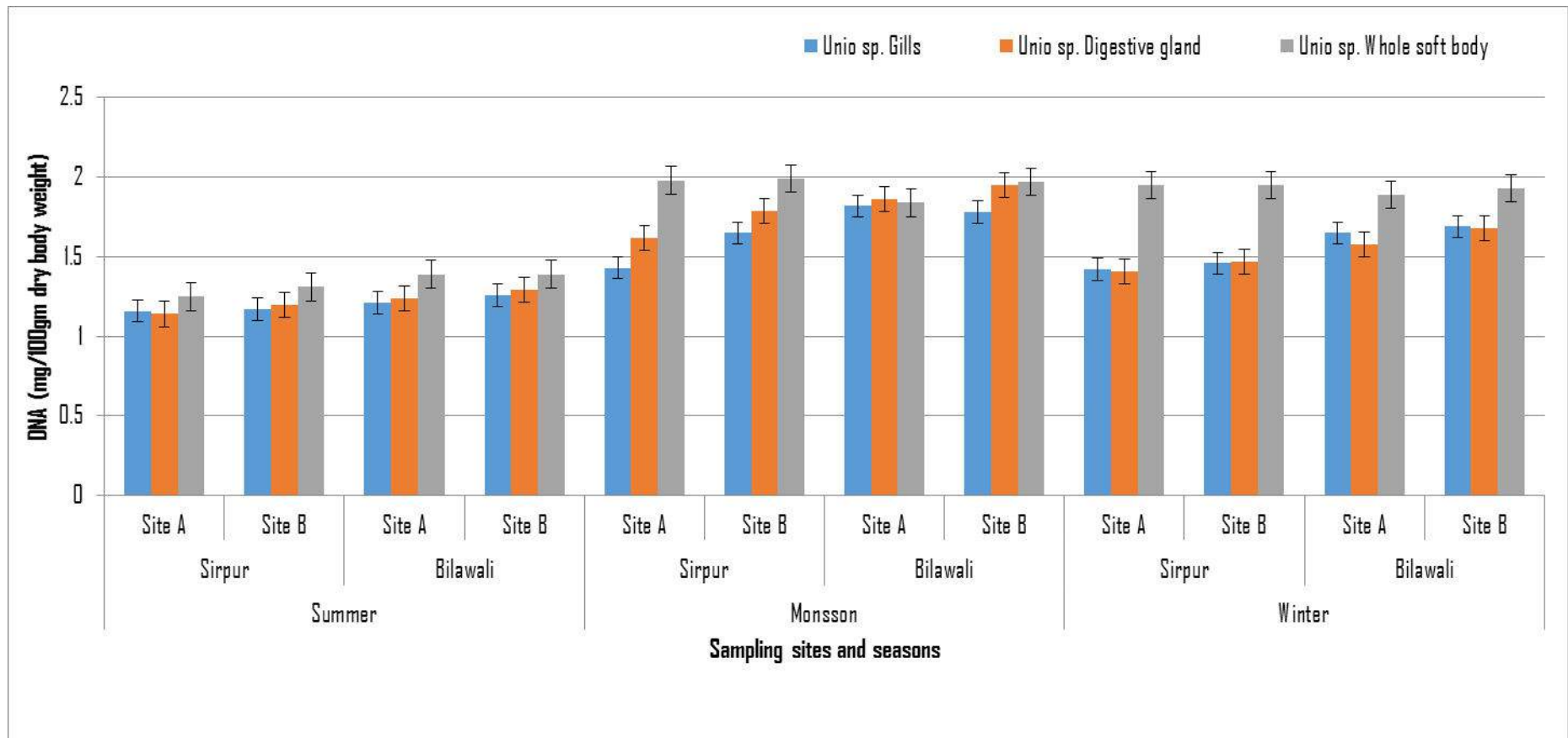
Fig. 61. RNA in gills, digestive gland and whole soft body of *Unio* sp. bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.



#### 3.5.1.3.4 DNA

DNA with standard error per mg dry weight of gills in collected *Unio* sp. bivalve samples of Sirpur and Bilawali Tank in summer, monsoon and winter seasons is shown in Table 12. DNA with standard deviation per mg dry weight of gills in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $1.16 \pm 0.19$  and  $1.17 \pm 0.19$  in summer;  $1.43 \pm 0.19$  and  $1.65 \pm 0.20$  in monsoon;  $1.42 \pm 0.21$  and  $1.46 \pm 0.19$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $1.21 \pm 0.21$  and  $1.26 \pm 0.16$  in summer;  $1.82 \pm 0.22$  and  $1.78 \pm 0.17$  in monsoon;  $1.65 \pm 0.21$  and  $1.69 \pm 0.24$  in winter seasons respectively. DNA with standard deviation per mg dry weight of digestive gland in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $1.14 \pm 0.23$  and  $1.20 \pm 0.17$  in summer;  $1.62 \pm 0.28$  and  $1.79 \pm 0.21$  in monsoon;  $1.41 \pm 0.18$  and  $1.47 \pm 0.24$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $1.24 \pm 0.22$  and  $1.29 \pm 0.21$  in summer;  $1.86 \pm 0.21$  and  $1.95 \pm 0.19$  in monsoon;  $1.58 \pm 0.21$  and  $1.68 \pm 0.24$  in winter seasons respectively. DNA with standard deviation per mg dry weight of whole body soft tissue in *Unio* sp. bivalve samples collected from Sirpur Lake site A and site B were  $1.25 \pm 0.21$  and  $1.31 \pm 0.22$  in summer;  $1.98 \pm 0.28$  and  $1.99 \pm 0.20$  in monsoon;  $1.95 \pm 0.19$  and  $1.95 \pm 0.25$  in winter seasons whereas, Bilawali Tank site A and site B were reported  $1.39 \pm 0.23$  and  $1.39 \pm 0.17$  in summer;  $1.84 \pm 0.21$  and  $1.97 \pm 0.19$  in monsoon;  $1.89 \pm 0.20$  and  $1.93 \pm 0.29$  in winter seasons respectively.

Fig. 62. DNA in gills, digestive gland and whole soft body of *Unio* sp. bivalves collected from different sampling sites of Sirpur and Bilawali Tanks of Indore in different seasons.



# *DISCUSSION*

## DISCUSSION

Physico-chemical analysis of surface water and heavy metal determination in water and soil sediments of Sirpur and Bilawali Tanks of Indore was studied. Dry weight whole body; metal concentration per unit body weight ( $\mu\text{g/g}$ ); metal Body burden ( $\mu\text{g/individual}$ ); BWAF (Bio-water accumulation factor); BSAF (Bio-sediment accumulation factor); protein content, ascorbic acid content and DNA content in gills, digestive gland and whole soft body in of three species of bivalves viz. *Lamellidens corrianus*, *Lamellidens marginali* and *Unio* sp. collected from Sirpur Lake and Bilawali Tank, Indore were studied to determine effect of pollution on such species in this study and discuss with previous studies.

### **Physico-chemical studies of surface water samples**

Quality water is needed to fulfill the need of human settlement. This is required for domestic purpose, drinking, fishing, irrigation and industries (Pruss, 1998; Thornburn *et al.*, 2003). India has 15 different ecological regions where 1.42 million population lives. About 37.7 million populations of this country are affected by waterborne diseases annually, in which approximately 1.5 million children suffer with diarrhea and allied disease every year. This is results nearly 600 million dollar burden to Indian economy. Over exploitation and pollution are main causes of deterioration of water quality (Pandey *et al.*, 2003). Polluted water quality may affect the aquatic life, animals, plants and humans (Chang *et al.*, 1998; Kara and Comlekci, 2004). A regular monitoring

of water quality is needed to maintain the quality of water. Physico-chemical parameters, heavy metal assessment and bioaccumulation studies will needed regular assessment of water quality. Water qualities, heavy metals assessment of water, sediment and bioaccumulation studies in bivalves of Sirpur and Bilawali Tank of Indore Madhya Pradesh in different seasons were studied. Results have shown the higher values of Total alkalinity (mg/l) in Sirpur Lake site A during summer season followed by Sirpur site B, Bilawali site A, Bilawali site B in summer; Sirpur site A, site B, Bilawali site A site B in monsoon and winter seasons. Higher values of Dissolved oxygen (mg/l) in Bilawali Tank site A during winter season and reported gradually decreased values in Sirpur site B, site A, Bilawali site B in winter; Bilawali site B, site A Sirpur site B, site A in monsoon and summer seasons. Highest total hardness (mg/l) was recorded in Sirpur site A and then Sirpur site B, Bilawali site A, site B in summer season. Gradually decreased values were observed in Sirpur site A, site B, Bilawali site A, site B in winter season; Sirpur site A, site B, Bilawali site A, site B in monsoon season. Highest Salinity (mg/l) was recorded in Sirpur site A and then Sirpur site B, Bilawali site A, site B in summer season. Gradually decreased values were observed in Sirpur site A, site B, Bilawali site A, site B in winter season; Sirpur site A, site B, Bilawali site A, site B in monsoon season. Highest Chloride (mg/l) was recorded in Sirpur site A and then Sirpur site B, Bilawali site A, site B in summer season. Gradually decreased values were observed in Sirpur site A, site B, Bilawali site A, site B in winter season; Sirpur site A, site B, Bilawali site A, site B in monsoon

season. Temperature was recorded within normal range everywhere. Highest pH (unit) was recorded in Sirpur site A in every season in comparison to other sampling sites. This result clearly indicated that Site A (Sirpur lake) was more polluted than Bilawali Tank sites and Sirpur site B. Obtained results are statistically proven with  $P < 0.05$ . The higher pH, total hardness, total alkalinity, chlorides, salinity, and lowest dissolved oxygen levels were found in Site A surface water (Sirpur lake) may be due to huge agricultural runoff, domestic waste, textile, and other manufacturing residues. Catchment areas of Sirpur Lake Site A are surrounded by agricultural field, which is polluted by different fertilizers and pesticides. Earlier report told the high pH values were recorded, may be due to heavy input of textile, printing and other effluent with municipal waste (Kalff and Knoechel, 1998). Richardson (1988) worked on pollution of water and reported heavy contamination in water, which alter the physico-chemical properties of water. Dehadri (1990) also reported the direct discharge of industrial effluents deplete different characters of water viz. dissolved oxygen, pH, increase the  $\text{CO}_2$  level in the water. Kolhe *et al.*, (2014) also told a story of pH modification after fusion of household and byproducts in water. In this analysis, higher pH was observed during the summer season than in the winter and monsoon seasons. This can be due to increased photosynthetic absorption of inorganic carbon dissolved by planktons (Subba rao and Govind, 1967; Goldman, 1972; Farrell *et al.*, 1979). Sharma and Jain (2000) recorded low pH in winter as well as higher in summer season. Mishra and Tripathi (2001); Singh *et al.*, (2002) also supported the results of like reports. The high



pH recorded for Venkatesharaju (2010) in winter is due to a decreased rate of decomposition.

### **Heavy Metal Concentrations in sediment and surface water samples**

Assessments of heavy metals in surface water samples and sediment of two different sites of Sirpur and Bilawali Tanks of Indore were performed and Median concentrations recorded of Zn, Cu, Pb Cd and Mn in surface water of Sirpur Site A and Site B were higher than site A and B of Bilawali Tank in summer and gradually lower in similar manner in winter and monsoon. The main reason behind the pollution of Site A (Sirpur lake), is catchment areas of surrounded by agricultural field, which is polluted by fertilizers and pesticides, these could acts as source of heavy metals Cu, Mn, Pb, Zn and As (Verkleji, 1993; de Meeus *et al.*, 2002). Discharges of untreated household waste and water into water bodies are a major cause of heavy metal contamination (Hadjmohammad, 1988; Rashed, 2001; Ravera, 2004; Aksoy *et al.*, 2005). Abaychi and DouAbul (1985) Stated that treated and untreated urban, industrial waste, agricultural run-off leads to the emissions from heavy metals Cu, Pb, Cd and Zn. Lokhande et al., (2011) stated that the major industries contributing to the contamination of Mn, Pb and Cu in the aquatic environment are colours, paints, textile industries. Jaishree and Khan (2014) claimed that textile dyeing and printing wastewater effluents include colours, bleaching agents, salts, acids, and heavy metals such as Cr, Cu, Pb, and Zn. The results show that the concentrations of metals in sediments varied widely and show variations in

values of Mn, Cu, Pb, Cd and Zn between different reservoirs. The results showed a concentration pattern close to that of its abundance in water . Sirpur Lake Site A carries enormous amounts of domestic waste, garment, printing, industrial dyeing and other effluent together with organic matter, run-off from agricultural waste and traffic run-off, which is driven by higher amounts of Mn, Cu, Pb ,Mn and Zn heavy metals in sediment samples collected from Site B. Halcrow et al., (1973) reported earlier that concentrations of heavy metals in sediments increased. Presley et al. (1980) reported that the elemental sediment concentrations depend not only on anthropogenic sources, but also on the quality of organic matter, textural characteristics, mineralogical composition and sediment deposition environment. Harland et al., (2000) researched and documented the reliance on organic matter and particle size of metal concentrations in the sediments. Accumulation of heavy metals in sediments may affect concentrations of heavy metals in aquatic organisms living in these sediments (Pourang, 1996; Wildi *et al.*, 2004; Kim and Kim, 2006). Overall results showed that highest concentration of heavy metals Mn, Cu, Pb, and Zn in sediments samples of Sirpur Lake Site A than Sirpur Lake Site B, Bilawali Tank Site A and Bilawali Tank Site. However, tracing Hg in both water and soil sediments was a cumbersome process with high degree of error.

### **Bioaccumulation Study**

Concentrations of heavy metals viz. Mn, Cu, Pb, Cd and Zn metal body burden, BWAf and BSAF values were determined in dry weight of whole soft body

tissues of three different freshwater bivalve species i.e. *Lamellidens corrianus*, *Lamellidens marginalis* and *Unio.sp.* collected from Sirpur Lake Site A, Site B; Bilawali Tank Site A, Site B of Indore Madhya Pradesh during monsoon, summer and winter seasons. Results indicate that mean highest values of concentrations of heavy metals in body, metal body burden and BWAF and BSAF in dry soft body tissues of all sampled bivalve species collected from Sirpur Lake Site A than Sirpur Lake Site B, Bilawali tank Site A and Bilawali tank Site B. Reported higher metal concentrations and body burden in whole dry soft body tissues of all sampled bivalve species collected from Sirpur Lake Site A might be due to exposure of higher pollutant than other sampling sites. Shinde (2013) reported in his study that metal concentrations in the dry soft body tissues of mollusc were related to metal levels present in water bodies and soil. Deshmukh (2013) also supported this report. It includes the BWAF / BSAF values used to identify the most suitable sentinel species to track heavy metal contamination in water. In this study BWAF values refer to the concentration of a specific metal in bivalve tissues per concentration of that metal in water. Biosediment Accumulation Factor is specified as the metal concentration ratio of the bivalve tissue to that in the sediment (Usero *et al.*, 2005; Szefer *et al.*, 1999). In moonsoon, the concentrations of heavy metals, metal body load, BWAF and BSAF throughout all bivalve species were low compared to summer and winter seasons, this could be due to the rise in water levels in water bodies (Patil *et al.*, 2004; Deshmukh, 2013).

## **Oxidative Stress**

Antioxidant enzyme activity: viz. SOD, CAT, GPx and GST; levels of GSH and LPO of three different freshwater bivalve species were assessed in the dry weight of the digestive gland tissues i.e. *Lamellidens corrianus*, *Lamellidens marginalis* and *Unio. sp.* collected from Sirpur Lake Site A, Site B; Bilawali Tank Site A, Site B of Indore Madhya Pradesh during monsoon, summer and winter seasons. Digestive gland is the main site of metal accumulation in the body and as it contains greater level of metallothionein (Pipe *et al.*, 1999; Canesi *et al.*, 2008; Waykar and Shinde, 2013; Deshmulh, 2013). Even digestive glands often also show higher level of antioxidant enzymes in body (Irato *et al.*, 2003). Different toxicant is generating reactive oxygen species and induces the LPO formation. This may increase the activity of GST, decrease the level of GSH and alter different antioxidant enzyme viz. SOD, CAT and GPx activities in different mollusk (Vasseur and Leguille, 2003; Box, *et al.*, 2007; Osman *et al.*, 2007; Shinde, 2013; Deshmukh, 2013). Such studies have indicated the oxidative stress indicator and act as a powerful and cost-effective tool to get information of environment and effect of contaminants on living biological resources. Highest level of LPO and Glutathione-S-transferase activity and lowest superoxide dismutase, glutathione peroxidase and catalase activity and low levels of decreased glutathione (GSH) in the bivalve species ' digestive glands, *Lamellidens corrianus*, *Lamellidens marginalis* and *Unio.sp* collected from Sirpur lake Site A than others. On the other side, the lowest level of lipid peroxidation and Glutathione-S-transferase activity and the highest activity in

the digestive glands of superoxide dismutase, glutathione peroxidase and catalase and reduced glutathione (GSH) of three bivalve species collected from Bilawali tank Site B than others. Lipid peroxidation is a key aspect of cell injury and seems to be largely results by free radical reactions in biological membranes, being rich in polyunsaturated fatty acids (Chesseman, 1982). Metals are known inducers of Reactive oxygen species which covalently bind to macromolecules and inflict peroxidative degeneration of the endoplasmic reticulum lipid membrane; which is rich in polyunsaturated fatty acids. This leads to formation of lipid peroxide, which in turn gives products like, MDA that causes damage to membrane. The substantial increase in lipid oxidation (MDA) may signify the vulnerability of lipid molecules to reactive oxygen species and the degree to which these molecules suffer oxidation harm (Jamil, 2001). Increased lipid peroxide (LPO) is one of the most important contributors to the loss of cell function in oxidative stress conditions (Hermes-Lima *et al.*, 1995). The glutathione-s-transferase (GST) enzyme involved in the detoxification of environmental pollutants, as well as endogenous toxic compounds such as lipid peroxidation products (Ketterer and Chistodoulides, 1994; Awasthi *et al.*, 1995). In the current investigation the highest activity of glutathione-S-transferase (GST) was observed in digestive glands of three bivalve species collected from Site A (Sirpur lake) than other three sites might be due to bivalve Species have been exposed to greater levels of toxins than three other sites. Evidence collected in the present study showed the highest concentrations of heavy metals Zn, Cu, Pb, Mn and Cd in surface water; soil

sediments and three bivalve species inhabiting at this reservoir than other three studied reservoirs. Metals are known inducers of reactive oxygen species (ROS). Higher GST activity at Site A (Sirpur lake) in bivalve digestive glands, the ability of the digestive glands to metabolize xenobiotics could be related; eliminate waste products (Gamble *et al.*, 1995) and it also suggest the defensive action against reactive oxygen radicals. Increase of GST enzyme activity also indicating activation of detoxification complexes in the digestive glands could be a good indicator of the pollutant exposure. Increased GST activity can therefore be due to higher detoxification of hydroperoxides. Many researchers revealed that the increase in activities of GST after exposure to toxicants. Sheehan *et al.*, (1995), reported an increase in GST activity in mussel, *Mytilus edulis* exposed to pollutants. Canesi *et al.*, (1999) reported an increase in GST activity in gill and digestive glands of *M. edulis* after exposure to Cu and Hg. Schuliga *et al.*, (2002) and Ventura-Lima *et al.*, (2007) have shown the modulation of GST activity on arsenic exposure. Compare to Site A (Sirpur lake) the bivalve species inhabiting at Site B (Sirpur lake) showed lesser level of LPO and activity of glutathione-S-transferase (GST) in digestive glands of three bivalve species, this might be related to bioaccumulated metal concentration in bivalves. Results indicate that bivalve inhabiting at Site B (Sirpur lake) are exposed to low levels of pollutants than as in Site A (Sirpur lake). In current study gathered data revealed the low concentration of heavy metals Zn, Cu, Pb, Mn and Cd in surface water, soil sediments and three bivalve species at Site B (Sirpur lake) than Site A (Sirpur lake). Therefore, this

study clearly indicated that Site B (Sirpur lake) was less polluted than Site A (Sirpur lake). Reduced glutathione (GSH) is the major non-protein thiol and is the important endogenous antioxidant which plays a crucial role in the defense against oxidative damage (Gohary *et al.*, 1999; Sies, 1999). GSH could be the first line of defense against metal contamination by metal chelation or participation in the GPx or GST detoxification process (Sies, 1999). It has been proved that reduced glutathione is One of the most effective ROS scavengers produced as cell metabolism by-products or during oxidative stress (O'Brien *et al.*, 2001; Tsukamoto *et al.*, 2002; Han *et al.*, 2008). GSH have antioxidant properties, and Its protective function toward oxidative stress induced toxicity is well known in aquatic organisms (Hasspielar *et al.*, 1994; Gohary *et al.*, 1999). Aquatic organisms have evolved a suite of enzymatic and non-enzymatic defenses to cope with the production of reactive oxygen species (ROS). The antioxidant defense enzyme system includes several enzymes such as Catalase (CAT), Superoxide Dismutase (SOD), and Glutathione peroxidase (GPx). Many among these antioxidants interact in a concerted manner to eliminate reactive oxygen species and prevent damage to cellular components. These enzymes activities can be affected by reactive oxygen species (ROS) and therefore they may represent indicators of oxidative stress (Valavanidis *et al.*, 2006 ;Pavlovic *et al.*, 2004). Various antioxidant enzyme activities are mostly likely used as indicators of oxidative stress (Cargnelutti *et al.*, 2006; Bocchetti *et al.*, 2008 ;Banni *et al.*, 2008; Zhou *et al.*, 2008). The lowest activity of CAT , SOD and GPx activity was associated with enhanced LPO formation. A

deficiency in these cellular defense enzymes might decline the capacity of aquatic organism to neutralize the production of ROS. Numerous researchers showed that the toxicants initiate the LPO formation, increases the activity of GST, decrease GSH level and alter antioxidant enzyme (SOD, CAT and GPx) activities in mollusk (Vasseur and Leguille, 2003; Osman *et al.*, 2007; Box, *et al.*, 2007; Shinde, 2013; Deshmukh, 2013).

### **Biochemical Study**

In the present investigation, biochemical constituents like protein, ascorbic acid, RNA and DNA contents were determined from the soft body tissues like gills, digestive glands and whole soft body tissues of three bivalve species, *Lamellidens corrianus*, *Lamellidens marginalis* and *Unio.sp* inhabiting the two reservoirs of Indore district during three seasons and gathered results were compiled in table nos. 10 to 12 and fig. nos. 51 to 62.

The results revealed lowest protein, ascorbic acid and DNA in soft body tissues of three groups of bivalves collected from Site A (Sirpur lake) than Site B (Sirpur lake), Site A (Bilawali tank) and Site B (Bilawali tank), though high protein, ascorbic acid and DNA content of three bivalve species sampled from Site B (Bilawali Tank) were observed in soft body tissues reservoir than other three studied reservoirs. The findings of the ANOVA test showed that the variation between the mean values of biochemical components between reservoirs, seasons and bivalve species ( $P < 0.05$ ) was significantly different.



In the present study obtained results showed the low level of protein content, ascorbic acid content RNA and DNA content in soft body tissues of three bivalve species located from Site A (Sirpur lake) than other three observed sites of both reservoirs, could be because of the bivalves present at Site A (Sirpur lake) were subject to higher pollutant load than three other sites of both reservoirs recorded. The findings of the present study showed highest concentrations of Zn, Cu, Mn, Pb and Cd heavy metals in surface water, sediments and three bivalve species sampled from Site A (Sirpur lake) than other three analyzed sites of reservoirs. The gained heavy metals were enough to interrupt the natural equilibrium of oxidation / reduction in the cells by generating reactive oxygen species which lead to oxidative stress (Abdullah *et al.*, 2004). In the appearance of reactive oxygen species (ROS), proteins can be weakened by oxidative attack, resulting in peptide chain fragmentation, site-specific variations of amino acids, disrupted electrical load, accumulation of cross-linked reaction products and increased proteolysis sensitivity (Grune, 2000; Requena *et al.*, 2003). The low protein content found in different tissues suggests that environmental pressure decreases translations levels or increases protein catabolism to meet the high energy demands of toxicants (Vincent *et al.*, 1995; Waykar and Lomte, 2001a). The lesser protein composition could be the result of cell destruction / necrosis and resulting degradation of protein synthetic machinery (Umminger, 1977; Bradbury *et al.*, 1987). Protein and amino acid catabolism contribute greatly to total energy output. It is known that structural proteins are used as energy source under stressful conditions

(Claybrook, 1983). Pottinger *et al.*, (2002) disclosed that protein synthesis, represents disruption of normal metabolic processes, may be suppressed at high pollution stress. This work provides information regarding the heavy metal level in surface water and soil sediment and in three bivalve species and biomarker responses in relation to heavy metal accumulation at two reservoirs of Indore district. This research focused on seasonal fluctuations in heavy metal concentrations of surface water and soil sediments, bivalve organisms, and their effects on biomarker responses and biochemical components.

*CONCLUSION  
AND  
RECOMMENDATIONS*

## CONCLUSION AND RECOMMENDATIONS

Reports of heavy metal accumulation in bivalves and their presence in water as well as in sediment are helpful in designing a system of heavy metals removal from water bodies and compliance. Bivalves are proven their usefulness as bio-indicator organisms and provide ideally, an estimate of trace elements availabilities in biomass of water bodies. Heavy metal body burden in bivalves have been used to identify and sketch the areas with exceedingly higher levels of trace metals and organic pollutants. This is due bivalves can be used as bio-monitors for aquatic environment. Bivalves can also providing the details of toxicity and alarm if anyone eat bivalves as food. The baseline data on metal concentrations in Sirpur Lake and Bilawali Tank may contribute in evaluating future impact of anthropogenic activities and progress. Biochemical studies are providing baseline data for assessment of bio-monitoring studies as reference. This study will deal the importance of biomarker research and incorporating the Indian freshwater monitoring program. Results of this study demonstrate the mean concentrations of Zn, Cu, Cd , Pb and Mn in surface water and soil sediment of Sirpur Site A and Site B were higher than site A and B of Bilawali Lake in summer and gradually lower in similar manner in winter and monsoon. Reported values were also higher than the recommendation of WHO limits for drinking water standard. Therefore, central water treatment plants urgently need to treat water and extract heavy metals before using this water for crop and drinking purposes. The haphazard anthropogenic activity near water bodies need to be controlled. Effective environmental monitoring exercise should also be encouraged to check the flow of heavy metals and other pollutant.

# *SUMMARY*

## SUMMARY

Human populations are increasing rapidly in the world and to fulfill the requirement of their needs, anthropological activities are progressively increasing.

Water is an elixir of life, so urbanization near water bodies is establishing. Settlement of population needs leads to technology development, industrialization, farming, and many more activities. These all have been adversely affecting environment and ecosystem.

Heavy metals viz. As, Zn, Cu, Pb, Cd, Mn, Hg, Ni and Cr are some trace metals present in polluted water. Higher concentration of these heavy metals makes harmfully affect to population of water bodies and even they are non-biodegradable and are persistent in nature.

Heavy metals may be transferred through accumulation in lower trophic organisms to higher trophic organism through food chain and leads to bad effect on ecological balance of environment and diversity of aquatic organisms.

The body burdens of metals in most bivalves have been used to identify and map areas with exceedingly high levels of trace metals and organic pollutants hence they can be used as biomonitors for aquatic environment.

Potential of bivalve to accumulate metals from water/sediments into its tissues can be estimated using Bio-water Accumulation Factor (BWAF) and Bio-sediment Accumulation Factor (BSAF). By comparing BWAF/BSAF values,

one can compare the potential of different bivalve species to accumulate metals from water and sediments in to their body tissues.

Bioaccumulation only detects the levels of heavy metals, in aquatic organisms, but a true evaluation of the damage inflicted by heavy metals should come from comprehensive biomarker studies.

India has 15 different ecological regions where 1.42 million population lives. About 37.7 million populations of this country are affected by waterborne diseases annually, in which approximately 1.5 million children suffer with diarrhea and allied disease every year. This is results nearly 600 million dollar burden to Indian economy. Over exploitation and pollution are main causes of deterioration of water quality. Polluted water quality may affect the aquatic life, animals, plants and humans. A regular monitoring of water quality is needed to maintain the quality of water.

In the present study different native species of freshwater bivalves, *Unio sp.*, *Lamellidens corrianus*, *Lamellidens marginalis* were selected to establish a local environmental monitoring network using bivalves as bioindicator species to evaluate the trends of bioaccumulation of Zn, Cu, Pb and Mn in freshwater ecosystem.

Sirpur Lake and Bilawali tank of Indore, Madhya Pradesh was taken for this study. Surface water, sediments soil and three species of bivalve viz. *Unio sp.*,

*Lamellidens corrianus* and *Lamellidens marginalis* were sampled from two different sites of each water bodies.

Surface water, sediments soil and three species of bivalve viz. *Unio sp.*, *Lamellidens corrianus* and *Lamellidens marginalis* were sampled.

Physico-chemical parameters viz. total alkalinity, dissolved oxygen, total hardness, salinity and chloride; heavy metals viz. Zn, Cu, Pb and Mn were determined in laboratory as per standard method of APHA 1998.

Metal bioaccumulation in bivalve species, Biowater Accumulation Factor (BWAFF) and Biosediment Accumulation Factor (BSAF) was calculated. The BWAFF factor is defined as the ratio between the concentration of metal in the organism and that in the water. The BSAF is defined as the ratio between the concentration of metal in the organism and that in the sediments.

Digestive glands of five animals of each species was collected, separated, surface blotted with tissues paper and thoroughly washed with phosphate buffer (50mM; pH 7.4). LPO, GSH, SOD, CAT, GPx and GST was performed for such samples.

Whole soft body tissues were removed and dried at 70<sup>0</sup> to 80<sup>0</sup>C in the oven till the constant weight of dry tissues was obtained. Protein, ascorbic acid, DNA and RNA content was estimated.

Results have shown the higher values of Total alkalinity (mg/l) in Sirpur Lake site A during summer season followed by Sirpur site B, Bilawali site A,



Bilawali site B in summer; Sirpur site A, site B, Bilawali site A site B in monsoon and winter seasons.

Higher values of Dissolved oxygen (mg/l) in Bilawali Lake site A during winter season and reported gradually decreased values in Sirpur site B, site A, Bilawali site B in winter; Bilawali site B, site A Sirpur site B, site A in monsoon and summer seasons.

Highest total hardness (mg/l) was recorded in Sirpur site A and then Sirpur site B, Bilawali site A, site B in summer season. Gradually decreased values were observed in Sirpur site A, site B, Bilawali site A, site B in winter season; Sirpur site A, site B, Bilawali site A, site B in monsoon season.

Highest Salinity (mg/l) was recorded in Sirpur site A and then Sirpur site B, Bilawali site A, site B in summer season. Gradually decreased values were observed in Sirpur site A, site B, Bilawali site A, site B in winter season; Sirpur site A, site B, Bilawali site A, site B in monsoon season.

Highest Chloride (mg/l) was recorded in Sirpur site A and then Sirpur site B, Bilawali site A, site B in summer season. Gradually decreased values were observed in Sirpur site A, site B, Bilawali site A, site B in winter season; Sirpur site A, site B, Bilawali site A, site B in monsoon season.

Temperature was recorded within normal range everywhere. Highest pH (unit) was recorded in Sirpur site A in every season in comparison to other sampling sites. This result clearly indicated that Site A (Sirpur lake) was more polluted

than Bilawali Lake sites and Sirpur site B. Obtained results are statistically proven with  $P < 0.05$ .

The higher values of pH, total alkalinity, total hardness, chlorides, salinity and lowest dissolved oxygen were observed in surface water of Site A (Sirpur lake) may be due to huge agricultural runoff, domestic waste, textile and other industrial effluents. Catchment areas of Sirpur Lake Site A are surrounded by agricultural field, which is polluted by different fertilizers and pesticides. There are two sugar and one cotton mill are located in the basin area of this Lake. Kalwan, Satana, Deola and some others are neighboring towns of this Lake and discharges untreated domestic waste to this Lake. Maximum pH in winter is due to decreased decomposition rate.

Assessments of heavy metals in surface water samples and sediment of two different sites of Sirpur and Bilawali Lakes of Indore were performed and reported the mean concentrations of Zn, Cu, Pb and Mn in surface water of Sirpur Site A and Site B were higher than site A and B of Bilawali Lake in summer and gradually lower in similar manner in winter and monsoon. The main reason behind the pollution of Site A (Sirpur lake), is rivulets of Site A (Sirpur lake) river namely Tamdi, Punand, Aram, Mosam, Masa Nadi, Baidki and markhandi etc. The catchment areas of these rivulets are surrounded by agricultural field, which is polluted by fertilizers and pesticides, these acts as source of heavy metals Cu, Cd, Pb, Zn and As. Overall results showed that highest concentration of heavy metals Mn, Cu, Pb, and Zn in sediments

samples of Sirpur Lake Site A than Sirpur Lake Site B, Bilawali Tank Site A and Bilawali Tank Site B.

Concentrations of heavy metals viz. Mn, Cu, Pb, Cd and Zn metal body burden, BSAF and BSAF values were determined in dry weight of whole soft body tissues of three different freshwater bivalve species i.e. *Lamellidens corrianus*, *Lamellidens marginalis* and *Unio*.species collected from Sirpur Lake Site A, Site B; Bilawali Tank Site A, Site B of Indore Madhya Pradesh during monsoon, summer and winter seasons. Results indicate that mean highest values of concentrations of heavy metals in body, metal body burden and BSAF and BSAF in dry soft body tissues of all sampled bivalve species collected from Sirpur Lake Site A than Sirpur Lake Site B, Bilawali tank Site A and Bilawali tank Site B. Reported higher metal concentrations and body burden in whole dry soft body tissues of all sampled bivalve species collected from Sirpur Lake Site A might be due to exposure of higher pollutant than other sampling sites.

Digestive gland is the main site of metal accumulation in the body and as it contains higher level of metallothionein. Different toxicant is generating reactive oxygen species and induces the LPO formation. This may increase the activity of GST, decrease the level of GSH and alter different antioxidant enzyme viz. SOD, CAT and GPx activities in different mollusk. Such studies have indicated the oxidative stress indicator and act as a powerful and cost-effective tool to get

information of environment and effect of contaminants on living biological resources.

Highest level of LPO and Glutathione-S-transferase activity and lowest activity of superoxide dismutase, catalase and glutathione peroxidase and low level of reduced glutathione (GSH) in the digestive glands of bivalve species, *Lamellidens corrianus*, *Lamellidens marginalis* and *Unio.sp* collected from Sirpur lake Site A than others. On other hand lowest level of lipid peroxidation and glutathione-S-transferase activity and highest activity of superoxide dismutase, catalase and glutathione peroxidase and level of reduced glutathione (GSH) in the digestive glands of three bivalve species collected from Bilawali tank Site B than others.

Lipid peroxidation is a main aspect in cellular injury and results largely from free radical reactions in biological membranes, which are rich in polyunsaturated fatty acids (Chapman, 1997). Metals are known inducers of reactive oxygen species which binds covalently to the macromolecules and induce peroxidative degeneration of the lipid membrane of endoplasmic reticulum, which is rich in polyunsaturated fatty acids. This leads to formation of lipid peroxide, which in turn gives products like, MDA that causes damage to membrane. The significant increase in lipid oxidation (MDA) may indicate the susceptibility of lipid molecules to reactive oxygen species and the extent of oxidative damage imposed on these molecules.

Increased lipid peroxide (LPO) is one of the most important contributors to the loss of cell function in oxidative stress conditions (Hermes-Lima *et al.*, 1995). The glutathione-s-transferase (GST) enzyme involved in the detoxification of environmental pollutants, as well as endogenous toxic compounds such as lipid peroxidation products

In the present investigation the highest activity of glutathione-S-transferase (GST) was observed in digestive glands of three bivalve species collected from Site A (Sirpur lake) than other three reservoirs might be due to bivalve species were exposed to higher level of pollutants than other three reservoirs. In the present study obtained data revealed highest concentrations of heavy metals Zn, Cu, Pb and Mn in surface water, soil sediments and three bivalve species inhabiting at this reservoir than other three studied reservoirs. Metals are known inducers of reactive oxygen species (ROS).

Higher GST activity at Site A (Sirpur lake) in the digestive glands of the freshwater bivalve might be related to the capacity of the digestive glands to metabolize xenobiotics, eliminate waste products and it also suggests the protective action against reactive oxygen radicals. Increase of GST enzyme activity indicating activation of detoxification mechanism in the digestive glands could be a good indicator of pollutant exposure. Increase of GST activity can therefore be due to increased detoxification of hydroperoxides.

Lowest level of LPO and activity of Glutathione-S-transferase (GST) in digestive glands of three bivalve species, this might be related to

bioaccumulated metal concentration in bivalves. Results indicate that bivalve inhabiting at Site B (Sirpur lake) was exposed to low level of pollutants than Site A (Sirpur lake). In the present study obtained data revealed the low concentration of the heavy metals Zn, Cu, Pb, Cd and Mn in surface water, soil sediments and three bivalve species at Site B (Sirpur lake) than Site A (Sirpur lake).

This study clearly indicated that Site B (Sirpur lake) was less polluted than Site A (Sirpur lake). Reduced glutathione (GSH) is the major non-protein thiol and is the important endogenous antioxidant which plays a central role in the defense against oxidative

In the present investigation the biochemical constituents like protein, ascorbic acid, DNA and RNA contents were determined from soft body tissues like mantle, gills, digestive glands and whole soft body tissues of three bivalve species, *Lamellidens corrianus*, *Lamellidens marginalis* and *Unio.sp* inhabiting the two reservoirs of Indore district during three seasons. The results showed, lowest protein, ascorbic acid and DNA contents in soft body tissues of three bivalves species sampled from Site A (Sirpur lake) than Site B (Sirpur lake), Site A (Bilawali tank) and Site B (Bilawali tank), while highest protein, ascorbic acid, DNA and RNA contents were observed in soft body tissues of three bivalve species sampled from Site B (Bilawali Tank) reservoir than other three studied reservoirs. The results of ANOVA test indicated that the difference between the mean values of biochemical constituents were varied

significantly between reservoirs, seasons and bivalve species ( $P < 0.05$ ). In the present study obtained results showed the low level of protein, ascorbic acid, DNA and RNA contents in soft body tissues of three bivalve species collected from Site A (Sirpur lake) than other three studied reservoirs, might be due to bivalves inhabiting at Site A (Sirpur lake) were exposed to higher load of pollutants than other three studied reservoirs.

Seasonal variation of heavy metal concentrations in surface water and soil sediment and bivalve species and their effect on biomarkers responses and biochemical components are focused in this study and reported information regarding heavy metal level in surface water and soil sediment and in three bivalve species and biomarker responses in relation to heavy metal accumulation at reservoirs of Indore district.

# *REFERENCES*



## REFERENCES

- Abaychi J.K. and Dou Abul A.A.Z. (1985) Trace metals in Shatt Al-Arab river, Iraq. *Water Res.* 19(4): 457-462.
- Abdelmoneim M. (1995) Assessment of cadmium, lead, copper and zinc in fish species reared in treated sewage effluents in Suez city, Egypt. *Bull High Inst Public Health.* 25: 227-234.
- Abdullah A.M., El-Mogy M.A., Farid N.M. and El-Sharabasy M.M. (2004) Purification and characterization of glutathione transferases from *Bulinus truncatus*. *J. of Genetic Eng. & Biotechnol (NRC).* 8773: 2.
- Aebi H. (1984) Catalase invitro. *Methods in Enzymol.* 105: 121-126.
- Ahmed Z., Akhter F., Hussain M.A., Haque M.S., Sayeed M.M.A. and Quashem M.A. (2002) Researches on jute and allied fibre plants. *Pak. J. Biol. Sci.* 5: 812-818.
- Aksoy A., Demirezen D. and Duman F. (2005) Bioaccumulation, detection and analysis of heavy metal pollution in Sultan Marsh and its environment. *Water Air and Soil Pollution.* 164: 241-255.
- Alkarkhi F.M.A., Ismail N. and Easa A.M. (2008) Assessment of arsenic and heavy metal contents in cockles (*Anadara granosa*) using multivariate statistical techniques. *J Haz Mat.* 150: 783-789.
- Apeti D.A., Johnson E. and Robinson L. (2005) A Model for Bioaccumulation of Metals in *Crassostrea virginica* from Apalachicola Bay, Florida. *American J Environ Sci.* 1: 239-248.
- APHA (1998) Standard methods for the examination of water and waste water. American Public Health Association Washington 20<sup>th</sup> edition, Washington DC (U.S.A.), 1.

- Awasthi Y.C., Zimniak P., Singhal S.S. and Awasthi S. (1995) Physiological role of glutathione S-transferases in protection mechanisms against lipid peroxidation: a commentary. *Biochem. Arch.* 11: 47-54.
- Banni M., Bouraoui Z., Ghedira J., Clerandau C., Guerbej H., Narbonne J.F. and Boussetta H. (2008) Acute effects of benzo[a]pyrene on liver phase I and II enzymes, and DNA damage on sea bream *Sparus aurata*. *Fish Physiol Biochem.* 34: 201-207.
- Barak N. and Mason C. (1990) A Survey of Heavy Metal Levels in Eels (*Anguilla anguilla*) from Some Rivers in East Anglia, England-The Use of Eels as Pollution Indicators. *Internationale Revue Der Gesamten Hydrobiologie.* 75(6): 827-833.
- Beauchamp C. and Fridovich I. (1973): Isozymes of superoxide dismutase from wheat germ. *Biochimica et Biophysica Acta.* 317: 50-64.
- Bebianno M.J., Nott J.A. and Langston W.J. (1993) Cadmium metabolism in the clam *Ruditapes decussata*: the role of metallothioneins. *Aquat Toxicol.* 27: 315-333.
- Biliaff B., O'Conner T.P., Daskalakis D.K. and Smith P.J. (1997) US mussel watch data from 1986 to 1994, Temporal trend detection at large spatial scales. *Environ Sci Technol.* 31: 1411-1415.
- Biliaff B., O'Conner T.P., Daskalakis D.K. and Smith P.J. (1997) US mussel watch data from 1986 to 1994, Temporal trend detection at large spatial scales. *Environ Sci Technol.* 31: 1411-1415.
- Blasco J., Arias A.M. and Saenz V. (1999) Heavy metals in organisms of the River Guadalquivir estuary. possible incidence of the Aznalcóllar disaster. *Sci Tot Environ.* 242: 249-259.

- Bocchetti R., Fattorini D., Pisanelli B., Macchia S., Oliviero L., Pilato F., Pellegrini D., Regoli F. (2008) Contaminant accumulation and biomarker responses in caged mussels, *Mytilus galloprovincialis*, to evaluate bioavailability and toxicological effects of remobilized chemicals during dredging and disposal operations in harbor areas. *Aquat Toxicol.* 89: 257-266.
- Bonneris E., Giguere A., Perceval O., Buronfosse T., Masson S., Hare L. and Campbell P.G.C. (2005) Sub-cellular partitioning of metals (Cd, Cu, Zn) in the gills of a freshwater bivalve, *Pyganodon grandis* role of calcium concretions in metal sequestration. *Aquat Toxicol.* 71: 319-334.
- Boran M. and Altinok N. (2010) A review of heavy metals in water sediment and living organism in the black sea. *Turkish Journal of Fishries and Aquatic Science.* 10: 565-572.
- Box A., Sureda A., Galgani F., Pons A. and Deudero S. (2007) Assessment of environmental pollution at Balearic Islands applying oxidative stress biomarkers in the mussel *Mytilus galloprovincialis*. *Comparative Biochemistry and Physiology.* 146(C): 531-539.
- Boyden C.R. (1974) Trace element content and body size in molluscs. *Nature.* 251: 311-314.
- Boyden C.R. (1974) Trace element content and body size in molluscs. *Nature.* 251: 311-314.
- Bradbury S.P., Symonik D.M., Coats J. R. and Atchison G. J. (1987) Toxicity of fenvalerate and its constituent isomers to the fat head minnow (*Pimephales promelas*), bluegill (*Lepomis macrochirus*). *Bull. Environ. Contam. Toxicol.* 38: 727-35.

- Bryan G.W., Gibbs P.E., Hummerstone L.G. and Burt G.R. (1986) Journal of the Marine Biological Association of the United Kingdom. 66: 611-640.
- Campanell L., Conti M.E., Cubadda F. and Sucapane C. (2001) Trace metals in seagrass, algae and molluscs from an uncontaminated area in the Mediterranean. Environ Pollut. 111: 117-126.
- Canesi L., Borghi C., Ciacci C., Fabbri R., Lorusso L.C., Vergani L., Marcomini A. and Poiana G. (2008) Short term effects of environmentally relevant concentrations of EDC mixtures on *Mytilus galloprovincialis* digestive gland. Aquat. Toxicol. 87: 272-279.
- Canesi L., Viarengo A., Leonzio C., Filippelli M. and Gallo G. (1999) Heavy metals and glutathione metabolism in mussel tissues. Aquat. Toxicol. 46: 67-76.
- Cargnelutti D., Tabaldi L.A., Spanevello R.M., Jucoski G.O., Battisti V., Redin M., Linares C.E.B., Dressler V.L., Flores E.M.M., Nicoloso F.T., Morsch V.M. and Schetinger M.R.C. (2006) Mercury toxicity induces oxidative stress in growing cucumber seedlings. Chemosphere. 65: 999-1006.
- Casas S., Gonzalez J.L., Andral B. and Cossa D. (2008) Relation between metal concentration in water and metal content of marine mussels (*Mytilus galloprovincialis*): impact of physiology. Environ Toxicol Chem. 27: 1543-1552.
- Censi P., Spoto S.E., Saiano F., Sprovieri M. and Mazzola S. (2006) Heavy metals in coastal water system. A case study from the North Western Gulf of Thailand. Chemosphere. 64: 1167-1176.

- Chang S., Zdanowicz V.S. and Murchelano R.A. (1998) Associations between liverlesions in winter flounder *Pleuronectes americanus*) and sediment chemical contaminants from north-east United States estuaries. *Journal of Marine Science*. 55(5): 954-969.
- Chapman P.M. (1997) Is bioaccumulation useful for predicting impacts? *Mar Pollut Bull*. 34(5): 282-283.
- Chase M.E., Jones S.H., Hennigar P., Sowless J., Harding G.C.H., Freeman K., Wells P.G., Krahforst C., Coombs K. and Crawford R. (2001) Gulf watch: monitoring spatial and temporal patterns of trace metal and organic contaminants in the Gulf of Maine [1991–1997] with the Blue Mussel *Mytilus edulis* L. *Mar Pollut Bull*. 42: 491-505.
- Cheng S.P. (2003) Heavy metal pollution in China: Origion, pattern and control. *Environ. Sci. Pollut. Res. Int*. 10: 192-198.
- Cheung K.C., Poon B.H.T., Lan C.Y. and Wong M.H. (2003) Assessment of metal and nutrient concentrations in river water and sediment collected from the cities in the Pearl River Delta, South China. *Chem*. 52: 1431-1440.
- Choudhary P., Dhakad N.K. and Jain R. (2014) Studies on the Physico-Chemical Parameters of Bilawali Tank, Indore (M.P.) India. *IOSR Journal of Environmental Science, Toxicology and Food Technology (IOSR-JESTFT)*. 8(1.I): 37-40.
- Claybrook D.C. (1983) In: *The Biology of Crustacea. Internal anatomy and Physiological Regulation* (Edited by Mantel L. H.) Academic press, New York. pp. 163-202.
- Cossu C., Doyotte A., Jacquin M., Babut M., Exinger A. and Vasseur P. (1997) Glutathione reductase, selenium-dependent glutathione peroxidase, glutathione levels, and lipid peroxidation in freshwater bivalves,

*Unio tumidus*, as biomarkers of aquatic contamination in field studies. *Ecotoxicology and Environmental Safety*. 38: 122-131.

D'Souza C. and Peretiakko R. (2002) The nexus between industrialization and environment: A case study of Indian enterprises. *Environ. Manag. Health*. 13: 80-97.

de Meeus C., Eduljee G.H. and Hutton M. (2002) Assessment and management of risks arising from exposure to cadmium in fertilizers I. *Sci. Total Environ*. 291(1-3): 167-187.

de Mora S., Fowler S.W., Wyse E. and Azemard S. (2004) Distribution of heavy metals in marine bivalves, fish and coastal sediments in the Gulf and Gulf of Oman. *Mar Pollut Bull*. 49: 410-424.

Dehadri P.V. (1990) Pollution and aquaculture of Land and water, metal mercuric chloride on the enzyme alkaline phosphates from *Labeo rohita*. *J.Aqua. Biol. Env. Poll. and Res*. 22(2): 142-144.

Deshmukh G.M. (2013) Biomonitoring of heavy metal pollution of jayakwadi reservoir at Paithan by using bivalves as bioindicators. Ph.D. thesis submitted to Dr. BAM University, Aurangabad, (MS) India.

Diagomanolin V., Farhang M., Ghazi-Khansari M. and Jafarzadeh N. (2004) Heavy metals (Ni, Cr, Cu) in the Karoon waterway river, Iran. *Toxicol Lett*. 151: 63-68.

Elder J.F. and Collins J.J. (1991) Freshwater molluscs as indicators of bioavailability and toxicity of metals in surface water systems. *Rev Environ Cont Toxicol*. 122: 36-79.

El-Gohary M., Awara W.M., Nassar S. and Hawas S. (1999) Deltamethrin-induced testicular apoptosis in rats: the protective effect of nitric oxide synthetase inhibitor. *Toxicology*. 132(1): 1-8.

- Ellman G.L. and Boyne A.F. (1972) A methodology for analysis of tissue sulfhydryl components. *Anal. Biochem.* 46: 639-653.
- Farkas A., Erratico C. and Vigano L. (2007) Assessment of the environmental significance of heavy metal pollution in surficial sediments of the River Po. *Chem.* 68: 761-768.
- Farkas A., Erratico C. and Vigano L. (2007) Assessment of the environmental significance of heavy metal pollution in surficial sediments of the River Po. *Chem.* 68: 761-768.
- Farrel T.P., Finlayson C.M. and Griffiths D.J. (1979) Studies on the hydrobiology of a tropical lake in North- Western Queensland. I. Seasonal changes in chemical characteristics. *Australian Journal of Marine and Freshwater Research.* 30: 579-595.
- Ferreira A.G. , Machado A.L.D.S. and Zalmon I.R. (2004) Temporal and Spatial Variation on Heavy Metal Concentrations in the bivalve *Perna perna* (LINNAEUS,1758) on the Northern Coast of Rio de Janeiro State, Brazil. *Braz Arch Biol Tech.* 47: 319-327.
- Flora S.J.S., Mittal M. and Mehta A. (2008) Heavy metal induced oxidative stress and its reversal by chelation therapy. *Ind. J. Med. Res.* 128: 501-523.
- Funes V., Alhama J., Navas J.I., Barea J.L. and Peinado J. (2006) Ecotoxicological effects of metal pollution in two mollusk species from the Spanish South Atlantic littoral. *Environ Poll.* 139: 214-223.
- Gamble S., Goldfrab P.S., Porte C. and Livingstone D.R. (1995) Glutathione peroxidase and other antioxidant enzyme function in marine invertebrates (*Mytilus eulis*, *Pecten maximus*, *Carcinus maenas* and *Asterias rubens*) *Mar. Environ. Res.* 39: 191-195.

- Gay D. and Maher W. (2003) Natural variation of copper, zinc, cadmium and selenium concentrations in *Bembicium nanum* and their potential use as a biomonitor of trace metals. *Wat Res.* 37: 2173-2185.
- Ghabbour E.A., Shaker M., El-Toukhy A., Abid I.M. and Davies G. (2006) Thermodynamics of metal cation binding by a solid soil-derived humic acid. Binding of Fe(III), Pb(II), and Cu(II). *Chem.* 63: 477-483.
- Giarratano E. and Amin O.A. (2010) Heavy metals monitoring in the southern most mussel farm of the world (Beagle Channel, Argentina) *Ecotox Environ Saf.* 73: 1378-1384.
- Goldman J.C. (1972) Effect of inorganic carbon on eutrophication. Proceedings of a Seminar on Eutrophication and Biostimulation. Brown R.L. and Tunzi M.G. (Eds.). California Development of Water Resources, San Francisco. pp. 30-53.
- Gregori I.D., Pinochet H., Gras N. and Mufioz L. (1996) Variability of cadmium, copper and zinc levels in molluscs and associated sediments from chile. *Environ Poll.* 92: 359-368.
- Grune T. (2000) Oxidative stress, aging and the proteasomal system. *Biogerontology.* 1: 31-40.
- Gundacker C. (2000) Comparison of heavy metal bioaccumulation in freshwater mollusks of urban river habitats in Vienna. *Environ Poll.* 110: 61-71.
- Gundacker C. (2000) Comparison of heavy metal bioaccumulation in freshwater mollusks of urban river habitats in Vienna. *Environ Poll.* 110: 61-71.



- Gupta S.K. and Singh J. (2011) Evaluation of mollusc as sensitive indicator of heavy metal pollution in aquatic system: A review. The IIOAB Journal special issue on environmental management for sustainable development. 2 (1): 49-57.
- Habig W.J., Babst M. J. and Jacoby W.J. (1974) Glutathione s-transferase the first step in mercapturic acid formation. J.B.C. 249: 7130.
- Hadjmohammad M.R. (1998) Quantitative and qualitative determination of heavy and toxic elements in the fish grown in treated wastewater and their comparison with other fish. Iran. J. Chem. Eng. 12: 20-31.
- Halcrow W., Mackay D.W. and Thornton I. (1973) The Distribution of trace metals and fauna in the firth of Clyde in relation to the disposal of sewage sludge. Journal of the Marine Biological Association of the United Kingdom. 53: 721-739.
- Han E.S., Muller F.L., Pérez V.I., Qi W., Liang H., Xi L., Fu C., Doyle E., Hickey M. and Cornell J. (2008) The in vivo gene expression signature of oxidative stress. *Physiol. Genomics*. 34: 112-126.
- Hargrave B.T., Phillips G.A., Vass W.P., Bruecker P., Welch H.E. and Siferd T.D. (2000) Seasonality in bioaccumulation of organochlorines in lower trophic level Arctic marine biota. *Environmental Science and Technology*, 34(6): 980-987.
- Harland B.J., Taylor D. and Wither A. (2000) The distribution of mercury and other trace metals in the sediments of the Mersey Estuary over 25 years: 1974-1998. *The Science of the Total Environment*. 253: 45-62.
- Hasspielar B.M., Behar J.V., Di Giulio R.T. (1994) Glutathione dependant defense in channel catfish (*Ictalurus punctatus*) and brown bullhead

(*Ameiurus nebulosus*). Ecotoxicology and Environmental Safety. 28: 8-90.

Hendozko E. and PiotrSzefer Jan W. (2010) Heavy metals in *Macoma balthica* and extractable metals in sediments from the southern Baltic Sea. Ecotox Environ Saf. 73: 152-163.

Hermes-Lima M., Willmore W.G. and Storey K.B. (1995) Quantification of lipidperoxidation in tissue extracts based on Fe(III) xylenol orange complexformation. Free Radic. Biol. Med. 19:271-280.

Idris A.M., Eltayeb M.A.H., Potgieter-Vermaak S.S., Van Grieken R. and Potgieter J.H. (2007) Assessment of heavy metals pollution in Sudanese harbours along the Red Sea Coast. Microchem J. 87: 104-112.

Injal A.S. and Raut P.D. (2009) Remedial effect of calcium on lead induced alterations in Proteins and phosphatase activities in the gills and mantle of freshwater bivalve, *Lamellidens marginalis*. J. Ecophysiol. Occup. Hlth. 9: 77-84.

Irato P., Santovito G., Cassini A., Piccinni E. and Albergoni V. (2003) Metal accumulation and binding protein induction in *Mytilus galloprovincialis*, *Scapharca inaequivalvis*, and *Tapes philippinarum* from the Lagoon of Venice. Archives of Environmental Contamination and Toxicology. 44: 476-484.

Jain C.K. (2004) Metal fractionation study on bed sediments of River Yamuna, India. Wat Res. 38:569-578.

Jain C.K. and Sharma M.K. (2001) Distribution of trace metals in Hindon river system, India. J Hydrol. 253: 81-90.

- Jain S. and Salman S. (1995) Heavy metal concentration in highly eutrophic lake sediments and overlying water. *Pollut. Res.* 14(4): 471-476.
- Jaishree and Khan T.I. (2014) Monitoring of heavy metal in textile waste water of Sanganer, Jaipur (Rajasthan). *International Journal of Scientific and Research Publications*, ISSN 2250-3153. 4:3
- Jamil K. (2001) Bioindicators and biomarkers of environmental pollution and risk assessment. Science publisher Inc. Enfield USA. pp. 136-146.
- Jan A.T., Ali A. and Haq Q.M.R. (2011) Glutathione as an antioxidant in inorganic mercury induced nephrotoxicity. *J. Postgrad. Med.* 57: 72-77.
- Kalff J. and Knoechel R. (1998) Phytoplankton and their Dynamics in Oligotrophic and Eutrophic Lakes. *Ann. Rev. Ecol. Syst.* 9: 475-495.
- Kara C. and Comlekci U. (2004) *Journal of KSU.* 7:1.
- Karageorgis A.P., Nikolaidis N.P., Karamanos H. and Skoulikidis N. (2003) Water and sediment quality assessment of the Axios River and its coastal environment. *Cont Shelf Res.* 23: 1929-1944.
- Kazanci N. and Dugel M. (2000) Ordination and classification of macro-invertebrates and environmental data of stream in Turkey. *Water Sci. Technol.* 47: 7-8.
- Ketterer B. and Chistodoulides L.G. (1994) Enzymology of cytosolic glutathione S-transferases. *Adv. Pharmacol.* 27:37-69.
- Kim H. and Kim J.G. (2006) Heavy metal concentration in the mollusk gastropod, *Cipangopaludina chinensis malleata* from Upwetland

reflect the level of heavy metal in the sediments. J. Ecol. FieldBiol. 29: 453-460.

Kolhe B., Zambare S.P. and Andhale S.B. (2014) Assesment of Physico-chemical water parameter using correlation analysis: A case study of Gangapur dam at Nasik district (M.S.) India

Lee C.S., Li X., Shi W., Cheung S.C. and Thornton I. (2006) Metal contamination in urban, suburban, and country park soils of Hong Kong. A study based on GIS and multivariate statistics. Sci Tot Environ. 356: 45-61.

Lee C.S., Li X., Shi W., Cheung S.C. and Thornton I. (2006) Metal contamination in urban, suburban, and country park soils of Hong Kong. A study based on GIS and multivariate statistics. Sci Tot Environ. 356: 45-61.

Lokhande R.S., Singare P.U. and Pimple D.S. (2011) Study of physicochemical parameters of wastewater effluents from Tajoja Industrial area of Mumbai India . Intern J Ecosystem. 1(1): 19.

Lopez M.A., Faria M., Sanjuan M.F., Lacorte S. and Barata C. (2010) Comparative toxicity of single and combined mixtures of selected pollutants among larval stages of the native freshwater mussels (*Unio elongatulus*) and the invasive zebra mussel (*Dreissena polymorpha*), Sci Tot Environ. 408: 2452-2458.

Lowry O.M., Rosenbrough N.J., Farr A.C. and Randall R.F. (1951) Protein estimation with Folin Phenol reagent. J. Biol. Chem., 193: 265-275.

Luoma S.N. (1983) Bioavailability of trace metals to aquatic organisms -A review. Sci Tot Environ. 28: 3-22.

- Mahajan S.S. (2007) Ascorbate effect on certain heavy metal induced physiological alteration in the freshwater bivalve, *Lamellidins marginalis* (Lamarck), Ph. D. Thesis submitted to North Maharashtra University, Jalgaon. (M. S.), India.
- Meharg A.A. (2004) Arsenic in rice- understanding a new disaster for south-east Asia. *Trends Plant. Sci.* 9:415-417.
- Meyer J.L. (2007) The contribution of headwater streams to biodiversity in river networks. *J. Am. Water Res. Assoc.* 43: 86-103.
- Mhatre G.N. (1991) bioindicators and biomonitoring of heavy metals. *J Environ Biol.* pp. 201-209.
- Mhatre G.N. (1991) Bioindicators and biomonitoring of heavy metals. *J Environ Biol.* pp. 201-209.
- Mishra B.P. and Tripathi B.D. (2001) Impact of city sewage discharge on physico-chemical characteristics of Gangawater. *Asian Journal of Microbiology, Biotechnology and Environmental Sciences.* 3:333-338.
- Mubiana V.K., Vercauteren K. and Blust R. (2006) The influence of body size, condition index and tidal exposure on the variability in metal bioaccumulation in *Mytilus edulis*. *Environ Poll.* 144: 272-279.
- Nakhle K.F., Cossa D., Khalaf G. and Beliaeff B. (2006) *Brachidontes variabilis* and *Patella* sp. as quantitative biological indicators for cadmium, lead and mercury in the Lebanese coastal waters. *Environ Poll.* 142: 73-82.
- Nandurkar H.P. and Zambare S.P. (2010) effect of board spectrum antibiotics on oxygen consumption rate of fresh bivalves *Parreysia cylindrical* (Annandale and Prashad). *J. Env. Sci.* 24(2):183-187.

- Newman M.C. and McIntosh A.W. (1982) Influence of lead in components of a fresh water ecosystem on molluscan tissue lead concentrations. *Aquatic Toxicol.* 2: 25-29.
- Niyogi S. and Wood C.M. (2004) Biotic ligand model, a flexible tool for developing site-specific water quality guidelines for metals. *Environmental Science and Technology.* 38: 6177-6192.
- Noorjahan C.M., Dawood S.S., Nausheen D. and Ghousia N. (2002) Studies on the Untreated Tannery Effluent and its Effects on Biochemical Constituents of Marine Crab scyllia. *Indian j. environ.Toxicol.* 15-17.
- Norkko J., Thrush S.F. (2006) Ecophysiology in environmental impact assessment: implications of spatial differences in seasonal variability of bivalve condition. *Marine Ecology Progress Series.* 326: 175-186.
- Norkko J., Thrush S.F. and Wells R.M.G. (2006) Indicators of short-term growth in bivalves: detecting environmental change across ecological scales. *Journal of Experimental Marine Biology and Ecology.* 337: 38-48.
- O'Brien A.M., Smith A.T. and O'Fagain C. (2003) Effects of phthalic anhydride modification on horseradish peroxidase stability and activity. *Biotechnology and bioengineering.* 81: 233-240.
- Ohkawa H., Ohishi N. and Yagi K. (1979) Assay for lipid peroxidation in animal tissues by thiobarbituric acid reaction. *Annals of Biochemistry.* 95: 351-358.
- Osman A., Heuvel H. and Noort P. (2007) Differential responses of biomarkers in tissues of a freshwater mussel, *Dreissena polymorpha*, to the exposure of sediment extracts with different levels of contamination. *Journal of Applied Toxicology.* 27: 51-59.

- Pandey A.K. and Pandey G.C. (2003) Physicochemical characteristics of city sewage discharge into river Saryu at Faizabad, Ayodhya. *Him.J.Env.Zool.* 17: 85-91.
- Patil G.V. and Pokhrel K. (2004) Biomedical solid waste management in an Indian hospital: a case study. *Waste Management.* 25:592-599.
- Pavlovic S.Z., Belic D., Blagojevic D.P., Radojicic M.R., Zikic V.R., Saicic S. Z., Grubor-Lajsic G. and Spasic M.B. (2004) Seasonal variations of cytosolic antioxidant enzyme activities in the liver and white muscle of thin lip gray mullet (*Liza ramada Risso*) from the Adriatic sea. *Cryo Letters.* 25: 273-285.
- Philips D.J.H. (1977) The use of biological indicator organisms to monitor trace metal pollution in marine and estuarine environments, a review. *Environ Poll.* 13: 281-317.
- Philips D.J.H. (1977) The use of biological indicator organisms to monitor trace metal pollution in marine and estuarine environments, a review. *Environ Poll.* 13: 281-317.
- Pipe R.K., Coles J.A., Carissan F.M.M. and Ramanathan K. (1999) Copperinduced immunomodulation in the marine mussel *Mytilus edulis*. *Aquat. Toxicol.* 46:43-54.
- Pottinger T.G., Carrick T. R., Yeomans W.E. (2002) The three-spined stickleback as an environmental sentinel: effects of stressors on whole-body physiological indices. *J Fish Biol.* 61: 207-229.
- Pourang N. (1996) Heavy metal concentrations in surficial sediments and benthic macroinvertebrates from Anzali wetland, Iran. *Hydrobiologia.* 331:53-61.

- Presley B.J., Trefry J.H. and Shokes R.F. (1980) Heavy metals inputs to Mississippi deltasediments, a historical view, *Water, Air andsoil Pollution*. 13: 481-494.
- Prüss A. (1998) Review of epidemiological studies on health effects from exposure to recreational water. *International Journal of Epidemiology*, 27(1): 1-9.
- Rainbow P.S., Wolowicz M., Fialkowski W., Smith B.D. and Sokolowski A. (2000) Biomonitoring of trace metals in the gulf of gdansk, using mussels (*Mytilus trossulus*) and barnacles (*Balanus improvisus*). *Wat Res*. 34: 1823-1829.
- Rao S.K., Murthy K.S., Reddy B.K., Swani K.S. and Chetty C.S.R. (1987) Effect of benthocarb on protein metabolism of fresh water teleost fish, *Labeo rohita* (Ham). *Proc. Acad. Env. Biol.*, 7: 143-148.
- Rashed M.N. (2001) Monitoring of environmental heavy metals in fish from Nasser Lake. *Environ. Int*. 27: 27-33.
- Ravera O. (2004) Importance and difficulties of research on the metal speciation in the aquatic ecosystem: an ecologist's viewpoint. *Annali di Chimica* 94: 495-504.
- Renier N.K.K. and Sparling D.W. (2001) Relationships between ambient geochemistry, watershed land-use and trace metal concentrations in aquatic invertebrates living in stormwater treatment ponds. *Environ Poll*. 112: 183-192.
- Requena J.R., Levine R.L. and Stadtman E.R. (2003) Recent advances in the analysis of oxidized proteins. *Amino Acids*, 25: 221–226.
- Richardson J.S. and Danehy R.J. (2007) Asynthesis of ecology of head water stream and their reparian zones in temperate forests.



- Romeo M., Bennani N., Gnassia-Barelli M., Lafaurie M., Girard J.P. (2000) Cadmium and copper display different responses towards oxidative stress in the kidney of the sea bass *Dicentrarchus labrax*. *Aquat. Toxicol.* 48:185-194.
- Romeo M., Frasila C., Barelli M.G., Damiens G., Micu D. and Mustata G. (2005) Biomonitoring of trace metals in the Black Sea (Romania) using mussels *Mytilus galloprovincialis*. *Wat Res.* 39: 596-604.
- Romeo M., Sidoumou Z. and Gnassia-Barelli M. (2000) Heavy metals in various molluscs from the Mauritanian coast. *Bull Environ Contam Toxicol.* 65: 269-276.
- Ron *van-der* O., Beyer J. and Vermeulen N.P.E. (2003) Fish bioaccumulation and biomarkers in environmental risk assessment: a review. *Environ Toxicol Pharm.* 13: 57-149
- Salanki J. (1986) Biological monitoring of the state of the environment: Bioindicators I.U.B.S. Monograph series No. 1 for I.C.S.U. by I.R.L.
- Salaskar G.M. and Nayak V.N. (2011) Nutritional quality of bivalve, *Crassostrea madrasensis* and *Perna viridis* in the Kali estuary, Karnataka, India. *Recent Research in Science and Technology.* 3(4): 6-11.
- Salchi P., Asghari B. and Mohammadi F. (2008) Removal of heavy metals from aqueous solutions by *Cercis siliquastrum* L. *J. Iran. Chem. Soc.* 5: 580-586.
- Schuliga M., Chouchane S. and Snow E.T. (2002) Up regulation of glutathione related genes and enzyme activities in cultured human cells by sub lethal concentration of inorganic arsenic. *Toxicol. Sci.* 70: 183-192.

- Schwarzenbach R.P., Egli T., Hofstetter T.B., von Gunten U. and Wehrli B. (2010) Global water pollution and human health. *Annual Review of Environment and Resources*. 35: 109-136.
- Shariati S.R.P., Bonakdarpour B., Zare N. and Ashtiani F.Z. (2011) The effect of hydraulic retention time on the performance and fouling characteristics of membrane sequencing batch reactors used for the treatment of synthetic petroleum refinery wastewater. *Bioresour. Technol.* 102(17): 7692-7699.
- Sharma D. and Jain R. (2000): Physico Chemical Analysis of Gopalpura Tank of Guna District (M.P.). *Ecol. Env. and con.* 6(4): 441-445.
- Sharma S. (2003) Biodiversity of littoral benthic organism & their tropical relationship with shorebirds & fishes in Krishnpura lake Indore M.P., D.A.V.V.
- Sharma S., Joshi V., Kurde S. and Sighavi M. (2007) Bio-diversity of benthic macro-invertebrates and fish species communities of krishnapura lake, Indore, M.P. *Aqua Bio.* 22(1):1-4.
- Sheehan D., McIntosh J., Power A. and Fitzpatrick P.J. (1995) Drug metabolizing enzymes of mussels as bioindicators of chemical pollution. *Biochem.Soc. Trans.* 23: 419-422.
- Shinde S. Waykar B. and Deshmukh G.M. (2011) Bioaccumulation of metal in fresh water pelecypod molluscs under experimental condition. *The Bioscan.* 6(4): 537-542.
- Sidoumou Y. Romeo M., Siau Z. and Gnasia M.B. (1999) Heavy Metal Distribution in different Fish Species from the Mauritania Coast. *Science of the Total Environment.* 232(3): 169-175.

- Sies H. (1999) Glutathione and its role in cellular functions. *Free Radic. Biol. Med.* 27: 916-921.
- Singh A.N., Shukla A.K., Jagannadham M.V. and Dubey V.K. (2010) Purification of a novel cysteine protease, procerain B, from *Calotropisprocera* with distinct characteristics compared to procerain. *Process Biochemistry.* 45: 399-406.
- Singh K.P., Malik A., Mohan D. and Sinha S. (2004) Multivariate statistical techniques for the evaluation of spatial and temporal variations in water quality of Gomti River (India) a case study. *Wat Res.* 38: 3980-3992.
- Singh M., Muller G. and Singh I.B. (2003) Geogenic distribution and baseline concentration of heavy metals in sediments of the Ganges River, India. *J Geochem Exp.* 80: 1-17.
- Singh S. P., Pathak D. and Singh R. (2002) Hydrobiological studies of two ponds of satna (M.P.), India, *Eco. Evn.and Cons.* 8(3): 289-292.
- Sokolova I.M., Ringwood A.H. and Johnson C. (2005) Tissue-specific accumulation of cadmium in subcellular compartments of eastern oysters *Crassostrea virginica* Gmelin (*Bivalvia. Ostreidae*). *Aquat Toxicol.* 74: 218-228.
- Solanki J., Farkas A., Kamardina T. and Rozsa K.S. (2003) Molluscs in biological monitoring of water quality. *Toxicol Lett.* 141: 403-410.
- Subba R.C. (1995) Ground Water Quality in Residential Colony. *Ind.J. environ. Health.* 37(4): 295-300.
- Subba Rao D. and Govind B.V. (1967) Hydrology of Tugab'hadra reservoir. *Indian Journal of Fisheries.* 321-344.

- Sunny A., Vattakeril and Diwan A.P. (1991) Community structure of benthic macro-invertebrates & their utility as indicators of pollution in river Kshipra, India. *J. Pollution Research*. 10: 1-11.
- Szefer P., Ali A.A., Ba-Haroon A.A., Rajeh A.A., Geldon J. and Nabrzyski M. (1999) Distribution and relationships of selected trace metals in molluscs and associated sediments from the Gulf of Aden, Yemen. *Environ Pollut*. 106: 299-314.
- Thorburn P.J., Cook F.J. and Bristow K.L. (2003a) Soil-dependent wetting from trickle emitters: implications for system design and management. *Irrig. Sci*. 22: 121-127.
- Thorburn P.J., Dart I.K., Biggs I.M., Baillie C.P., Smith M.A. and Keating B.A. (2003b) The fate of nitrogen applied to sugarcane by trickle irrigation. *Irrig Sci*. 22: 201-209.
- Thorne R.S. and William W.P. (1997) The response of benthic macroinvertebrates to pollution in developing countries. A multimetric system of bioassessment. *Freshw. Biol*. 37: 671-686.
- Tsukamoto H. (2002): Redox regulation of cytokine expression in Kupffer cells. *Antiox Redox Signal*. 4: 741-748.
- Umminger B.L. (1977) Relation of whole blood sugar concentration in vertebrates to standard metabolic rates. *Comparative Biochemistry and Physiology*. 55: 457-460.
- Usero J., Izquierdo C., Morillo J. and Gracia I. (2003) Heavy metals in fish (*Solea vulgaris*, *Anguilla anguilla* and *Liza aurata*) from salt marshes on the southern Atlantic coast of Spain. *Environ Int*. 29: 949-956.

- Usero J., Marilla J. and Graccia I. (2005) Heavy metal concentrations in mollusc from the Atlantic Coast of Sothern Spain. *Chemosphere*. 59: 1175-1181.
- Valavanidis A., Vlahogianni T., Dassenakis M. and Scoullou M. (2006) Molecular biomarkers of oxidative stress in aquatic organisms in relation to toxic environmental pollutants. *Ecotoxicol. Environ. Safe*. 46: 178-189.
- van Vuren J.H.J., Du Preez H.H. and Deacon A.R. (1994) Effect of Pollutants on the Physiology of Fish in the Olifants River (Eastern Transvaal). WRC Report No 350/1/94. Water Research Commission, Pretoria. pp. 214.
- Vasseur P. and Cossu-Leguille C. (2003) Biomarkers and community indices as complementary tools for environmental safety. *Environment International*. 28: 711-717.
- Venkatesharaju K., Ravikumar P., Somashekar R.K. and Prakash K.L. (2010) Physico-chemical and Bacteriological Investigation on the river Cauvery of Kollegal Stretch in Karnataka, *Journal of science Engineering and technology*. 6(1): 50-59.
- Ventura-Lima J, Sandrini J.Z., Ferreira-Cravo M., Piedras F.R., Moraes T.B., Fattorini D., Notti A., Regoli F., Geracitano L.A., Marins L.F. and Monserrat J.M. (2007) Toxicological responses in *Laeonereis acuta* (Annelida, Polychaeta) after As exposure. *Environ. Intern*. 33:559-564.
- Verkleji J.A.S. (1993) The effects of heavy metals stress on higher plants and their use as bio monitors. In: Markert B (ed) *Plant as bioindicators: indicators of heavy metals in the terrestrial environment*. VCH, New York. Pp. 415-424.

- Verlecar X.N., Jena K.B., Chainy G.B.N. (2007) Biochemical markers of oxidative stress in *Perna viridis* exposed to mercury and temperature, *Chemico. Biological Interactions*. 167: 219-226.
- Vincent S., Ambhore T., Kumar L.C.A. and Selvanayagam M. (1995) Biochemical response of the Indian major carp, *Catlacatla* (Ham) to Chromium toxicity. *Indian J. Environ. Health*. 37(3): 190-196.
- Wagner A., Boman J. (2003) Biomonitoring of trace elements in muscle and liver tissue of freshwater fish. *Spectrochim Acta*. 58: 2215-2226.
- Waykar B and Shinde S (2011) Assesment of the heavy metal of bioaccumulation in three species fresh water bivalves. *Bull Env. Contam. Toxicol*. 87(3):267-271.
- Waykar B., Lomte V.S. and Zambare S.P. (2001) Effect of cypermethrin on ascorbic acid content in the mantle, foot, gill, digestive gland and whole body tissues of freshwater bivalve, *Unio.sp.* *J. Aqua. Biol*. 16: 57-61.
- Wetzel R. G. (1975) *Limnology*. Philadelphia: Saunders, (1975) 1983. Freshwater Ecology Program, Dept. Biological Sciences. Univ. Alabama, Tuscaloosa, AL. pp 860.
- Wildi W. (2004) *Research and Management*. pp. 75.
- Zhou Q., Zhang J., Fu J., Shi J. and Jiang G. (2008) Biomonitoring: An appealing tool for assessment of metal pollution in the aquatic ecosystem. *Anal.Chim.Acta*. 606: 135-150.
- Zhoua Q., Zhanga J., Fua J., Shia J. and Jiang G. (2008) Biomonitoring: An appealing tool for assessment of metal pollution in the aquatic ecosystem. *Ana Chim Acta*. 606: 135-150.

Zhoua Q., Zhanga J., Fua J., Shia J. and Jiang G. (2008) Biomonitoring: An appealing tool for assessment of metal pollution in the aquatic ecosystem. *Ana Chim Acta*. 606: 135-150.

Zhoua Q., Zhanga J., Fua J., Shia J. and Jiang G. (2008) Biomonitoring: An appealing tool for assessment of metal pollution in the aquatic ecosystem. *Ana Chim Acta*. 606: 135-150.

# *Appendices*